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Search Notes		
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JAN 125480

SEARCH REQUEST FORM

Scientific and Technical Information Center

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=> fil wpix FILE 'WPIX' ENTERED AT 10:40:01 ON 07 JUL 2004 COPYRIGHT (C) 2004 THOMSON DERWENT FILE LAST UPDATED: 2 JUL 2004 <20040702/UP> MOST RECENT DERWENT UPDATE: 200442 <200442/DW> DERWENT WORLD PATENTS INDEX SUBSCRIBER FILE, COVERS 1963 TO DATE >>> FOR A COPY OF THE DERWENT WORLD PATENTS INDEX STN USER GUIDE, PLEASE VISIT: http://www.stn-international.de/training center/patents/stn guide.pdf <<< >>> FOR DETAILS OF THE PATENTS COVERED IN CURRENT UPDATES, SEE http://thomsonderwent.com/coverage/latestupdates/ <<< >>> FOR INFORMATION ON ALL DERWENT WORLD PATENTS INDEX USER GUIDES, PLEASE VISIT: http://thomsonderwent.com/support/userguides/ <<< >>> NEW! FAST-ALERTING ACCESS TO NEWLY-PUBLISHED PATENT DOCUMENTATION NOW AVAILABLE IN DERWENT WORLD PATENTS INDEX FIRST VIEW - FILE WPIFV. FREE CONNECT HOUR UNTIL 1 MAY 2004. FOR FURTHER DETAILS: http://www.thomsonderwent.com/dwpifv >>> NEW! IMPROVE YOUR LITIGATION CHECKING AND INFRINGEMENT MONITORING WITH LITALERT. FIRST ACCESS TO RECORDS OF IP LAWSUITS FILED IN THE 94 US DISTRICT COURTS SINCE 1973. FOR FURTHER DETAILS: http://www.thomsonscientific.com/litalert >>> THE DISPLAY LAYOUT HAS BEEN CHANGED TO ACCOMODATE THE NEW FORMAT GERMAN PATENT APPLICATION AND PUBLICATION NUMBERS. SEE ALSO: http://www.stn-international.de/archive/stnews/news0104.pdf <<< => d all abeq tech abex tot L62 ANSWER 1 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2004-203810 [19] AN WPIX DNN N2004-161906 DNC C2004-080473 ΤI Determination of concentration of free thrombin in e.g. blood, involves measuring physical property of the product as function of time, and determining time derivative of the concentration of alpha-macroglobulin-thrombin complex. DC B04 D16 S03 IN HANSSON, G PA (ASTR) ASTRAZENECA AB CYC 105 PΙ WO 2004016807 A1 20040226 (200419) * EN 32 C12Q001-56 RW: AT BE BG CH CY CZ DE DK EA EE ES FI FR GB GH GM GR HU IE IT KE LS LU MC MW MZ NL OA PT RO SD SE SI SK SL SZ TR TZ UG ZM ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NI NO NZ OM PG PH PL PT RO RU SC SD SE SG SK SL SY TJ TM TN TR TT TZ UA UG US UZ VC VN YU ZA ZM ZW ADT WO 2004016807 A1 WO 2003-SE1273 20030813 PRAI SE 2002-2454 20020816 IC ICM C12Q001-56

NOVELTY - A concentration of free thrombin in sample is

ICS G01N033-49; G01N033-86

WO2004016807 A UPAB: 20040318

ICA A61P007-02

AB

determined as a function of time by measuring a physical property of the product as a function of time, computing a time derivative of the physical property to determine a rate of reaction from the substrate to product, and determining the concentration of free thrombin, from a time derivative of the concentration of alpha-macroglobulin-thrombin complex (alpha 2MT).

DETAILED DESCRIPTION - Determination of concentration of free thrombin in a sample as a function of time comprises providing a substrate that reacts with thrombin to form a product, measuring a physical property of the product as a function of time, computing a time derivative of the physical property to determine a rate of reaction from the substrate to product, deterring the concentration of alpha 2MT as a function of time, and determining the concentration of free thrombin , from a time derivative of the concentration of alpha 2MT.

INDEPENDENT CLAIMS are also included for:

- (1) an apparatus for determining a concentration of free thrombin in a sample as a function of time comprising physical property measuring element, time derivative computing device, and free thrombin determining device. used in the invention; and
- (2) assessing the effectiveness of putative anticoaqulant, comprising providing a sample containing a thrombin precursor, determining the concentration of free thrombin in the sample as a function of time using the method, and assessing whether the rate of free thrombin formation determined is indicative of anticoagulant activity.

USE - For determining a concentration of free thrombin in a sample, e.g. blood and/or blood plasmas.

ADVANTAGE - The inventive method provides more accurate determination of free thrombin concentration in the sample.

DESCRIPTION OF DRAWING(S) - The drawing shows a schematic diagram of the process for thrombin induced conversion of a substrate to product yielding a measurable change in optical properties of the product. Dwq.1/3

FS CPI EPI

FΑ AB; GI

MC CPI: B04-B04D4; B04-B04D5; B04-H19;

B11-C08E; B12-K04; D05-H08; D05-H09; D05-H13

EPI: S03-E14H; S03-E14H1

TECH UPTX: 20040318

TECHNOLOGY FOCUS - INSTRUMENTATION AND TESTING - Preferred Method : The method further comprises iteratively varying values for parameter k, expressing the relationship between the rate of change of concentration of alpha2MT and a concentration of free thrombin, or parameter, kcat2, expressing a rate of turnover of substrate by alpha2MT, to reduce an optimization value. The determining of the concentration of alpha2MT includes assuming the initial condition of t = 0 of (alpha2MT) = 0. The method also includes predetermining a rate of turnover of substrate from thrombin, kcat2, and predetermining the Michaelis constant, k-M for the substrate, thrombin, and alpha2MT. Preferred Parameters: The optimization value incorporates a value, RMSE, expressing a difference between a measured value of the physical property of the product and a calculated combined value related to the concentrations of free thrombin and alpha2MT. The optimization value further comprises a factor related to the calculated average free thrombin concentration during a preceding period of time. The value RMSE comprises the mathematical equation (I). (Structure (I), page 20) P = concentration of product at time; al = reaction rate from substrate to product by thrombin;

kcat1 = rate of turnover of substrate by thrombin;

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and
     kcat2 = rate of turnover of substrate by alpha-MT.
L62 ANSWER 2 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     2003-723490 [69]
ΔN
                        WPTX
DNN N2003-578477
                        DNC C2003-199290
     Test strip qualification method for measuring
ТT
     prothrombin time with whole blood by obtaining
     prothrombin time results, and comparing result
     from two control areas to two control qualification criteria.
DC
     B04 S03
IN
     PATEL, H; PATEL, H I
     (LIFE-N) LIFESCAN INC; (PATE-I) PATEL H
PΑ
CYC 38
    ·EP 1345030
PΤ
                     A1 20030917 (200369) * EN
                                                11
                                                      G01N033-49
         R: AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LI LT LU LV
            MC MK NL PT RO SE SI SK TR
                     A 20030915 (200369)
     NO 2003000903
                                                      G01N033-52
                     A1 20030914 (200372)
     CA 2421467
                                                      G01N033-86
                                           EN
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     JP 2003294759
                     A 20031015 (200376)
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                                                      G01N033-86
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                     A 20031001 (200382)
     CN 1445550
                                                      G01N033-86
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                     A1 20030918 (200382)
     US 2003175978
                                                      G01N031-00
     KR 2003074409
                     A 20030919 (200410)
                                                      G01N033-52
                     B2 20040106 (200411)
     US 6673617
                                                      G01N031-00
     AU 2003200892
                     A1 20031002 (200428)
                                                      G01N033-86
     US 2004096980
                    A1 20040520 (200434)
                                                      G01N021-00
ADT EP 1345030 A1 EP 2003-251534 20030313; NO 2003000903 A NO 2003-903
     20030226; CA 2421467 A1 CA 2003-2421467 20030310; JP 2003294759 A JP
     2003-68305 20030313; CN 1445550 A CN 2003-120515 20030313; US 2003175978
     A1 US 2002-100531 20020314; KR 2003074409 A KR 2003-15615 20030313; US
     6673617 B2 US 2002-100531 20020314; AU 2003200892 A1 AU 2003-200892
     20030228; US 2004096980 Al Cont of US 2002-100531 20020314, US 2003-712679
     20031112
FDT US 2004096980 A1 Cont of US 6673617
PRAI US 2002-100531
                          20020314; US 2003-712679
IC
     ICM G01N021-00; G01N031-00; G01N033-52; G01N033-86
     ICS A61B005-000; C12Q001-56; G06F015-04
ICA
    G01N033-49
          1345030 A UPAB: 20031027
AB
     EΡ
     NOVELTY - Test strip qualification method comprises:
          (1) providing a test strip (2);
          (2) obtaining prothrombin time (PT) results for
     each reaction area;
          (3) comparing result from first and second control areas to
     first and second control qualification criteria, respectively; and
          (4) outputting message to user indicating test strip reliability.
          DETAILED DESCRIPTION - Test strip qualification method
     comprises:
          (1) providing a test strip;
          (2) obtaining prothrombin time (PT) results for
     each reaction area;
          (3) comparing result from the first and second control
     areas to first and second control qualification criteria, respectively;
     and
          (4) outputting a message to user indicating test strip reliability.
          The test strip comprises assay reaction area, and two control
     reaction areas. The first control qualification criteria comprise upper
     and lower limits. The first control upper limit is at least partially
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dependent upon assay reaction area PT result.

and

INDEPENDENT CLAIMS are also included for:

(1) system programmed to operate according to the method;

(2) computer-readable medium using a program to direct system to perform the method. USE - The method is useful in measuring prothrombin time (PT) (claimed) with whole blood. ADVANTAGE - The method provides test strip that gives highly accurate and reliable results. DESCRIPTION OF DRAWING(S) - The figure shows a top view of the test strip. Test strip 2 Measurement areas 4, 6, 8 Introduction port 10 Bladder 12 Channel 14 Junction 16 Bypass channel 18 Dwg.1a/5 FS CPI EPI AB; GI; DCN FA MC CPI: B04-B04D5; B04-H19; B11-C08; B12-K04 EPI: S03-E14H; S03-E14H1 TECH UPTX: 20031027 TECHNOLOGY FOCUS - MECHANICAL ENGINEERING - Preferred Components: The upper limit is dependent upon assay reaction area PT results at most2 international normalized ratio (INR). The upper limit comprises linear function dependent upon assay reaction area PT results. The upper limit also comprises a value independent of assay reaction area PT result for reaction area PT result at least2 INR. The lower limit comprises value independent of assay reaction PT result. The PT results obtained for each reaction area are INR values. The second control qualification criteria comprise upper and lower limits. The lower limit has two sections dependent upon assay reaction area PT results. The second control drops-off from the first section. L62 ANSWER 3 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2002-489927 [52] AN WPIX DNN N2002-387327 DNC C2002-139068 ΤI Novel reagent useful for assessment of hemostatic potential of blood or plasma sample, comprises a coagulation activator. DC B04 D16 P31 BAGLIN, T; DOOBAY, H; FISCHER, T J; LUDDINGTON, R; IN TEJIDOR, L PA (ALKU) AKZO NOBEL NV; (BIOM-N) BIO MERIEUX INC; (INMR) BIOMERIEUX INC CYC PΙ WO 2002034109 A2 20020502 (200252)* EN 44 A61B000-00 RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PH PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW AU 2002015382 A 20020506 (200257) A61B000-00 <--EP 1337858 A2 20030827 (200357) EN G01N033-86 < - -R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI TR US 6645768 B1 20031111 (200382) G01N033-86 < - ~ BR 2001014942 A 20031223 (200406) G01N033-86 <--W 20040422 (200428) JP 2004512511 70 G01N033-48 <---ADT WO 2002034109 A2 WO 2001-US32563 20011018; AU 2002015382 A AU 2002-15382 20011018; EP 1337858 A2 EP 2001-983998 20011018, WO 2001-US32563 20011018; US 6645768 B1 US 2000-698589 20001027; BR 2001014942 A BR 2001-14942 20011018, WO 2001-US32563 20011018; JP 2004512511 W WO 2001-US32563

20011018, JP 2002-537169 20011018 FDT AU 2002015382 A Based on WO 2002034109; EP 1337858 A2 Based on WO 2002034109; BR 2001014942 A Based on WO 2002034109; JP 2004512511 W Based on WO 2002034109 PRAI US 2000-698589 20001027 ICM A61B000-00; G01N033-48; G01N033-86 ICS C07K004-00; C12Q001-56; G01N035-02 WO 200234109 A UPAB: 20020815 AB NOVELTY - A reagent (I) comprising a coagulation activator at a concentration of 11 picomolar or less, for assessment of the hemostatic potential of a blood or plasma sample, is new. DETAILED DESCRIPTION - An INDEPENDENT CLAIM is also included for a kit for assessing the hemostatic potential of a test sample comprising a coagulation activator at a concentration of 11 picomolar or less, or the activator and instructions for diluting the activator, vesicles, a metal divalent cation or a metal salt capable of dissociating into a metal divalent cation, instructions for adding the activator , metal cation or metal salt and vesicles to a test sample, and instructions for assessing the hemostatic potential of the test sample. USE - The reagent and the kit are useful for indicating a sample to be hypocoagulable, normal or hypercoagulable , depending upon the condition of the patient from which the sample was taken, for indicating a patient to have thrombotic tendency, hemorraghic tendency, or stasis, and also for assessing hemostatic potential of a blood or plasma sample (claimed). (I) is useful in the drug discovery and drug development processes by modifying the components or concentrations of the reagent. (I) is useful to determine the amount of plasma to be modified in order to restore coagulability to normal. ADVANTAGE - The reagent allows for globally assessing both the hypercoagulable potential and hypocoagulable potential of a patient in a single assay, which is accurate, sensitive and easy. The test is simple and can be automated on standard laboratory coagulometers. The test is based on the rate of fibrin polymerization which allows detection of perturbances in the propagation, amplification and polymerization pathways, whereas in the traditional prothrombin time test, these parts of the coagulation pathway are overshadowed by the excessive amounts of Factor IIa produced by the initiation phase. Dwg.0/10 CPI GMPI FS AB; DCN FΑ MC CPI: B04-B04D4; B04-B04D5; B04-H19; B04-N02; B05-A01B; B05-A03A; B05-B01P; B05-C07; B11-C08E; B12-K04A; D05-H09 TECH UPTX: 20020815 TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Reagent: (I) further comprises vesicles or liposomes. The vesicles comprise platelets, cellular debris, phospholipid vesicles (prepared by dilution, sonication, dialysis or extrusion), or platelet microparticles. The coagulation activator comprises tissue factor which is a recombinant or purified, truncated tissue factor, or cells expressing tissue factor on their surface. The tissue factor comprises a metal cation, especially a divalent metal cation such as magnesium, calcium or manganese or metal salt (5-50, preferably 15-35 mM), preferably a halide of magnesium, calcium or manganese, which dissociates into a metal cation. The tissue factor is at a

concentration of 11, 8 or 6 picomolars, preferably 3 picomolars or less. The vesicles comprise phospholipids (at a concentration of 10-300 micromolar, preferably 50-200 micromolar) which comprise one or more of phosphatidylcholine, phosphatidylethanolamine or phosphatidylserine at a ratio of 0-10, preferably 10 %, by mole phosphatidylserine, 5-30, preferably 20 %, by mole phosphatidylethanolamine and the remainder, preferably 70 %, by mole phosphatidylcholine. The coagulation activator comprises tissue factor-rich mammalian tissue extracts, tissue factor purified from mammalian tissue or thromboplastin. The coagulation activator is capable of detecting defects in the initiation phase. (I) further comprises an activator of an anticoagulant pathway, preferably an activator of protein C which is a purified human or non-human mammalian thrombomodulin, soluble or membrane associated thrombomodulin, native thrombomodulin or thrombomodulin reconstituted with phospholipids, partially or fully glycosylated thrombomodulin, and fully deglycosylated thrombomodulin. The protein C activator (thrombomodulin) is at a concentration of 30 nanomolar or less, preferably 5-20 nanomolar. The thrombomodulin comprises heparin or heparin-like molecules and is relipidated with phospholipids comprising 10 % phosphatidylethanolamine. (I) further comprises buffers and/or stabilizers, or phospholipids. Preferred Kit: The kit further comprises calcium cation or calcium salt that dissociates into a calcium cation, and an activator of an anticoagulant pathway and instruction for adding the activator to the test sample. The thrombomodulin is provided separately from the coagulation activator, and mixed with heparin, heparin sulfate or heparin-like molecules. The kit has a first container having the coagulation activator which is a tissue factor at a concentration of 11 picomolars or less mixed with vesicles which are phospholipids at a concentration of 10-300 picomolar, a second container having a metal salt at a concentration of 5-50 mM, and third container having the coagulation activator mixed with vesicles and an activator of an anticoagulant pathway which is thrombomodulin at a concentration of 300 nanomolar or less.

ABEX

UPTX: 20020815

EXAMPLE - An assay was conducted for detecting the coagulability , by adding 50 micro-l of plasma to 50 micro-l of the activator and 50 micro-l of the start reagent which consisted of 0.25 M calcium chloride. A normal sample, a hypocoagulable sample (factor VIII deficient plasma) and a hypercoagulable plasma (protein S deficient plasma) were evaluated at various dilutions of the activator. The activator was diluted with a buffer at two dilutions, 1:100 and 1:50000 of its original concentration. The assay was conducted at 37 degrees C, and the reaction was monitored at 580 nm for 300 seconds. Endpoints were calculated for time and rate indices of clot formation. The ratio of the endpoint of reagent dilution (x) for specimen/endpoint of reagent dilution (y) for specimen to the endpoint of reagent dilution (x) for npp/endpoint of reagent dilution (y) for npp was calculated, where x is 1:100 dilution and y is a series of dilutions. The results were expressed as the magnitude of deviation at a given dilution or as the dilution required to deviate from ideal (normal value or normal range). As the dilution of the reagent was greater (y became larger) the results for the two abnormal plasmas (the hypercoagulable and hypocoagulable plasmas) tested began to deviate from the calculated endpoints or ratios

of the normal plasma. The hypocoagulable specimen produced ratios that were greater than 1 and the hypercoagulable specimen had ratios that were less than 1 for the endpoint (clot time)/ratio combination. L62 ANSWER 4 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2002-463332 [49] WPIX DNN N2002-365287 DNC C2002-131735 Determining hyper- or hypocoagulable condition of a patient, comprises initiating coagulation in patient sample by fibrin polymerization activator and monitoring formation of fibrin polymer to drive time dependent profile. B04 D16 P31 BAGLIN, T; FISCHER, T J; TEJIDOR, L (ALKU) AKZO NOBEL NV; (INMR) BIOMERIEUX INC; (BAGL-I) BAGLIN T; (FISC-I) FISCHER T J; (TEJI-I) TEJIDOR L A2 20020502 (200249)* EN WO 2002034110 68 A61B000-00 RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PH PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW AU 2002014619 A 20020506 (200257) C12Q001-56 <--EP 1337660 A2 20030827 (200357) EN <--C12Q001-56 R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI TR A 20031007 (200373) BR 2001014936 C12Q001-00 <--A1 20040318 (200421) <--US 2004053351 C12Q001-56 US 6743596 B1 20040601 (200436) <--C12Q001-56 JP 2004518111 W 20040617 (200440) 106 G01N033-86 <---ADT WO 2002034110 A2 WO 2001-US32564 20011018; AU 2002014619 A AU 2002-14619 20011018; EP 1337660 A2 EP 2001-983170 20011018, WO 2001-US32564 20011018; BR 2001014936 A BR 2001-14936 20011018, WO 2001-US32564 20011018; US 2004053351 Al Div ex US 2000-697934 20001027, US 2003-663449 20030916; US 6743596 B1 US 2000-697934 20001027; JP 2004518111 W WO 2001-US32564 20011018, JP 2002-537170 20011018 FDT AU 2002014619 A Based on WO 2002034110; EP 1337660 A2 Based on WO 2002034110; BR 2001014936 A Based on WO 2002034110; JP 2004518111 W Based on WO 2002034110 PRAI US 2000-697934 20,001027; US 2003-663449 20030916 ICM A61B000-00; C12Q001-00; C12Q001-56; G01N033-86 ICS G01N021-75; G01N033-15; G01N033-49; G01N033-50

AB WO 200234110 A UPAB: 20020802

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NOVELTY - Determining (M1) if a patient is hypercoagulable, hypocoagulable or normal, comprises initiating coagulation in the test sample of patient in the presence of an activator for carrying out intrinsic tenase-dependent fibrin polymerization (IP), and monitoring formation of IP over time to drive a time-dependent profile, where the results determine whether the patient is hyper- or hypocoagulable, or normal, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

- (1) assessing the coagulation system in a test sample;
- (2) detecting defects in the propagation and/or amplification phase

in the coagulation system of a test sample;

- (3) monitoring an antithrombotic or procoagulant pharmaceutical therapy;
- (4) evaluating the efficacy of an antithrombotic or procoagulant pharmaceutical; and
 - (5) assessing the hemostatic potential of a sample.

ACTIVITY - Thrombolytic; Anticoagulant;

Coagulant. No supporting data is given in the source material.

MECHANISM OF ACTION - None given in the source material.

USE - For determining if a patient is hypercoagulable, hypocoagulable or normal, for assessing the coagulation system in a test sample, monitoring an antithrombotic or procoagulant pharmaceutical therapy, evaluating the efficacy of an antithrombotic or procoagulant pharmaceutical and assessing hemostatic potential of a sample (claimed). The method is useful for assessing the hemostatic potential of a sample. The method is also useful for determining how much the plasma needs to be modified in order to restore coagulability to normal.

ADVANTAGE - The method allows for globally assessing both the hypercoagulable potential and hypocoagulable potential of a patient in a single assay. The method is accurate and easy to use. Disturbances in the propagation and amplification loops are accessible in this method, whereas in the traditional prothrombin (PT) test, the parts of the coagulation pathway are overshadowed by the excessive amounts of Factor IIa produced by the initiation phase.

Dwg.0/10

FS CPI GMPI

FA AB; DCN

MC CPI: B04-B04D; B04-H19; B11-C07B2; B12-K04A2

; B14-F04; B14-F08; D05-H09

TECH

UPTX: 20020802

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: The time-dependent profile, at least part of which includes initiation of clot formation, overall change in profile, slope of profile after initiation of clot formation and/or acceleration at the time of clot initiation, is compared to a time-dependent profile of a known sample. At least two time-dependent fibrin polymerization profiles are obtained, an additional profile obtained for a known sample from computer memory or by adding the activator at least one concentration to a known sample and monitoring the formation of fibrin polymerization over time. At least two time -dependent fibrin polymerization profiles are obtained for two different activator concentrations, and/or one or more profiles for a known sample at one or more activator concentrations. Parameter from each time-dependent fibrin polymerization profile having varying activator concentrations is determined and a concentration at which at least one parameter of the sample being tested deviates from normal is determined. The parameter is the index and value of the minimum of first derivative, the time index and value for the minimum and maximum of the second derivative or overall magnitude of change. The part is rate of acceleration of fibrin polymerization compared to known sample. A difference or ratio of parameters for test sample and normal sample are determined. Parameter is clot time and a ratio of clot times at different activator concentrations is determined. The parameter includes the time of initiation of clot formation,

rate of clot formation, maximum acceleration of clot formation, turbidity at a predetermined time period or total change in turbidity. The parameters are measures of defects in the thrombin propagation and/or amplification phases. A ratio of one parameter in test and normal sample, and ratio for multiple concentrations of activator, are determined. A concentration at which ratio departs from 1 is determined. An activator of one or more anticoagulant pathways, and an activator of protein C e.g. thrombomodulin or its derivatives given in the specification are added. A fibrin polymerization profile is obtained with and without thrombomodulin. The activator comprises tissue factor and phospholipids. A metal salt (e.g. halide of magnesium, calcium or manganese) which dissociates into a metal divalent cation when added to the test sample, is added as part of the activator. The activator comprises homogenized cerebral tissue. M1 further involves adding phopholipids together with or separately from the activator, adding buffers and/or stabilizers to the test sample e.g. patient plasma sample. The time dependent measurement profile is an optical absorbance or transmittance profile provided on an automated analyzer. A visible light beam is directed through a container holding the test sample and activator and light absorbed or transmitted is monitored to form the time dependent measurement profile. The activator comprises recombinant or purified tissue factor, truncated tissue factor or cells expressing tissue factor on their surface, sufficiently diluted to determine hypercoagulable, normal or hypocoagulable depending upon the condition of the patient. Defects in formation of intrinsic tenase complex are detected. One or more endpoints from the time-dependent measurement profile are calculated, the endpoints selected from the time of clot initiation and the rate of polymerization. Sample is whole blood or platelet rich plasma. M1 further involves adding vesicles (e.g. platelets, cellular debris, phospholipid vesicles or platelet microparticles) to the test sample. M1 further involves adding less than 11 pM concentration of tissue factor that generates intrinsic dependent fibrin polymerization in the patient sample, measuring formation of fibrin polymerization, and determining whether the patient is hypercoagulable, normal or hypocoagulable, based on the measured fibrin polymerization. Fibrin polymerization profile is obtained at multiple concentrations of activator which triggers thrombin explosion. The fibrin polymerization measurement is used to adjust the patient's therapy to result in a fibrin polymerization profile approximating normal. UPTX: 20020802

EXAMPLE - The assay was conducted by adding 50 micro liter of plasma to 50 micro liter of the activator and then adding 50 micro liter of the start reagent. A normal sample, a hypocoagulable sample (Factor VIII deficient plasma) and a hypercoagulable plasma (protein S deficient plasma) were evaluated at various dilutions of the activator. The activator was a commercially available thromboplastin diluted with a buffer at two dilutions, a 1:100 and 1:500000 of its original concentration. The start reagent consisted of 0.25 M Calcium chloride. The assay was conducted at 37 degrees C and the

0.25 M Calcium chloride. The assay was conducted at 37 degrees C and the reaction was monitored at 580 nm for 300 seconds. Endpoints were calculated for time and rate indices of clot

formation. Ratios of endpoints were compared to other

AREX

dilutions and other samples. As the dilution of the reagent become greater, the results for the two abnormal plasmas (hypercoagulable and hypocoagulable plasmas) tested began to deviate from the calculated endpoints or ratios of the normal plasma. The results were expressed as the magnitude of deviation at a given dilution or as the dilution required to deviate from ideal (normal value or normal range). The hypercoagulable and hypocoagulable results deviating in opposite directions indicating the ability to differentiate between the two conditions, were shown graphically. L62 ANSWER 5 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN AN 2002-268645 [31] WPTX DNN N2002-209092 DNC C2002-079631 Assaying activated protein C in TТ blood, for screening drugs, comprises contacting plasma with blood coagulation cascade activator and assaying activity of activated protein C. DC B04 S03 HOSAKA, Y; IMADA, K; OHMORI, Y; SHIRAKAWA, K; TAKAHASHI, Y IN PA (MOCH) MOCHIDA PHARM CO LTD CYC 96 A1 20011227 (200231)* JA PΙ WO 2001098782 49 G01N033-53 RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW AU 2001074597 A 20020102 (200233) G01N033-53 JP 2002504491 Х 20030916 (200362) G01N033-53 <--ADT WO 2001098782 A1 WO 2001-JP5376 20010622; AU 2001074597 A AU 2001-74597 20010622; JP 2002504491 X WO 2001-JP5376 20010622, JP 2002-504491 20010622 AU 2001074597 A Based on WO 2001098782; JP 2002504491 X Based on WO FDT2001098782 PRAI JP 2000-227966 20000622 IC ICM G01N033-53 C12Q001-56; G01N033-15; G01N033-50; G01N033-86 ICA C12N009-99 AB WO 200198782 A UPAB: 20020516 NOVELTY - Assaying the amount of activated protein C in a blood sample comprising contacting plasma in a sample with a substance activating a blood coagulation cascade, and assaying the activity of an activated protein C and/or the amount of an antigen in a protein C activated protein, is DETAILED DESCRIPTION - An INDEPENDENT CLAIM is also included for a kit for conducting the above assay. USE - The method is useful for assaying the amount of activated protein C in a blood sample (claimed) which can be used to screen drugs. ADVANTAGE - Allows an assay of protein C to be conducted under conditions similar to physiological conditions. Dwg.0/7 FS CPI EPI FA AB; DCN MC CPI: B04-B04D2; B04-C01; B04-H19; B04-N02; B11-C08E; B11-C10; B12-K04E; B14-L01; B14-L06 EPI: S03-E14H4 TECH UPTX: 20020516 TECHNOLOGY FOCUS - ORGANIC CHEMISTRY - Preferred Method: The

plasma in the blood sample is firstly contacted with a drug. The substance activating the blood coagulation cascade is thromboplastin. Thrombin activity inhibitor may be added just before the assaying step..

- L62 ANSWER 6 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
- AN 2002-122222 [16] WPIX
- DNN N2002-091675 DNC C2002-037464
- TI Detection of a complex of lipoprotein and an acute phase protein useful for predicting an increased probability of system failure or mortality involves adding a reagent to a sample, and measuring the formed complex over time.
- DC B04 S03
- IN DOWNEY, C; FISCHER, T J; NESHEIM, M; SAMIS, J A; TEJIDOR, L; TOH, C H; WALKER, J B
- PA (ALKU) AKZO NOBEL NV; (INMR) BIOMERIEUX
 INC; (DOWN-I) DOWNEY C; (FISC-I) FISCHER T J; (NESH-I) NESHEIM M;
 (SAMI-I) SAMIS J A; (TEJI-I) TEJIDOR L; (TOHC-I) TOH C H; (WALK-I) WALKER J B
- CYC 97
- PI WO 2001096864 A2 20011220 (200216) * EN 83 G01N033-49 <-RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ
 NL OA PT SD SE SL SZ TR TZ UG ZW
 - W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW
 - AU 2001066795 A 20011224 (200227) G01N033-49 <-US 2002193949 A1 20021219 (200303) G01N031-00
 - EP 1309857 A2 20030514 (200333) EN G01N033-49 <-R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
 RO SE SI TR
 - BR 2001011987 A 20031028 (200374) G01N033-49 <--JP 2004503254 W 20040205 (200412) 126 C12Q001-56 <--
- ADT WO 2001096864 A2 WO 2001-US18611 20010608; AU 2001066795 A AU 2001-66795 20010608; US 2002193949 A1 WO 2001-US18611 20010608, US 2001-19087 20011219; EP 1309857 A2 EP 2001-944378 20010608, WO 2001-US18611 20010608; BR 2001011987 A BR 2001-11987 20010608, WO 2001-US18611 20010608; JP 2004503254 W WO 2001-US18611 20010608, JP 2002-510942 20010608
- FDT AU 2001066795 A Based on WO 2001096864; EP 1309857 A2 Based on WO 2001096864; BR 2001011987 A Based on WO 2001096864; JP 2004503254 W Based on WO 2001096864
- PRAI US 2000-591642 20000609; US 2001-19087 20011219
- IC ICM C12Q001-56; G01N031-00; G01N033-49
 ICS G01N033-86
- AB WO 200196864 A UPAB: 20020308
 - NOVELTY Detection of a complex of lipoprotein and an acute phase protein involves: adding at least one reagent to a test sample from a patient in order to cause formation of the complex; measuring the formation of the complex over time so as to derive a time-dependent measurement profile; and determining a slop and/or a time-dependent measurement profile so as to diagnose a condition of the patient.

DETAILED DESCRIPTION - Detection of a complex of at least one human lipoprotein and at least one acute phase protein involves:

- (a) adding at least one reagent to a test sample from a patient comprising at least one part of a **blood** sample from the patient in order to cause formation of the complex, while causing no fibrin polymerization;
- (b) measuring the formation of the complex over time so as to derive a time-dependent measurement profile; and
- (c) determining a slope and/or a time-dependent measurement profile so as to diagnose a condition of the patient.

INDEPENDENT CLAIMS are also included for the following:

- (1) predicting an increased probability of system failure or mortality of a patient involving: obtaining a blood sample from a patient, obtaining plasma or serum from the blood sample, adding the reagent, taking at least one measurement of a parameter of the plasma or serum and correlating the measured parameter to complex formation if present, and correlating the complex formation to the probability of system failure or mortality of the patient; and
 - (2) testing the effectiveness of a therapeutic involving:
 - (a) taking a test sample from a test subject;
- (b) adding a reagent which causes formation of the complex in the test sample;
 - (c) administering to the subject a therapeutic;
 - (d) repeating the steps (a) and (b); and
 - (e) determining if the amount of complex formed has changed.

USE - For predicting an increased probability of system failure or mortality in a patient; diagnosing and treating patient with hemostatic dysfunction (claimed).

ADVANTAGE - The method detects particular abnormality and also monitors the progression of the disease in a single patient. The method is not only useful as early diagnostic and single monitoring marker of disseminated intravascular coagulation (DIC), but the quantifiable and standardizable changes also allow for prognostatic applicability in clinical management.

DESCRIPTION OF DRAWING(S) - The figures illustrate transmittance waveform, on the activated partial thromboplastin time (APTT) assay. Figure A shows a normal appearance, and (B) shows a biphasic appearance. The clot time is indicated by an arrow.

Dwg.1/50

FS CPI EPI

FA AB; GI; DCN

MC CPI: B04-N05; B11-C07; B12-K04A2

EPI: S03-E14H; S03-E14H1

TECH UPTX: 20020308

TECHNOLOGY FOCUS - INORGANIC CHEMISTRY - Preferred Reagent: The reagent is metal ion (preferably divalent transition metal ion). The metal ion comprises at least one calcium, magnesium, manganese, iron or barium. Optionally a clot inhibitor is provided as part of the reagent or as part of an additional reagent added to the test sample. The reagent is capable of causing precipitate formation completely in the absence of fibrin polymerization. The precipitate inhibiting reagent comprises an apolipoprotein capable of binding to a lipoprotein-acute phase protein binding site. The precipitate inhibiting reagent is capable of inhibiting the association of C-reactive protein (CRP) with chylomicrons or their remnants, low density lipoprotein (LDL), very low density lipoprotein (VLDL) and/or intermediate density lipoprotein (IDL).

TECHNOLOGY FOCUS - PHARMACEUTICALS - Preferred Clot Inhibitor:
The clot inhibitor comprises at least one of hirudin, heparin,
PPACK (RTM), I2581 (RTM) or antithrombin.
Preferred Process: The formation of the complex is correlated to the
increase probability of death of the patient, greater the formation of the
complex, the greater the probability of death of the patient. The
time-dependent measurement profile is an optical transmission
profile, and greater the decrease of optical transmittance through the
test sample, greater the formation of the complex. Diagnosing of the
condition of the patient involves a prediction of the probability of
mortality of the patient. The formation of the precipitate is measured at
least once after time to. A single endpoint measurement is made
of precipitate formation after time to. The amount of

fibrin polymerization causes no change in optical transmittance. The method can also involve measuring a formation of a precipitate having the acute phase protein and the lipoprotein followed by addition of inhibiting reagent, before or after adding the precipitate causing reagent, which inhibits at least in part formation of the precipitate and determining the extent of inhibition of the inhibiting reagent. Several measurements are made after addition of the reagent in order to derive the time-dependent measurement profile. Rate of change of the measurements or a total change is determined and hemostatic dysfunction is determined based on the determined total and/or rate change. A single reagent is used prior to taking the measurements such as transmission or absorbance through the sample. The measurements are unaffected by clot formation due to lack of fibrin polymerization. The precipitate inhibiting reagent is either added after all or substantially all of the lipoprotein has become associated with acute phase protein so as to form the precipitate, or added prior to adding the precipitate causing reagent. Measurements are performed over time to derive time -dependent measurement profile. The formation of a complex and additional complex are measured over time to provide respective first and second time-dependent measurement profiles. The measured additional complex and measured initial complex together are correlated to a total amount of acute phase in the test phase. The formation of the complex can also be correlated to a concentration of the lipoprotein.

TECHNOLOGY FOCUS - BIOLOGY - Preferred Components: The human lipoprotein comprises at least one chylomicrons or their remnants, VLDL, IDL, LDL or high density lipoprotein (HDL). The acute phase protein comprises C-reactive protein (CRP) and/or serum amyloid A (SAA) (preferably CRP).

Preferred Complex: A majority of the complex comprises CRP bound to VLDL. ABEX UPTX: 20020308

EXAMPLE - Freshly collected blood samples requiring a prothrombin (PT) or activated partial thromboplastin time (APTT) were analyzed prospectively over a two week working period. The samples were taken in 0.105 M tri-sodium citrate in a ratio of 1 part anticoagulant to 9 parts whole blood. The platelet-poor plasma was analyzed on the multichannel discrete analyzer (MDA). The clot time were derived for PT (normal 11.2 - 15 seconds) using MDA simplastin LS (RTM) and APTT (normal 23-35 seconds) using MDA platelin LS (RTM) with 0.025 M calcium chloride. On analysis the transmittance waveform (TW) for APTT was performed at a wavelength of 580 nm. To ensure no cases of disseminated intravascular coagulation (DIC) were overlooked, a full DIC screen was performed to include the thrombin time; fibrinogen, and D-dimer levels on the Nyocard D-dimer (RTM). Platelet counts performed or an EDTA sample at the same time were recorded. A total of 1,470 samples from 747 patients were analyzed. 174 samples (11.9%) from 54 patients showed the bi-phasic waveform change. DIC was diagnosed in 41 patients with 30 of those requiring transfusion support with fresh frozen plasma, cryoprecipitate or platelets. 40 of the 41 patients with DIC showed the bi-phasic TW. The one false negative result (DIC without a bi-phasic TW) occurred in a patient with pre-eclampsia where the single sample showed a prolonged PT of 21 second, APTT of 44 seconds and raised D-dimer of 1.5 mg/liter. The results showed that the bi-phasic TW had a sensitivity of 97.6% and specificity of 98% for the diagnosis of DIC. The positive predictive value of the test was 74%, which increased with increasing steepness of the bi-phasic slope and decreasing

levels of light transmittance.

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2000-572005 [53]; 2003-605936 [57]; 2003-615959 [58]; 2004-141748 [14];
CR
     2004-224645 [21]
                        DNC C2002-030637
DNN
    N2002-072440
     Measuring efficacy of anti-platelet agents, comprises
TI
     determining blood coagulation parameters of
     blood samples obtained in the presence and absence of anti-
     platelet therapy, using blood coagulation
     analyzers.
DC
     B04 D16 S03
IN
     COHEN, E
     (HAEM-N) HAEMOSCOPE CORP
PA
CYC
    97
     WO 2001096879
                     A2 20011220 (200213) * EN
PΙ
                                                20
                                                      G01N033-86
        RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ
            NL OA PT SD SE SL SZ TR TZ UG ZW
         W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK
            DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR
            KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU
            SD SE SG SI SK SL TJ TM TR TT TZ UA UG UZ VN YU ZA ZW
     AU 2001075249
                     A 20011224 (200227)
     EP 1287349
                     A2 20030305 (200319)
                                           EN
                                                      G01N033-49
         R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
            RO SE SI TR
     US 6613573
                     B1 20030902 (200359)
                                                      G01N033-86
     CN 1436304
                     A 20030813 (200373)
                                                      G01N033-49
                                                                      <--
                    W 20040205 (200412)
     JP 2004503781
                                                34
                                                      G01N033-86
                                                                      <--
    WO 2001096879 A2 WO 2001-US18154 20010605; AU 2001075249 A AU 2001-75249
ADT
     20010605; EP 1287349 A2 EP 2001-941938 20010605, WO 2001-US18154 20010605;
     US 6613573 B1 CIP of US 1999-255099 19990222, US 2000-591371 20000609; CN
     1436304 A CN 2001-810898 20010605; JP 2004503781 W WO 2001-US18154
     20010605, JP 2002-510957 20010605
FDT
    AU 2001075249 A Based on WO 2001096879; EP 1287349 A2 Based on WO
     2001096879; US 6613573 B1 CIP of US 6225126; JP 2004503781 W Based on WO
     2001096879
PRAI US 2000-591371
                          20000609; US 1999-255099
                                                         19990222
     ICM G01N033-49; G01N033-86
         A61K031-195; A61K031-197; A61K031-352; A61K031-727; A61K038-46;
          A61K038-48; A61P007-02; A61P007-04; G01N033-15
AB
     WO 200196879 A UPAB: 20040421
     NOVELTY - Measuring (M) the efficacy of anti-platelet agents
     (A), involves determining a first blood coagulation
     parameter (P1) of a first blood sample obtained in the absence
     of anti-platelet therapy (AT) and determining a second
    blood coagulation parameter (P2) of a second
    blood sample in the presence of AT, using a blood
     coagulation analyzer (I), and determining the efficacy of (A)
     based on P1 and P2.
          DETAILED DESCRIPTION - Measuring (M) the efficacy of anti-
    platelet agents (A), involves determining a first blood
     coagulation parameter (P1) of a first blood sample
     obtained in the absence of anti-platelet therapy (AT) and
     determining a second blood coagulation parameter (P2)
     of a second blood sample in the presence of AT, using a
    blood coagulation analyzer (I), and determining the
     efficacy of (A) based on P1 and P2. (I) is capable of measuring a
     clot strength in the range of about 100-1000 dyn/cm2.
          An INDEPENDENT CLAIM is also included for an apparatus for measuring
     the efficacy of AT, which comprises:
          (a) a blood coagulation analyzer (I) (10)
     operable to measure a first blood sample (13) in the absence of
    AT and a second blood sample in the presence of AT to
    respectively generate P1 and P2, where P1 and P2 are related to
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blood clot strength; and

(b) a processor having an associated control program for directing the operation of the processor for determining the efficacy of AT based upon P1 and P2. USE - (I) and (M) are useful for measuring the efficacy of antiplatelet agents (claimed). The measured blood coagulation parameters permit confirmation of the attainment of therapeutic level of GPIIb/IIIa receptor blockade, individualized dosing assessment to evaluate attainment of adequate GPIIb/IIIa receptor blockage, individualized dosing assessment required to reach adequate GPIIb/IIIa receptor blockade, illustration of the rate of diminishment of platelet inhibition or inhibition recovery after treatment with platelet-inhibition drugs, evaluation of the interaction effect of a combination of thrombolytic or any other agents or conditions effecting hemostasis and platelet -inhibiting agents on patient hemostasis. ADVANTAGE - The blood coagulation analyzer is utilized to measure continuously in real time, the clotting process from the initial fibrin formation, through platelet fibrin interaction and lysis to generate blood coagulation parameters. DESCRIPTION OF DRAWING(S) - The figure shows the blood coagulation analyzer. Blood coagulation analyzer 10 Blood sample 13 Dwg.2/4 CPI EPI AB; GI; DCN CPI: B04-B04D5; B04-F04; B04-H19; B11-C08; B14-F04; D05-H09 EPI: S03-E14H TECH UPTX: 20020226 TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: Determination of P1 involves: (a) testing a sampled portion of the first blood sample to determine a first characteristic; and (b) testing a portion of the first **blood** sample treated in vitro to inhibit thrombin activation, and to preserve fibrinogen and platelet activation to determine a second characteristic. Determination of P2 involves: (a) testing a sampled portion of the second blood sample to determine a first characteristic; and (b) testing a portion of the second blood sample treated in vitro to inhibit thrombin activation, and to preserve fibrinogen and platelet activation to determine a second characteristic. (M) is performed in conjunction with administration of at least one of the blood coagulation therapies including: (i) anti-coagulation therapy utilizing the administration of heparin or warfarin; (ii) thrombolytic therapy utilizing the administration of tPA, steptokinase and urokinase; (iii) anti-fibrinolytic therapy utilizing e-amino-caproic acid, trasylol and tranexamic acid; (iv) anti-platelet therapy and a blood component transfusion therapy. (M) is performed in connection with assessment of thrombotic risk, hypercoagulable condition or hypocoagulable condition. P1 and P2 are based upon the latency period of clot formation, rate of clot formation, maximum clot strength or rate of clot lysis. P1 and P2 correspond to an estimate of fibrin-platelet

interaction. P1 is represented by a first maximum amplitude measurement

FS FA

MC

and P2 is represented by a second maximum amplitude measurement, and (M) involves comparing the amplitudes. Preferred Apparatus: (I) provides for the substantially simultaneous testing of the first and second blood samples. ABEX UPTX: 20020226 EXAMPLE - None given. L62 ANSWER 8 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2002-091713 [13] WPIX AN DNC C2002-028520 DNN N2002-067550 Testing system for coagulation promoting substance has sample TI wells for measuring test clotting indicator time of patient's blood and coagulation promoting substance as test sample, and of patient's blood as control sample. B04 D16 S03 DC IN GOLDSTEIN, S (GOLD-I) GOLDSTEIN S PA CYC 28 PΙ EP 1162457 A2 20011212 (200213) * EN 17 G01N033-49 R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI TR AU 2001051826 A 20011213 (200213) G01N035-02 A1 20011209 (200213) EN CA 2349959 G01N033-86 EP 1162457 A2 EP 2001-202205 20010608; AU 2001051826 A AU 2001-51826 ADT 20010608; CA 2349959 A1 CA 2001-2349959 20010608 PRAI US 2000-591329 20000609 ICM G01N033-49; G01N033-86; G01N035-02 IC ICS C12Q001-56 AB 1162457 A UPAB: 20020226 NOVELTY - An automated multiple coagulation testing system has sample wells for measuring a test clotting indicator time of the patient's blood and coagulation promoting substance as a test sample; and for measuring a baseline clotting indicator time of the patient's blood as a control sample. An appropriate therapy is determined by comparing clotting indicator time of control sample and test sample.

DETAILED DESCRIPTION - An automated multiple coagulation testing system includes at least three sample wells for receiving patient's blood (35), at least two other sample wells (75A-D) for measuring a test clotting indicator time of the patient's blood and coagulation promoting substance (105A) as a test sample. At least one of the sample wells (75-E) is for measuring a baseline clotting indicator time of the patient's blood as a control sample. The control sample wells are free of coagulation promoting substance. The test sample wells (95A-D) each contain a different coagulation promoting substance. The coagulation substance is an agent or combination of agents capable of improving clotting function in the patient. The sample wells are constructed and arranged to allow detection of a clotting indicator in the patient's blood for measuring clotting indicator time. An appropriate therapy for improving clotting function in the patient is determined by comparison of the baseline clotting indicator time of the control sample with the test clotting indicator time of the patient's blood and the coagulation promoting substance.

An INDEPENDENT CLAIM is also included for a method of determining an appropriate coagulation promoting substance for administration to a patient as a therapy for improving clotting function involving adding a selected amount of a patient's blood to each of the at least three sample wells, and adding a different coagulation promoting substance to each of the test sample wells.

USE - For determining an appropriate **coagulation** promoting substance for administration to a patient as a therapy for improving **clotting** function.

ADVANTAGE - The system produces results indicating a proper course treatment without resort to a shotgun approach, which requires an addition of multiple agents to a patient and thus avoids several of the complications inherent in using such approach. The system thus allows rapid determination of a specific treatment in a hemorrhaging situation without awaiting standard laboratory test results.

DESCRIPTION OF DRAWING(S) - The figure shows a schematic view of the testing system.

Blood 35

Sample wells for measuring test **clotting** indicator time of test sample 75A-D

Sample well for measuring baseline clotting indicator time of control sample 75-E

Sample wells containing **coagulation** promoting substance 95A-D

Coagulation promoting substance 105A-D Coagulation detector 125A-E

Dwg.2/2

FS CPI EPI

FA AB; GI; DCN

MC CPI: B04-B04D; B04-H19; B04-H20A; B11-C07B4; B11-C08E;

B12-K04E; D05-H09
EPI: S03-E14H; S03-E14H1

TECH UPTX: 20020226

TECHNOLOGY FOCUS - INSTRUMENTATION AND TESTING - Preferred Component: The sample wells include tubes for containing the blood, and filter paper for receiving the blood. The system has a magnetic rod in each of the tubes, and a magnetic detector (125A-E) triggerable by displacement of the magnetic rods. It has a light source, a photo-optical detector, a viscometer, holder for containing a patient's blood, an aliquot meter connected with the holder for withdrawing a predetermined measured amounts of the patient's blood, and dosing meters connected with the test sample wells for withdrawing a preselected equivalent dose of the coagulation promoting substances from the test sample wells. The holder is removably attached in connection with the aliquot meter. The test sample wells are removably attached in connection with the dosing meters. Each well contains diatomaceous powder for increasing the surface area for contact of substances involved in clotting. There are 4-10 wells used. Preferred Mechanism: The clotting indicator is detected by the

magnetic detector when displacement of the magnetic rods due to blood clotting occurs in any of the tubes, or when a change of light transmission from the light source to the detector due to blood clotting occurs in any of the sample wells. The clotting indicator is detected by the viscometer when a change of viscosity due to blood clotting occurs in the sample wells.

wells.

Preferred Substance: The coagulation promoting substance is coagulation factors, recombinant coagulation factors, bovine coagulation factors, coagulation factor VIII:C, von Willebrand factor, platelets, fibronectin, thrombin, desmopressin acetate, epsilo-amino caproic acid, cryoprecipitate, fresh frozen plasma, protamine, aprotinin or calcium ion. The coagulation factor is coagulation factor I (fibrinogen), Ia (fibrin), II (prothrombin), IIa (thrombin), III (thromboplastin), or IV-XIII, preferably recombinant factor VIII. The cryoprecipitate is bovine or human cryoprecipitate. The fresh frozen plasma is bovine or human fresh frozen plasma. The coagulation inhibiting substance is heparin,

aprotinin, carbacyclin, prostacyclin, prostaglandin El, or abciximab.

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L62 ANSWER 9 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     2002-075349 [10]
                        WPIX
DNN N2002-055548
                        DNC C2002-022532
TT
     Inducing or reducing hemostasis in a subject for treating
     hypocoagulation disorders, vasculature-associated diseases and
     thrombotic disorders, comprises administering to the subject a
     modulator of P-selectin activity.
DC
     B04 D16 S03
IN
     ANDRE, P; HARTWELL, D W; HRACHOVINOVA, I; WAGNER, D D
     (BLOO-N) CENT BLOOD RES; (BLOO-N) CENT BLOOD RES INC; (ANDR-I) ANDRE P;
PA
     (HART-I) HARTWELL D W; (HRAC-I) HRACHOVINOVA I; (WAGN-I) WAGNER D D
CYC
     97
PΙ
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                                                      A61K038-48
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     EP 1289552
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    WO 2001089564 A2 WO 2001-US16021 20010517; AU 2001061735 A AU 2001-61735
ADT
     20010517; US 2002031508 A1 Provisional US 2000-205734P 20000519, US
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    AU 2001061735 A Based on WO 2001089564; EP 1289552 A2 Based on WO
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                                                         20010517
     ICM A61K038-48; A61K039-395; A61K045-00
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         A61K035-14; A61K038-00; A61K038-17; A61K048-00; A61P007-00;
          A61P007-02; A61P007-04; A61P009-00; A61P009-10; A61P035-00;
          A61P043-00; G01N033-50; G01N033-68;
          G01N033-86
AB
     WO 200189564 A UPAB: 20020213
     NOVELTY - Inducing (M1) or reducing hemostasis in a subject, comprising
     administering an inducer or inhibitor of P-selectin activity, a soluble
     P-selectin polypeptide (I), an isolated polynucleotide comprising a
     sequence encoding (I), a recombinant cell expressing (I), or an isolated
     nucleic acid molecule comprising a sequence antisense to a sequence
     encoding (I), is new.
          DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the
     following:
          (1) modulating (M2) hemostatic potential in a subject,
     involves modulating P-selectin activity in the subject;
          (2) diagnosing (M3) a procoagulant state in a subject,
     involves determining a P-selectin activity in a biological sample of the
     subject, where an increased P-selectin activity in the sample indicates a
    procoagulant state in the subject;
          (3) identifying (M4) a subject having a thrombotic
     disorder, or at risk for developing a thrombotic disorder,
     comprising determining P-selectin activity in a biological sample of the
     subject, where an increased P-selectin activity in the sample identifies a
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- (4) identifying (M5) a compound capable of modulating hemostasis, comprising:
 - (a) assaying the ability of the compound to modulate a P-selectin

subject having a thrombotic disorder, or at risk for developing

a thrombotic disorder;

activity; and

- (b) identifying a compound capable of modulating hemostasis;
- (5) a pharmaceutical composition (II) for modulating hemostasis comprising a compound identified by M5; and
- (6) a pharmaceutical composition (III) for modulating hemostasis containing at least one compound which is a modulator of P-selectin activity.

ACTIVITY - Hemostatic; antitumor; antiarteriosclerotic; thrombolytic; antianginal; vasotropic; antidiabetic; ophthalmological; antipsoriatic; dermatological; antiinflammatory; antiallergic; cytostatic; gynecological; antirheumatic; antiarthritic; cerebroprotective; cardiant; cytostatic. delta CT mice were treated with soluble P-selectin (PSGL)-Ig. Soluble PSGL-Ig infusion decreased the procoagulant phenotype of delta CT mice as shown by a significant decrease in the number of microparticles and a prolonged clotting time of plasma. Infusion of control Ig had no such effect. The clotting time was significantly longer in mice treated with soluble PSGL-Ig than in control Ig treated group.

MECHANISM OF ACTION - Modulator of hemostasis (claimed). No biolgical data provided.

USE - M1 is useful for modulating hemostasis in a subject. M1 is useful for treating or preventing a disorder (such as hemorrhagic disorder or hemophilia) associated with hypocoagulation in a subject, by administering to the subject an inducer of P-selectin activity or (I)1, such that the disorder associated with hypocoagulation is treated or prevented. M1 is useful for treating a vasculatureassociated disease such as tumor in a subject, by administering an inducer of P-selectin activity or (I), to the subject. The subject is further treated with a molecule effective to induce a procoagulant state in tumor associated vasculature. The molecule comprises a first binding region that binds to a component of a tumor cell or tumor associated vasculature, operatively linked to a coagulation factor or a second binding region that binds to a coagulation factor. The first binding region comprises an antibody or its fragment, that binds to VCAM-1, operatively linked to tissue factor. M1 is useful for treating or preventing a thrombotic disorder in a subject, by administering to the subject an inhibitor of P-selectin activity, such that the thrombotic disorder is treated or prevented. The thrombotic disorder is arteriosclerosis; deep vein thrombosis, angina or restenosis following medical intervention (claimed). M1 is useful for treating disorders resulting from a deficiency in clotting factors or platelet ligands, e.g., a deficiency in von Willebrand's factor resulting in von Willebrand's disease, vasculature-associated diseases such as benign and malignant tumors or growth, diabetic retinopathy, vascular restenosis, neovascular glaucoma, psoriasis, synovitis, dermatitis, endometriosis, angiofibroma, rheumatoid arthritis, atherosclerotic plaques, corneal graft neovascularization, hemophilic joints, hypertrophic scars, and Osler-Weber syndrome, and thrombotic disorder such as thromboembolism, stroke, myocardial infarction, inflammatory disorders, cancer metastasis or sickle cell disease. Dwg.0/14 CPI EPI

FS

AB; DCN FA

MC CPI: B04-B04D5; B04-C01; B04-E03F; B04-E06; B04-F0100E; B04-N02;

B11-C08E; B12-K04A2; B14-F01D; B14-F01G; B14-F04;

B14-F07; B14-F08; B14-H01B; D05-C07; D05-C11;

D05-H09; D05-H12A; D05-H12D2; D05-H14B2; D05-H17A6

EPI: S03-E14H

TECH UPTX: 20020213

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: In M1, the inducer of P-selectin activity increases the level of (I) in the plasma of the subject, increases the proteolytic cleavage of

P-selectin from a cell surface or increases P-selectin gene expression. The inducer of P-selectin activity binds to a P-selectin receptor or ligand and mimics the activity of a P-selectin polypeptide. The inducer of P-selectin activity is an antibody to a P-selectin receptor of ligand, where the antibody or ligand is PSGL-1. The inhibitor of P-selectin activity decreases the level of soluble P-selectin polypeptide in the plasma of the subject, decreases the proteolytic cleavage of P-selectin from the cell surface, or decreases P-selectin gene expression. The inhibitor of P-selectin activity is an anti-P-selectin antibody, or a recombinant soluble PSGL-1. In M2, modulating involves administering to the subject a modulator of P-selectin activity, where the modulator regulates the level of soluble P-selectin in the plasma of the subject. M3 involves providing a test sample of blood from a subject and comparing the level of soluble P-selectin in the test sample to the level of soluble P-selectin in a control blood sample from a subject with normal hemostatic activity, where an increased level of soluble P-selectin in the test samples as compared to the control sample is an indication of a procoagulant state in the subject. M4 involves contacting a sample of blood obtained from the subject with a P-selectin binding substance, and detecting the presence of increased levels of soluble P-selectin in the sample, thus identifying a subject having a thrombotic disorder, or at risk for developing a thrombotic disorder. In M5, the P-selectin activity is the expression of soluble P-selectin.

ABEX

UPTX: 20020213

WIDER DISCLOSURE - Also disclosed as new, is an active agent which modulates soluble P-selectin expression or activity.

ADMINISTRATION - Administration is parenteral, intravenous, intradermal, subcutaneous, oral, by inhalation, transdermal, topical, transmucosal or rectal. Dosage is 0.001-30 mg/kg body weight, preferably 5-6 mg/kg body weight.

EXAMPLE - No relevant example is given.

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L62 ANSWER 10 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT ON STN
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AN 2001-316462 [33] WPIX

DNN N2001-227460 DNC C2001-097551

TI Monitoring the effect of Factor Xa inhibitors, comprises collecting plasma sample from patient, who has received FXa inhibitor, an anticoagulant, an antithrombotic agent, adding solution of Russell's viper venom to plasma sample.

DC B04 S03

IN FUNG-HWEI, V C; CHU, V F

PA (AVET) AVENTIS PHARMA DEUT GMBH; (CHUV-I) CHU V F

CYC 95

PI WO 2001033217 A2 20010510 (200133)* EN 11 G01N033-50 <-RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ
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AU 2001015157 A 20010514 (200149) G01N033-50 <--EP 1230382 A2 20020814 (200261) EN C12Q001-56 <--

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ADT WO 2001033217 A2 WO 2000-EP10646 20001028; AU 2001015157 A AU 2001-15157

20001028; EP 1230382 A2 EP 2000-977438 20001028, WO 2000-EP10646 20001028; US 2003049704 A1 Cont of WO 2000-EP10646 20001028, US 2002-125627 20020418; JP 2003513280 W WO 2000-EP10646 20001028, JP 2001-535051 20001028; EP 1230382 B1 EP 2000-977438 20001028, WO 2000-EP10646 20001028 AU 2001015157 A Based on WO 2001033217; EP 1230382 A2 Based on WO 2001033217; JP 2003513280 W Based on WO 2001033217; EP 1230382 B1 Based on WO 2001033217

PRAI GB 1999-30535 19991223; US 1999-163161P 19991102

IC ICM C12Q001-56; G01N033-50; G01N033-86 ICS G01N033-15

WO 200133217 A UPAB: 20010615

AB

NOVELTY - A method of monitoring the effect of Factor Xa (FXa) inhibitors comprising:

- (a) collecting a plasma sample from a patient, who has received a FXa inhibitor, an anticoagulant, an antithrombotic agent, or any combination;
- (b) adding a solution of Russell's viper venom to the plasma sample and
- (c) measuring clotting time or a chromogenic change.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

- (1) a method of monitoring the effect of Factor Xa inhibitors comprising of:
 - (a) collecting a plasma sample from a mammal;
- (b) providing a Factor X deficient plasma sample of the same species to be used to make serial dilutions of the normal plasma sample;
- (c) adding a solution of Russell's viper venom to the plasma samples;
- (d) comparing the clotting time measured for the plasma sample with the clotting time measured for the plasma samples diluted with Factor X deficient plasma;
- (e) constructing a **standard** curve of % FXa activity (proportional to the **normal plasma** content) and **clotting time** prolongation; and
- (f) knowing the clotting time of the mammal, which received FXa inhibitor treatment, % residual FXa activity or % FXa inhibition is obtained from the standard curve;
- (2) a method of monitoring the effect of Factor Xa inhibitors comprising:
 - (a) collecting a plasma sample from a mammal;
- (b) dividing the **plasma** sample into portions, saving one portion as the control **normal plasma** and adding serial dilutions of Factor Xa inhibitor to other portions;
- (c) adding a solution of Russell's viper venom to plasma samples defined in (b);
- (d) comparing the clotting time measured for the plasma sample without Factor Xa inhibitor with that plasma samples with added Factor Xa inhibitor;
- (e) constructing a dose-dependent clotting time prolongation curve and determining the concentration of a FXa inhibitor required to prolong the clotting time twice longer than the control plasma clotting time; and
- (3) a method of monitoring the effect of Factor Xa inhibitors comprising:
 - (4) collecting a plasma sample from a mammal;
- (a) dividing said plasma sample into portions, saving one portion as the control plasma and adding serial dilutions of Factor Xa inhibitor to other portions;
- (b) adding a solution of Russell's viper venom (RVV-X) to
 plasma samples defined in (b);

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(c) comparing the FXa activity measured for the control
     plasma with the residual FXa activity measured for the other
     plasma samples with added Factor Xa inhibitor;
          (d) constructing a standard curve of dose-dependent
     inhibition of RW-X induced by FXa inhibitor; and
          (e) the concentration of FXa inhibitor is estimated by using the
     standard curve.
          USE - Russell's viper venom-induced plasma factor Xa
     monitors the safety and efficacy of intravenous or orally active FXa
     inhibitors, thrombin inhibitors and indirect FXa inhibitors such
     as anticoagulants for factors upstream of the
     coagulation cascade and heparin, more particularly low molecular
     weight heparin.
          ADVANTAGE - None given.
     Dwg.0/0
     CPI EPI
    AB; DCN
    CPI: B04-B04D4; B04-B04G; B04-H19; B07-D04; B11-C08E;
          B11-C08E2; B12-K04A; B12-K04E; B14-F08; B14-L01; B14-L06
     EPI: S03-E14H
TECH
                    UPTX: 20010615
     TECHNOLOGY FOCUS - PHARMACEUTICALS - Preferred Method: The
     residual FXa activity is measured chromogenically. Cephalin is added to
     the plasma samples. The plasma samples were incubated
     in a buffer having a pH from 7 - 8.
     Preferred Inhibitor: The FXa inhibitor is methyl-3-(4'-Noxopyridylphenoyl)-
     3-methyl-2-(m-amidinobenzyl)-propionate.
    ANSWER 11 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
L62
    2001-315989 [33]
                        WPTX
DNN N2001-227171
                        DNC C2001-097248
     Quantitative determination of soluble fibrin in opaque sample,
     useful for early detection of disseminated intravascular
     coagulation, by determining time taken to detect
     fibrin precipitate.
    B04 D16 S03
    BULL, B S; KORPMAN, R A; HAY, K L
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    JP 2003508060
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    US 2003059836
                    A1 20030327 (200325)
                                                      G01N033-53
                                                                     <---
ADT
    WO 2001016360 A1 WO 2000-US24106 20000901; AU 2000073431 A AU 2000-73431
    20000901; EP 1214448 A1 EP 2000-961486 20000901, WO 2000-US24106 20000901;
    US 6436655 B1 CIP of US 1998-21062 19980209, US 1999-388796 19990902; JP
    2003508060 W WO 2000-US24106 20000901, JP 2001-520905 20000901; US
    2003059836 Al CIP of US 1998-21062 19980209, Cont of US 1999-388796
    19990902, US 2002-136037 20020429
    AU 2000073431 A Based on WO 2001016360; EP 1214448 A1 Based on WO
    2001016360; JP 2003508060 W Based on WO 2001016360; US 2003059836 A1 Cont
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19990902; US 1998-21062

19980209;

FS

FA MC

AN

ТT

DC

IN

PA

PΙ

PRAI US 1999-388796

US 2002-136037 20020429
IC ICM C12Q001-56; G01N033-53
ICS C07K007-08; C12Q001-37; G01N033-49;
G01N033-86

AB WO 200116360 A UPAB: 20010615

NOVELTY - Determining (M1) the existence and the amount of soluble fibrin (SF) contained in an opaque specimen fluid is new.

DETAILED DESCRIPTION - (M1) comprises:

- (1) mixing a portion of the opaque specimen fluid in a transparent container with a sufficient amount of precipitating reagent under a condition to cause the soluble **fibrin** to precipitate;
- (2) aggregating and concentrating the soluble **fibrin** precipitates in a region of the container for rendering the precipitates optically detectable in the opaque specimen fluid;
 - (3) optically detecting the precipitates; and
- (4) recording the **time** when the precipitates are first become optically detectable in the opaque specimen fluid, where the time elapsed from the addition of the precipitating reagent to the detection of the aggregated precipitates is an inverse measure of the quantity of soluble **fibrin** present in the opaque specimen fluid.

USE - The method is particularly used to detect SF during major surgery or in cases of severe trauma, where disseminated intravascular coagulation (DIC) is a risk. SF is a sensitive indicator of early DIC.

ADVANTAGE - The method is rapid and quantitative, and can be performed on whole blood, i.e. in the operating theater without need for separation of plasma, allowing immediate action to be taken to combat DIC. By collecting precipitate in a selected region of the container, the precipitate can be detected optically, even in opaque samples.

Dwg.0/3 CPI EPI

FS CPI EPI

FA AB; DCN

MC CPI: B04-B04D5; B04-B04H; B04-C03; B04-N02; B11-C07B2; B11-C08D3; B11-C09; B12-K04A; D05-H09; D05-H13

EPI: S03-E09; S03-E14H1

TECH

UPTX: 20010615

TECHNOLOGY FOCUS - BIOLOGY - Preferred Process: The time at which the precipitate sticks to, and rotates with, the container is also measured; it also is inversely related to SF content. Step (ii) particularly involves placing the container in a rocking/rotating device. The measured time is compared with a standard curve, produced by analyzing samples of known SF contents. The time is determined e.g. by direct visual examination, using a charge-coupled device camera etc. Optionally a second portion of the sample is tested separately, using either a different precipitant or a different concentration of the same precipitant, the result processed as above and an average value of the two determinations taken. Preferred Sample: This is whole blood, a bloody effusion or a bloody cerebrospinal fluid sample, preferably diluted with buffer (pH 5-9) and processed at 37 degrees centigrade.

Preferred Precipitant: This is protamine sulfate and/or polybrene. ABEX UPTX: 20010615

EXAMPLE - Blood samples (150 microliter) were taken at 30 min intervals from a subject undergoing a liver transplant operation. Samples were diluted with saline (450 microliter) then treated with protamine sulfate (20 microliter) and the mixture placed in a hemostatic analyzer that rocked and rotated the sample. The sample was kept at 37 degrees centigrade and the time at which precipitate was first visible was taken as the first end point and the time at which the precipitate adhered to, and rotated with, the container was taken as the second end point. The level of soluble fibrin (SF) was below 20 units during the early stages of the operation but increased sharply,

to about 70 units, during the anhepatic stage, followed by a slight decline. The SF level was strongly positively correlated with levels of both fibrin degradation products and D-dimer, also strongly negatively correlated with levels of fibrinogen.

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L62 ANSWER 12 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     2001-234924 [24]
AN
                        WPIX
CR
     1997-052574 [05]; 1999-458146 [38]; 2000-514997 [46]; 2001-091718 [10];
     2002-147005 [19]
DNC
    C2001-070340
ΤI
     Detecting presence of hemostatic dysfunction, useful e.g. for
     diagnosing or monitoring of disseminated intravascular coagulation
     , by precipitation without fibrin polymerization.
DC
     B04
     DOWNEY, C; FISCHER, T J; TOH, C H
IN
     (ALKU) AKZO NOBEL NV; (DOWN-I) DOWNEY C; (FISC-I)
PA
     FISCHER T J; (TOHC-I) TOH C H; (INMR) BIOMERIEUX SA
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FDT
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PRAI US 1999-372954
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     US 1997-1647
                          19971231; US 2002-156462
                                                         20020528
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     ICM G01N033-86
     TCS
         A61P007-04; G01N033-15; G01N033-48;
          G01N033-50; G01N033-53
AB
     WO 200113125 A UPAB: 20030603
    NOVELTY - Method comprising treating a test sample, containing
     at least one component of blood, with a reagent (R) then
     measuring formation of a precipitate (P) over time to produce a
     time-dependent measurement profile. (R) forms a precipitate
     without significant polymerization of fibrin.
          DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the
     following:
          (1) determining presence of a complex (C) of proteins, comprising at
     least one of serum amyloid A and C-reactive protein (CRP);
          (2) methods for determining possibility or probability of
    hemostatic dysfunction;
          (3) method for monitoring an inflammatory condition using
     (R);
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(4) method for diagnosing or treating hemostatic

(5) immunoassay for diagnosing HD by detecting or quantifying CRP or

dysfunction (HD) using (R);

a 300 kD protein (I); and

(6) method for testing new drugs on humans or animals having an inflammatory condition and/or HD. ACTIVITY - Anticoagulant; Antithrombotic; Antiarthritic; Antiinflammatory; Antibacterial; Immunosuppressive; Antirheumatic. MECHANISM OF ACTION - None given. USE - (R) is used (i) to diagnose hemostatic dysfunction (HD), particularly disseminated intravascular coagulation (DIC) or a condition that can lead to DIC, bleeding or thrombosis, also to optimize and monitor treatment, (ii) to monitor an inflammatory condition (rheumatoid arthritis, sepsis or conditions caused by surgical trauma) or (iii) to screen for new drugs for treatment of HD or inflammation. ADVANTAGE - The method provides early indication of disseminated intravascular coagulation, and since it can be standardized and made quantitative, it is suitable for prognosis and monitoring. It is simple and provides results quickly. Dwg.0/29 CPI AB; DCN CPI: B04-B04D4; B04-B04D5; B04-N02; B05-A01B; B05-A03A; B11-C07A; B12-K04A2; B14-A01; B14-C03; B14-C09B; B14-F04; B14-G02; B14-S12 TECH UPTX: 20010502 TECHNOLOGY FOCUS - BIOLOGY - Preferred reagent: (R) contains a metal ion, preferably divalent, and especially calcium, magnesium, manganese, iron or barium. It may also include a clotting inhibitor (CI), e.g. hirudin, heparin, PPACK, I2581 or antithrombin, or CI is provided in another reagent. (R) causes formation of (P) completely in absence of fibrin polymerization. Preferred precipitate: (P) comprises a protein of about 20 kD that is insoluble in saline, ethylenediamine tetraacetic acid or imidazole but soluble in 5 M urea. Preferred process: The formation of (P) is correlated with HD, with increased amounts of (P) indicating more severe dysfunction, and this can be quantified by constructing a reference curve for comparison with the patient sample. Especially the profile is an optical transmission or absorbance profile, with a greater reduction in transmission indicating a greater formation of (P). If any fibrin polymerization does occur, then it does not cause a change in optical transmittance. (R) is added in absence of clot-inducing reagents and either a single (end-point) measurement is made or several measurements, in which case HD is detected from the rate of change. The test sample is particularly plasma and the test may be repeated at different (R)/plasma ratios or at different times (to monitor progression or regression of disease). In method (a), a test sample (preferably blood or a blood component) is treated with an alcohol (especially (m)ethanol), CI and metal cation. The precipitate forms contains (C). In method (b), a coagulation reagent (specifically a prothrombin (PT) or activated partial thromboplastin time (APTT) reagent) is added to a sample and formation of fibrin monitored over time by measuring some parameter that changes due to addition of reagent. The rate of change of this parameter, before fibrin is formed in the sample, is determined and if the rate exceeds a predetermined value, a second aliquot of sample is treated with (R) and the formation of precipitate monitored over time. In method (c), some parameter indicative of (P) is measured over time, the rate of change calculated and the process repeated at various times, with a change in the rate indicating progression or regression of the inflammatory

state. The parameter is optical transmission or absorbance. In method (d), a sample is treated with (R) and some parameter that

FS

FA

MC

changes due to formation of (P) is measured over time and its rate of change calculated. HD is diagnosed if the rate exceeds a predetermined level and appropriate treatment is administered, e.g. (i) antibiotic and/or CI or (ii) identification and correction of the underlying cause, e.g. administration of broad-spectrum antibiotic; evacuation of the uterus in abruptio placentae; blood replacement; administration of platelet concentrate (to correct thrombocytopenia), fresh plasma, blood factors and/or interleukin-1. The procedure may be repeated to optimize treatment. In method (e), a test sample is treated with a ligand (L) that can bind to CRP or (I), and this detected as part of a complex of proteins formed by adding a divalent metal cation. CRP may be intact, modified, cleaved or mutant. In method (f), a test sample is treated with (R) and kinetic or end-point measurements of precipitate formation made. A drug is then administered and the assay repeated, with an increase/decrease in precipitation indicating an effective drug. UPTX: 20010502 EXAMPLE - The plot of transmission against time in a

EXAMPLE - The plot of transmission against time in a standard activated partial thromboplastin time (APTT) assay is normally sigmoid but in patients with disseminated intravascular coagulation (DIC) it is biphasic, with an initial region of low gradient and a subsequent region of steeper slope. The slope measured before start of clot formation is a significantly more specific and sensitive indicator of DIC than analysis of transmittance at a particular time. Particularly this slope was -0.001, or more negative, for all DIC patients and was -0.005 or more negative for 85 of 91 of them. Normal subjects, and those with abnormalities other than DIC, never had values more negative than -0.0002.

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L62
    ANSWER 13 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     2001-091718 [10]
                        WPIX
     1997-052574 [05]; 1999-458146 [38]; 2000-514997 [46]; 2001-234924 [24];
CR
     2002-147005 [19]
DNN
    N2001-069462
                        DNC C2001-027107
     Presentation of relationship between data comprises deriving at least one
TT
     time-dependent measurement of the known blood samples,
     and computing standard deviation and determining the z-score for
     each derived predictor variable.
```

DC B04 S03

IN BECK, L; BRAUN, P; GIVENS, T B; GIVENS, T

PA (ALKU) AKZO NOBEL NV; (BECK-I) BECK L; (BRAU-I) BRAUN P; (GIVE-I) GIVENS T; (INMR) BIOMERIEUX INC

CYC 24

ABEX

PI WO 2001001152 A1 20010104 (200110) * EN 36 G01N033-86 <-RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE
W: AU CA JP KR US

AU 2000060676 A 20010131 (200124)

US 2002010553 A1 20020124 (200210) G06F019-00 EP 1188060 A1 20020320 (200227) EN G01N033-86

R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE

KR 2002042539 A 20020605 (200277) G01N033-86 <-

US 6502040 B2 20021231 (200305) G01N021-00

JP 2003529046 W 20030930 (200365) 28 G01N033-86 <-AU 768436 B 20031211 (200404) G01N033-86 <--

ADT WO 2001001152 A1 WO 2000-US18310 20000630; AU 2000060676 A AU 2000-60676 20000630; US 2002010553 A1 CIP of US 1997-1647 19971231, US 1999-345080 19990630; EP 1188060 A1 EP 2000-947001 20000630, WO 2000-US18310 20000630; KR 2002042539 A KR 2001-716754 20011228; US 6502040 B2 CIP of US 1997-1647 19971231, US 1999-345080 19990630; JP 2003529046 W WO 2000-US18310

20000630, JP 2001-507105 20000630; AU 768436 B AU 2000-60676 20000630

FDT AU 2000060676 A Based on WO 2001001152; US 2002010553 A1 CIP of US 6321164; EP 1188060 A1 Based on WO 2001001152; JP 2003529046 W Based on WO

19971231

2001001152; AU 768436 B Previous Publ. AU 2000060676, Based on WO 2001001152

PRAI US 1999-345080 19990630; US 1997-1647

ICM G01N021-00; G01N033-86; G06F019-00

AB WO 200101152 A UPAB: 20040115

NOVELTY - Relationship between data is presented by providing data from each of the known **blood** samples; deriving at least one time-dependent measurement on the unknown **blood** sample; transforming data into predicator variable(s); computing **standard** deviation for each predictor variable; determining the z-score for each predictor variable.

DETAILED DESCRIPTION - Presentation of the relationship between data comprises:

- (a) providing data from at least one time dependent measurement profile for each of the known blood samples;
- (b) measuring a respective property over time to derive at least one time-dependent measurement on the unknown blood sample;
- (c) transforming data from (a) and (b) to predicator variable(s) which captures the content of both the unknown and known **blood** sample's **time**-dependent measurement profiles;
- (d) computing standard deviation for each variable in (c) of the known sample of (a); and
- (e) determining the z-score for each predictor variable, and determining if z-score(s) for the unknown sample is greater than a predetermined limit, signifying that the unknown sample is different from the known population represented by the model.

USE - The method is for presenting the relationship between data from an assay relating to thrombosis-hemostasis on an unknown sample, and data from assays relating thrombosis -hemostasis from known populations.

ADVANTAGE - The method facilitates analysis of information embedded in the data from coagulation assays that is not included in the conventional analysis. The additional information discriminates between underlying conditions and aid in the identification of undetected conditions.

DESCRIPTION OF DRAWING(S) - The figure is a diagram illustrating the key aspect of the invention.

Dwg.7/7

FS CPI EPI

FA AB; GI; DCN

MC CPI: B04-C02E1; B04-H19; B11-C08E; B12-K04A2

EPI: S03-E14H

TECH UPTX: 20010220

TECHNOLOGY FOCUS - INSTRUMENTATION AND TESTING - Preferred Method: The data in (c) is transformed into predicator variable(s) that includes timing, rate and/or magnitude of changes during the time -dependent profile. Coagulation assays are performed in (a) and (b) to provide profiles. The unknown sample is removed by an automated probe from the sample container to a test well. Reagent(s) are added to the test well to initiate property changes within the sample. The development of the property over time is monitored to derive the optical data profile. The z-scores is stored in memory of the analyzer and/or displayed on the analyzer. Maps are provided for presenting the data.

Preferred Variable: The predictor variable in (c) includes a minimum(s) of the two derivative of the profile; time index(es) on the minimum of two derivatives; maximum(s) and time index(es) of the second derivative; overall change(s) in the coagulation parameter during the time-dependent measurement on the unknown sample; clotting time(s); and slope(s) of the profile before and after clot formation.

Preferred Sample: The known blood samples are whole

blood or plasma from which information is known relating to intrinsic or extrinsic(s) clotting factors and/or therapeutic agents or are normal samples. Preferred Profile: The time-dependent measurement profile(s) comprise(s) profile from PT, APTT, preferably optical measurements made from multiple wavelengths that correspond to light scattering changes or light absorption in the sample. At least one optical profile is provided with an automated analyzer for thrombosis and hemostasis testing. L62 ANSWER 14 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2001-024512 [03] WPIX DNN N2001-019191 DNC C2001-007324 Detection of coagulation abnormalities, e.q., those associated with autoimmune disorders, by comparison of thrombin generation rates achieved using defibrinated plasma samples. B04 S03 KRILIS, S (SESY-N) SOUTHEASTERN SYDNEY AREA HEALTH SERVICE WO 2000062077 A1 20001019 (200103) * EN 24 G01N033-86 RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL OA PT SD SE SL SZ TZ UG ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY CA CH CN CR CU CZ DE DK DM DZ EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW A 20001114 (200108) AU 2000036486 EP 1173771 A1 20020123 (200214) EN G01N033-86 R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI JP 2002541484 W 20021203 (200309) 28 G01N033-86 <--B 20031127 (200404) AU 768038 G01N033-86 <--WO 2000062077 A1 WO 2000-AU309 20000412; AU 2000036486 A AU 2000-36486 20000412; EP 1173771 A1 EP 2000-915041 20000412, WO 2000-AU309 20000412; JP 2002541484 W JP 2000-611088 20000412, WO 2000-AU309 20000412; AU 768038 B AU 2000-36486 20000412 AU 2000036486 A Based on WO 2000062077; EP 1173771 A1 Based on WO 2000062077; JP 2002541484 W Based on WO 2000062077; AU 768038 B Previous Publ. AU 2000036486, Based on WO 2000062077 PRAI AU 1999-9712 19990412 ICM G01N033-86 ICS C12Q001-56; G01N033-49 WO 200062077 A UPAB: 20010116 NOVELTY - A comparison of thrombin generation rates achieved by reaction of an activator of thrombin with defibrinated normal plasma, in the presence and absence of a defibrinated test plasma sample, is used to detect coagulation abnormalities in the test plasma sample. DETAILED DESCRIPTION - Detection of coagulation abnormalities in a plasma sample comprises: (a) determining a test rate of thrombin generation over a given time interval by reacting an activator of thrombin with defibrinated normal plasma (DNP) in the presence of a defibrinated test plasma (b) determining a control rate of thrombin

defibrinated test plasma; and (c) comparing the rates of thrombin

activator of thrombin with DNP in the absence of any

generation over the same time interval as in (a) by reacting an

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AB

generation in steps (a) and (b). Any significant difference between the two thrombin generation rates is indicative of a coagulation abnormality in the test plasma.

USE - The process is useful for detecting clotting abnormalities in plasma samples. It may be used to detect decreased clotting potential, which can occur as a result of sepsis, severe trauma or autoimmune conditions. It may be used to detect increased clotting potential, which can occur with lupus, other autoimmune conditions or the presence of autoantibodies to beta 2-GPI, prothrombin or other antigens that induce antiphospholipid antibodies to occur. It can be used to identify patients at risk of clinical thrombotic events.

ADVANTAGE - The process is simple to carry out, and can identify both anticardiolipin and lupus **anticoagulant** type antibodies. It can discriminate between infective and autoimmune type anticardiolipin autoantibodies.

Dwg.0/9

FS CPI EPI

FA AB; DCN

MC CPI: B04-B04D4; B04-H19; B11-C08; B11-C08A; B12-K04E

data points on a linear portion of the curve.

EPI: S03-E14H

TECH UPTX: 20010116

TECHNOLOGY FOCUS - PHARMACEUTICALS - Preferred Process: The activator of thrombin is thromboplastin, kaolin, Russell Viper venom or silica, especially thromboplastin. The time interval is 20 minutes. The thrombin generation is measured by adding a substrate which is converted by the generated thrombin to a detectable product. This substrate is especially spectrozyme. Thrombin formation in (a) and (b) is measured in optical density units, and the units obtained are placed on the Y axis and plotted over time (in minutes) on an X axis to give a sigmoidal curve. A shift in the curve to the left indicates accelerated thrombin generation, and a shift to the right indicates inhibition of thrombin formation in the test plasma sample. Changes may be quantified by expressing the results as a ratio of the normal curve using the mean of three

ABEX UPTX: 20010116

EXAMPLE - A typical process used ELISA 96 well plates to which thromboplastin (diluted 1/10 in 0.9% NaCl; 25 microl), test antibody (diluted in 0.9% NaCl; final concentration 6.25 microg/ml; 25 microl) and pooled defibrinated plasma (1/4 dilution in 0.9% NaCl; 50 microl) were added. The mixture was incubated at 37degreesC for 10 minutes. Spectrozyme (diluted 1/5 in 0.9% NaCl; 50 microl) was added at room temperature, followed by CaCl (30 mM in 0.9% NaCl; 50 microl) at room temperature. The time started on the addition of CaCl. The absorbance was read at 405 nm every 2 minutes. A typical curve was a skewed S shape, and the readings were stopped when a plateau was reached at an optical density of 1.2.

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L62 ANSWER 15 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
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AN 2000-663458 [64] WPIX

DNN N2000-491495 DNC C2000-200997

TI Method for diagnosing hemostasis system disorder in circulatory shock patients.

DC B04 S03

IN KNYSH, A S; SHIPAKOV, V E; SHPISMAN, M N; TYUTRIN, I I

PA (KNYS-I) KNYSH A S; (SHIP-I) SHIPAKOV V E; (SHPI-I) SHPISMAN M N; (TYUT-I) TYUTRIN I I; (UYSI-R) UNIV SIBE MED

CYC :

PI RU 2151401 C1 20000620 (200064)* G01N033-86

ADT RU 2151401 C1 RU 1998-114538 19980716

PRAI RU 1998-114538 19980716

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IC
     ICM G01N033-86
     ICS G01N033-49
AB
          2151401 C UPAB: 20001209
     NOVELTY - Method involves determining response period,
     spontaneous blood platelets aggregation intensity,
     prothrombin constant, maximum amplitude, clot
     fibrin-blood-platelet structure formation
     time, general retraction and spontaneous clot lysis
     index. Response period being found to be within 5-7 min, spontaneous
     blood platelets aggregation intensity being from -2 to
     -6 relative units, prothrombin constant being equal to 4-6 min,
     maximum amplitude being within the limits of 500-700 relative units,
     clot fibrin-blood- platelet
     structure formation time being within the limits of 40-60 min,
     general retraction and spontaneous clot lysis index being equal
     to 10-20%, no shock case is to be diagnosed. Spontaneous blood
     platelets aggregation intensity being from -8 to -14 relative
     units and no changes in the other values being observed, compensated shock
     stage is to be diagnosed. Response period being found to be less than 4
     min, spontaneous blood platelets aggregation intensity
     being from -8 to -14 relative units, prothrombin constant being
     less than 3 min, maximum amplitude being greater than 750 relative units,
     clot fibrin-blood-platelet structure
     formation time being less than 40 min, general retraction and
     spontaneous clot lysis index being equal to 10-20%,
     decompensated reversible shock stage is to be diagnosed. Response period
     being found to be greater than 10 min, spontaneous blood
     platelets aggregation intensity being equal to -4 relative units,
     prothrombin constant being greater than 9 min, maximum amplitude
     being less than 400 relative units, clot fibrin-
     blood-platelet structure formation time being
     greater than 70 min, general retraction and spontaneous clot
     lysis index being greater than 20% or less than 5%, decompensated
     irreversible shock stage is to be diagnosed.
     USE - Medicine.
          ADVANTAGE - Enhanced accuracy of the method; accelerated
     method. 1 dwg
     Dwg.0/0
FS
     CPI EPI
FA
     AB
MC
     CPI: B04-B04D5; B11-C08E; B12-K04A
     EPI: S03-E14H; S03-E14H1
L62
    ANSWER 16 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     2000-514997 [46]
                        WPIX
CR
     1997-052574 [05]; 1999-458146 [38]; 2001-091718 [10]; 2001-234924 [24];
     2002-147005 [19]
DNN
    N2000-380594
                        DNC C2000-153705
TI
     Predicting hemostatic dysfunction by measuring property of
     blood sample over time and creating time
     -dependent measurement profile.
DC
     B04 P31 S03
IN
     DOWNEY, C; FISCHER, T J; TOH, C H
PA
     (ALKU) AKZO NOBEL NV
CYC
    24
     WO 2000046603
PΙ
                   A1 20000810 (200046) * EN 111
        RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE
         W: AU CA JP KR US
     AU 2000034833
                    A 20000825 (200059)
                     A1 20011024 (200171)
     EP 1147423
                                          EN
                                                      G01N033-86
         R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE
     JP 2002541431
                   W 20021203 (200309)
                                               105
                                                      G01N033-86
ADT WO 2000046603 A1 WO 2000-US2987 20000204; AU 2000034833 A AU 2000-34833
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20000204; EP 1147423 A1 EP 2000-913371 20000204, WO 2000-US2987 20000204; JP 2002541431 W JP 2000-597634 20000204, WO 2000-US2987 20000204 AU 2000034833 A Based on WO 2000046603; EP 1147423 A1 Based on WO 2000046603; JP 2002541431 W Based on WO 2000046603 PRAI US 1999-244340 19990204 IC ICM G01N033-86 ICS A61B005-145 AB WO 200046603 A UPAB: 20030603 NOVELTY - Predicting hemostatic dysfunction comprises defining predictor variables (110), e.g. the slope of a time-dependent profile. A model is derived (113) that represents the relationship between the dysfunction and the variables from known samples (115). The model is used to predict the existence of hemostatic dysfunction. USE - The predicted hemostatic dysfunction is especially disseminated intravascular coagulation. ADVANTAGE - The prediction can be made from a time -dependent measurement profile without artificial manipulation of samples. DESCRIPTION OF DRAWING(S) - The figure is a chart illustrating the steps of the process. Definition of predictor variables 110 Derivation of model 113 Predictor variables from known samples 115 Dwg.31/45 FS CPI EPI GMPI AB; GI FA CPI: B11-C08E; B12-K04A2 MC EPI: S03-E14H TECH UPTX: 20000921 TECHNOLOGY FOCUS - BIOLOGY - Preferred method: The time -dependent measurement is an optical profile provided by an automated analyzer for thrombosis and hemostatic testing. A number of optical measurements are taken at one or more wavelengths to derive the optical profile, the measurements corresponding to changes in light transmission through the sample. Results of predicted congenital or acquired imbalance or therapeutic condition are automatically stored in a memory of an automated analyzer and/or displayed. Assays for confirming the existence of the condition are automatically performed. The time-dependent measurement profiles are derived from assays initiated with PT, APTT, fibrinogen or TT reagents. Additional patient medical data e.g. fibrinogen, D-dimer or platelet count information, may also be used for predicting the condition. The dysfunction prediction may be performed a number of times to monitor disease progression or regression in the patient. L62 ANSWER 17 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2000-412008 [35] AN WPIX DNN N2000-307978 DNC C2000-124866 TIPerformance of blood coagulation assays with clotting monitored by piezoelectric sensing. DC A96 B04 S03 MORENO, M; WU, J R IN PA (ALKU) AKZO NOBEL NV CYC PΤ A1 20000602 (200035) * EN WO 2000031529 40 G01N033-00 RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE W: AU CA JP KR US AU 2000017331 A 20000613 (200043) G01N033-00 < - -B1 20010313 (200120) US 6200532 G01N033-00 <--EP 1141699 A1 20011010 (200167) G01N033-00 EN R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE ADT WO 2000031529 A1 WO 1999-US27287 19991117; AU 2000017331 A AU 2000-17331

19991117; US 6200532 B1 US 1998-197481 19981120; EP 1141699 A1 EP

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1999-960444 19991117, WO 1999-US27287 19991117
    AU 2000017331 A Based on WO 2000031529; EP 1141699 A1 Based on WO
FDT
     2000031529
PRAI US 1998-197481
                          19981120
IC
     ICM G01N033-00
     WO 200031529 A UPAB: 20000725
AB
     NOVELTY - A reaction chamber (1) in a housing has a blood sample
     inlet. A generator (6) passes electromagnetic waves through the sample in
     the reaction chamber. A piezoelectric device (3) monitors changes to the
     waves after passing through the sample to detect a changing
     coagulation parameter of the sample.
          DETAILED DESCRIPTION - Mechanical vibration is created using a bender
     (2) made of a thin iron film attached to the piezoelectric film (3).
     Variations in the bender movement are detected by the piezoelectric device
     that provides a signal corresponding to the time required for
     the formation of a fibrin clot. An electric circuit
     (7) collects the signal generated by the piezoelectric device. A
     differential amplifier enhances the signal. A separation membrane may be
     used to separate red blood cells from whole blood in
     the event that a plasma sample is desired. The membrane may be
     provided as part of the point-of-care device. A mechanism may be provided
     to compensate for the effect of the different hematocrit content in a
     patient's whole blood sample in a device for measuring one or
     more coagulation parameter.
          USE - The device performs blood coagulation
     assays, particularly prothrombin times,
     activated partial thromboplastin times and
     other clotting tests.
          ADVANTAGE - It is easy to use, accurate and rapid for routine testing
     at a patient's bedside, physician's office, operating room, or patient's
     home for patients undergoing anticoagulant therapy.
          DESCRIPTION OF DRAWING(S) - The figure shows a cross-sectional view
     through the test device.
          reaction chamber 1
          magnetic bender 2
          piezoelectric film 3
          electromagnetic wave generator 6
          electric circuit 7
     Dwg.2/14
FS
     CPI EPI
FΑ
     AB; GI; DCN
MC
     CPI: A12-V03B; B04-B04D5; B04-H19; B11-C08B;
          B12-K04A2
     EPI: S03-E02X; S03-E12; S03-E14H1
L62 ANSWER 18 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1999-561378 [47]
                        WPIX
DNN
    N1999-414814
                        DNC C1999-163562
TI
     Analytical method for qualitative and optionally quantitative
     determination of polymerization and/or coagulation in
     fluids containing polymerizable and/or coagulable
     components.
DC
     A35 B04 D14 D16 J04 S03
     LUNDSTROEM, I; TENGVALL, P; LUNDSTROM, I
IN
     (GLOB-N) GLOBAL HEMOSTASIS INST MGR AB
PA
CYC
                     A1 19990902 (199947) * EN
PT
     WO 9944060
                                                24
                                                      G01N033-49
        RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL
            OA PT SD SE SZ UG ZW
         W: AL AM AT AU AZ BA BB BG BR BY CA CH CN CU CZ DE DK EE ES FI GB GD
            GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV
            MD MG MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT
            UA UG US UZ VN YU ZW
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AU 9930296
                    A 19990915 (200004)
                                                     G01N033-49
                                                                    <--
                    A1 20001213 (200066) EN
                                                     G01N033-49
    EP 1058848
                                                                    <--
        R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE
     JP 2002505429 W 20020219 (200216)
                                               27
                                                     G01N021-27
    US 6379976
                   B1 20020430 (200235)
                                                                    <--
                                                     G01N033-53
                   B1 20020918 (200269) EN
    EP 1058848
                                                     G01N033-49
                                                                    <--
        R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE
    DE 69903002
                   E 20021024 (200278)
                                                     G01N033-49
    WO 9944060 A1 WO 1999-EP1156 19990223; AU 9930296 A AU 1999-30296
ADT
    19990223; EP 1058848 A1 EP 1999-911696 19990223, WO 1999-EP1156 19990223;
    JP 2002505429 W WO 1999-EP1156 19990223, JP 2000-533757 19990223; US
     6379976 B1 WO 1999-EP1156 19990223, US 2000-622063 20001114; EP 1058848 B1
    EP 1999-911696 19990223, WO 1999-EP1156 19990223; DE 69903002 E DE
     1999-603002 19990223, EP 1999-911696 19990223, WO 1999-EP1156 19990223
FDT AU 9930296 A Based on WO 9944060; EP 1058848 A1 Based on WO 9944060; JP
    2002505429 W Based on WO 9944060; US 6379976 B1 Based on WO 9944060; EP
     1058848 B1 Based on WO 9944060; DE 69903002 E Based on EP 1058848, Based
     on WO 9944060
PRAI SE 1998-590
                         19980226
    ICM G01N021-27; G01N033-49; G01N033-53
         G01J003-45; G01N033-00; G01N033-48;
         G01N033-543; G01N033-86
AB
          9944060 A UPAB: 19991116
    NOVELTY - Analytical method for qualitative and optionally
     quantitative determination of polymerization or
     coagulation in fluids containing polymerizable or
     coagulable components involves measuring changes in reflected
    beams of incident beams of electromagnetic radiation.
         DETAILED DESCRIPTION - Analytical method of qualitative and
     optionally quantitative determination of polymerisation or
     coagulation comprises:
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- (a) initiating polymerization or coagulation in fluid;
- (b) contacting fluid or sample with a film of electrically conducting material on a support that is transparent for electromagnetic radiation used and is more optically dense than the fluid;
- (c) directing incident beams of electromagnetic radiation through the support, to the back of the film, at angles equal to or greater than the critical angle for total reflection;
- (d) measuring changes in the reflected beams due to changes in the surface plasmon resonance angle;
 - (e) repeating (c) and (d) at least once;
- (f) registering the occurrence and magnitude of changes measured in (d) and
- (g) correlating the occurrence and magnitude of changes with qualitative and optionally quantitative occurrence of polymerization or coagulation in the fluid.

USE - Used for qualitative and optionally quantitative determination of polymerization or coagulation in fluids containing polymerizable or coagulable components (claimed) including biological fluids, such as blood or blood plasma (claimed) and milk, lymph, sperm, and synovial fluid, and polymerization reaction fluids, including paints, lacquers, glues, thermosetting plastics, thermoplastics, acrylamides, agarose and foodstuffs including sauce, custard, mousse, jelly or sugar (claimed).

The method is also used to determine presence of initiators such as thromboplastin.

A drop of whole **blood** (50 mu l), obtained by skin puncture of a fingertip of an apparently healthy volunteer, was transferred with a plastic pipette tip onto a 45 nm gold film covered with dry immobilized thermoplastin, a **coagulation** inhibitor. The gold film was supported on a glass slide, which was placed on the flat side of the semi-spherical rod prism of a modified BIAlite (RTM) surface plasmon

resonance instrument. The prism and slide with the god film were temperature equilibrated at 22 deg. C. An instrument run, with continuous registrations of shift in surface plasmon resonance angle, was started some minutes prior to applying the drop of **blood**.

The sensogram produced was interpreted as follows. Application of the drop of blood on the gold film resulted in large changes in instrument readout at 180 seconds. The readout then stabilized at 33500 RU. At 240 seconds, the readout began to increase and increased continuously for some minutes, reaching a plateau of 35500 RU at 400 seconds. In total, during the time period 240-400 seconds, the readout increased by 2000 RU. This relatively large increase, a relatively large signal, was caused by coagulation of the blood and was related to the magnitude of coagulation. In separate experiments, the coagulation was confirmed by touching the drop of blood on the gold film with the tip of a plastic pipette. At 240 seconds, the drop of blood was liquid, but at 400 seconds, the drop of blood was coagulated, i.e. transformed into a gel. The results demonstrate that the occurrence of coagulation in whole blood, initiated by thromboplastin, was readily qualitatively and optionally quantitatively determined by the analytical method.

ADVANTAGE - The method places no, or small, requirements on fluid volume, thus fluid volume size and precision in fluid transfer is not of importance for analytical results. The method allows determinations in severe-condition situations e.g. inside high-pressure and -temperature polymerization reactors, such as in the production of polyethylene, and on-line determinations inside flow through industrial reactors and the circulatory system.

DESCRIPTION OF DRAWING(S) - Sensogram of coagulation in a drop of whole blood initiated by immobilized thromboplastin. Instrument response (shift in surface plasmon resonance angle) expressed in resonance units (RU) is plotted against time.

Dwg.1/3

FS CPI EPI

FA AB; GI; DCN

MC CPI: A09-B; A10-B01; A10-C01; A10-D; B04-B04D5; B04-C03B; B05-A01B; B05-A03B; B05-B02C; B11-C01; B11-C08; B12-K04; D03-K03; D03-K04; D05-H09; J04-C

EPI: S03-E14H; S03-E14H1

TECH

UPTX: 19991116

TECHNOLOGY FOCUS - INSTRUMENTATION AND TESTING - Preferred method : (a), (b) and (c) are effected simultaneously or in an alternative order. (e) is effected continuously. Initiation of polymerization or coagulation is spontaneous is effected by changes of one or several conditions within the fluid chosen from temperature, pressure, ionic strength, pH and/or redox potential, or is effected by at least one initiator, preferably attached to the surface of the film in contact with the fluid. Changes in the reflected beams due to changes in the surface plasmon resonance angle are polarization changes in relation to the polarization of incident beams or incident changes. The wavelength of incident light is chosen to coincide with the absorption wavelength(s) of the fluid. Preferred films: The film of electrically conducting material is comprising at least one of gold, silver, platinum, aluminum or electrically conducting polymer. The thickness of the film is 10-1000 nm. Preferred film support: The film support comprises glass or plastic and is

Preferred film support: The film support comprises glass or plastic and is in the form of a plate, half sphere, half-spherical rod, optical fiber, beaker, cuvette, test tube or reactor window.

TECHNOLOGY FOCUS - BIOLOGY - The fluid is a biological fluid, preferably blood or blood plasma, more preferably mixed

gitomer - 10 / 663449 with at least one coagulation inhibitor, especially heparin, hirudin, anti-thrombin, tissue factor pathway inhibitor, C1-inhibitor and/or Ca2+ activity lowering agent and/or at least one coagulation initiator, especially negatively charged surfaces including silica and ellergic acid derivatives, phospholipids, thromboplastin, endothelial cells and cell membranes, thrombocytes and their cell membranes, monocytes and their cell membranes or any required coagulation factor lacking from the fluid. TECHNOLOGY FOCUS - POLYMERS - The fluid is a polymerization reaction fluid, preferably a paint, lacquer, qlue, thermosetting plastic, thermoplastic, acrylamide, agarose, foodstuff including sauce, custard, mousse, jelly or sugar. L62 ANSWER 19 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1999-458146 [38] WPIX 1997-052574 [05]; 2000-514997 [46]; 2001-091718 [10]; 2001-234924 [24]; 2002-147005 [19] N1999-342695 DNC C1999-134481 Predicting presence of abnormal levels of protein in blood clotting cascade. B04 S03 T01 BRAUN, P; FISCHER, T J; GIVENS, T B (ALKU) AKZO NOBEL NV; (BRAU-I) BRAUN P; (FISC-I) FISCHER T J; (GIVE-I) GIVENS T B; (INMR) BIOMERIEUX SA

CR DNN TIDC IN PA CYC WO 9934208 A1 19990708 (199938) * EN 94 PΤ G01N033-49 RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE W: AU CA JP KR US AU 9919503 A 19990719 (199951) EP 1042669 A1 20001011 (200052) EN R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE KR 2001033817 A 20010425 (200164) G01N033-49 B1 20011120 (200174) US 6321164 G01N021-00 JP 2002500360 W 20020108 (200206) 82 G01N033-68 US 2002019706 A1 20020214 (200214) G06F019-00 US 6564153 B2 20030513 (200335) G01N021-00 ADT WO 9934208 A1 WO 1998-US27865 19981230; AU 9919503 A AU 1999-19503

19981230; EP 1042669 A1 EP 1998-964342 19981230, WO 1998-US27865 19981230; KR 2001033817 A KR 2000-707373 20000630; US 6321164 B1 Cont of US 1995-477839 19950607, CIP of US 1997-859773 19970521, US 1997-1647 19971231; JP 2002500360 W WO 1998-US27865 19981230, JP 2000-526808 19981230; US 2002019706 A1 Cont of US 1995-477839 19950607, CIP of US 1997-859773 19970521, Cont of US 1997-1647 19971231, US 2001-850255 20010507; US 6564153 B2 Cont of US 1995-477839 19950607, CIP of US 1997-859773 19970521, Cont of US 1997-1647 19971231, US 2001-850255 20010507

FDT AU 9919503 A Based on WO 9934208; EP 1042669 A1 Based on WO 9934208; US 6321164 B1 Cont of US 5708591, CIP of US 6101449; JP 2002500360 W Based on WO 9934208; US 2002019706 Al Cont of US 5708591, CIP of US 6101449, Cont of US 6321164; US 6564153 B2 Cont of US 5708591, CIP of US 6101449, Cont of US 6321164

PRAI US 1997-1647 19971231; US 1995-477839 19950607; 19970521; US 2001-850255 US 1997-859773 20010507

IC ICM G01N021-00; G01N033-49; G01N033-68; G06F019-00 ICS G01N031-00; G01N033-86

AN

AB 9934208 A UPAB: 20030603 NOVELTY - Predicting the presence of an abnormal level of at least 1 protein in the clotting cascade from at least one time -dependent measurement profile involves using a model that represents the relationship between the abnormal level of the protein in the clotting cascade and a set of predictor variables.

DETAILED DESCRIPTION - Predicting the presence of an abnormal level of at least 1 protein in the clotting cascade from at least one time-dependent measurement profile comprises:

- (1) performing at least one time-dependent measurement on an unknown sample and measuring a respective property over time to derive a time-dependent measurement profile;
- (2) defining a set of predictor variables which define the data of the time-dependent measurement profile;
- (3) deriving a model that represents the relationship between the abnormal level of the protein in the clotting cascade and the set of predictor variables and
- (4) using the model of step (c) to predict the existence of the abnormal level of the protein in the clotting cascade and to predict which protein or proteins in the clotting cascade are at an abnormal level.

The prediction of the protein or proteins at an abnormal level is a better prediction than an abnormal **clot time** alone.

An INDEPENDENT CLAIM is also included for presenting a relationship between data from an assay relating to thrombosis-hemostasis on an unknown sample and data from assays relating to thrombosis-hemostasis from known sample populations which comprises:

- (A) providing data from at least one time dependent measurement profile for each of known blood samples;
- (B) performing at least one time-dependent measurement on an unknown blood sample and measuring a respective property overtime to derive at least one time-dependent measurement profile for the unknown blood sample;
- (C) transforming data from step (2) to predictor variables which capture the information content of both the unknown **blood** sample time-dependent measurement profile and the known **blood** sample time-dependent measurement profiles and
- (D) presenting the data from the unknown **blood** sample time-dependent measurement profile relative to the data from the known **blood** sample **time**-dependent measurement profiles.

USE - Used for estimating the concentration of at least one protein in the clotting cascade.

Dwg.0/30

FS CPI EPI

FA AB; DCN

MC CPI: B04-B04D2; B04-N04; B11-C07; B12-K04

EPI: S03-E14H; S03-E14H1; T01-J

TECH

UPTX: 19990922

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred method: In the prediction of the protein at an abnormal level, the specificity is at least 0.85 and the sensitivity is greater than 0.6. The samples with a measured concentration of less than 30% of normal for a specific factor are defined as being at an abnormal level. The time-dependent measurement profile is at least one optical profile is produced by an automated analyzer for thrombosis and hemostasis testing. A number of optical measurements at one or more wavelengths are taken over time to derive at least one optical profile. The optical measurements correspond to changes in light scattering and/or light absorption in the unknown sample. The optical measurements are taken over time to derive at least one optical profile, and the optical measurements are each normalized to a first optical measurement. At least one optical profile is produced automatically by an analyzer. The unknown sample is automatically removed by an automated probe from a sample container to a test well. One or more reagents are automatically added to the test well to initiate the property changes within the sample. The development of the property over time is automatically optically monitored. A predicted congenital or acquired imbalance or therapeutic condition is

automatically stored in a memory of the automated analyzer and/or displayed on the automated analyzer. Assays for confirming the existence of the congenital or acquired imbalance or therapeutic condition are automatically effected. A set of data from known samples is produced which is used for deriving the model. The data from known samples is obtained by performing assays on the known samples. Time dependent measurement profiles include at least two profiles from assays initiated with PT reagents, APTT reagents, fibrinogen reagents and TT reagents. In (1), steps (3) and (4) comprise transforming a set of input parameters from the time-dependent measurement profiles for the known blood samples and the unknown blood sample, to corresponding individual output neurons whose location on an output map corresponds to the respective input data. Step (4) also comprises: selecting weight vectors; (2) selecting a sample from a training set; (3) identifying best matching winning neuron at a particular time (4) updating weight vectors and (5) repeating steps (1)-(4) until the map reaches equilibrium. In step (3), data from the time-dependent measurement profiles is transformed into predictor variables that characterize timing, rate and magnitude of changes during the time-dependent measurement profile and the predictor variables are used as input for neural networks. The definition of the predictor variables is a position in a self-organizing feature map, trained with data from the time -dependent measurement profiles for the known blood samples. Preferred materials: The proteins comprise factors II, V, VII, VIII, IX, X, XI and/or XII. The known blood samples and the unknown blood sample are samples of whole blood, plasma , or other part of whole blood. The known blood samples are samples of which are information is known relating to one or more intrinsic or extrinsic clotting factors and/or therapeutic agents. The known blood samples are samples of which are known the presence or absence of one or more abnormalities relating to at least one of fibrinogen level, oral anticoagulant, heparin, and one or more factor levels. In step (4) one or more of normal sample, presence of heparin, and one or more factor deficiencies are presented on a PT map, or at least one of normal specimen, presence of heparin, abnormal fibrinogen, oral anticoagulant, and one or more factor deficiencies are presented on an APTT map. The predictor variables are in terms of a standard deviation from a mean of at least one known blood sample population, and the unknown blood sample is characterized by variation from the mean of the known blood samples for each predictor variable. UPTX: 19990922 EXAMPLE - No relevant example given. L62 ANSWER 20 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1999-373178 [32] WPIX DNN N1999-278551 DNC C1999-110250 Detecting haemostasis defects, using clotting agent and/or anticoagulating agent. B04 D16 S03 KRAUS, M; SCHELP, C; WIEGAND, A (DADE-N) DADE BEHRING MARBURG GMBH CYC 29 EP 924523 A2 19990623 (199932) * GE 17 G01N033-86

R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT

G01N033-50 G01N033-50

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ABEX

AN

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DC

IN

PA

PΤ

RO SE SI

DE 19756773 A1 19990624 (199932) AU 9897182 A 19990708 (199938)

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                   A 19990907 (199947)
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                                                      G01N033-86
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                   A1 19990619 (199949) EN
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     US 2003027235 A1 20030206 (200313)
                                                      C12Q001-56
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    US 6750032
                    B2 20040615 (200439)
                                                                     <--
                                                      C12Q001-52
ADT EP 924523 A2 EP 1998-122888 19981202; DE 19756773 A1 DE 1997-1056773
     19971219; AU 9897182 A AU 1998-97182 19981217; JP 11242035 A JP
     1998-360662 19981218; CA 2256089 A1 CA 1998-2256089 19981215; US 6187594
     B1 US 1998-215167 19981218; US 6482653 B1 Cont of US 1998-215167 19981218,
     US 2000-604271 20000626; US 2003027235 A1 Cont of US 1998-215167 19981218,
     Div ex US 2000-604271 20000626, US 2002-255632 20020927; US 6750032 B2
     Cont of US 1998-215167 19981218, Div ex US 2000-604271 20000626, US
     2002-255632 20020927
    US 6482653 B1 Cont of US 6187594; US 2003027235 A1 Cont of US 6187594, Div
FDT
     ex US 6482653; US 6750032 B2 Cont of US 6187594, Div ex US 6482653
PRAI DE 1997-19756773
                          19971219
     ICM C12Q001-52; C12Q001-56; G01N033-50;
          G01N033-86
     ICS
         G01N021-75; G01N021-76; G01N021-77; G01N033-546
AB
           924523 A UPAB: 19991122
     NOVELTY - Detecting haemostasis defects comprises adding a
     clotting agent and/or an anticoagulating agent at
     time intervals which causes or inhibits an energy transfer between
     the agents, which can be measured.
          DETAILED DESCRIPTION - An INDEPENDENT CLAIM is also included for a
     diagnostic composition comprising the blood clotting
     substances and anticoagulating substances used in the
     method above.
         USE - The method can be used to detect genetic and/or
     acquired lack of blood clotting factors or factors of
     the fibrinolytic system, genetic defects of thrombocytes,
     disease-associated or disease therapy-associated haemostasis
     defects and/or genetic and/or acquired defects of the complement system.
     The method can be used to establish the lack of certain factors
     necessary for blood clot formation, and for testing
     the aggregation ability of platelets (all claimed).
     Dwg.0/3
     CPI EPI
FS
     AB; DCN
FA
MC
     CPI: B04-B01B; B04-C02C; B04-C03; B04-N04; B05-B01B; B05-B02C; B05-U02;
          B06-A03; B06-D18; B06-F02; B10-F02; B11-C07B3; B11-C08E3; B12-K04A;
          D05-H09
     EPI: S03-E14H
TECH
                    UPTX: 19990813
     TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred materials: The energy
     transfer is caused by short-lived molecules such as singlet oxygen,
     radiation with low intensity, e.g. radioactive beta-radiation and/or
     energy transfer according to Foerster. The activity of the agents can be
     enhanced or inhibited by other substances, which will change the
     measurable signal strength by changing the polarity and intensity of the
     light, inhibiting or enhancing of enzyme activity and/or changing
     fluorescence. The agents are capable to bind to the blood
     clot via direct or indirect non-specific hydrophobic or
     electrostatic exchange. The agents are bound to or are encased in
     suspendable particles via covalent binding, specific binding and
     adsorption. The particles are selected from colored crystals, alloys,
     silicas, magnet particles, oil droplets, lipid particles, dextran, protein
     aggregates and, especially latex particles. The particles can be further
     treated after their initial preparation and can comprise adsorptively or
     covalently bound coating layers or shells, comprising proteins,
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carbohydrates, lipophilic substances, bio-polymers and/or

organic polymers. The method also measures the

blood clotting time, partial thromboblast time, thromboblast time, protein C activation time, Russel viper venom time and thrombin time. The diagnostic substances used in the method also comprises factors of the blood clotting reaction.

TECHNOLOGY FOCUS - ORGANIC CHEMISTRY - Preferred Agent: The agents are selected from photosensitising and chemiluminescent compounds which can be activated to generate singlet oxygen and light radiation, respectively. The photosensitising compounds are acetone, benzophenone, 9-thioxanthone, eosin, 9,10-dibromoanthrazene, chlorophyll, buckminsterfullerene, methylene blue, rose Bengal, porphyrines, phthalocyanine and/or their derivatives. The chemiluminescent compounds are selected from olefins, 9-alkylidenexanthane, 9-alkylidene-N-alkylacridane, enolether, enamine, arylvinylether, dioxene, arylimidazone and/or lucigenin. The chemiluminescent compounds are brought into contact with fluorophores to enhance emission of light of higher wavelength. Fluorescent compounds, e.g. 1,3-di-phenylisobenzofuran, which react with singlet oxygen by photobleaching, or, e.g. oxenumbelliferylether, which react with singlet oxygen to form fluorophore precursors.

ABEX UPTX: 19990813

EXAMPLE - None given.

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L62 ANSWER 21 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
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AN 1999-359879 [31] WPIX

DNC C1999-106698

TI Determination of antithrombin III in plasma sample, useful for diagnosing patients at risk of thrombosis.

DC B04 D16

IN TRISCOTT, M X

PA (SIGM-N) SIGMA-ALDRICH CO; (TRIN-N) TRINITY BIOTECH MFG LTD

CYC 29

PI EP 927767 A2 19990707 (199931)* EN 24 C12Q001-56 <-R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
RO SE SI

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A 19990701 (199937)
A 19990831 (199946)
AU 9895156
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JP 11235197
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                A1 19990609 (199948) EN
CA 2255255
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US 5985582
                A 19991116 (200001)
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AU 730231
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                                                   C12Q001-56
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                A 20010329 (200124)#
AU 2000072346
                                                   G01N033-96
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                B 20020214 (200223)#
AU 744025
                                                   G01N033-96
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ADT EP 927767 A2 EP 1998-310023 19981208; AU 9895156 A AU 1998-95156 19981202; JP 11235197 A JP 1998-349902 19981209; CA 2255255 A1 CA 1998-2255255 19981208; US 5985582 A US 1997-987038 19971209; AU 730231 B AU 1998-95156 19981202; AU 2000072346 A Div ex AU 1998-95156 19981202, AU 2000-72346 20001218; AU 744025 B Div ex AU 1998-95156 19981202, AU 2000-72346 20001218

FDT AU 730231 B Previous Publ. AU 9895156; AU 2000072346 A Div ex AU 730231; AU 744025 B Previous Publ. AU 2000072346, Div ex AU 730231

PRAI US 1997-987038 19971209; AU 2000-72346 20001218

IC ICM C12Q001-56; G01N033-86; G01N033-96 ICS C08B037-10; C12P019-04; C12Q001-44; G01N021-78;

AB EP 927767 A UPAB: 20040408

G01N033-50

NOVELTY - Determination of antithrombin III in a plasma sample comprises, combining the plasma sample with exogenous thrombin and with a heparin derivative (prepared by enzymatically digesting heparin) to form an assay mixture, forming a complex between the antithrombin III and the thrombin, determining the uncomplexed thrombin, and correlating the determined uncomplexed thrombin to the antithrombin III.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

- (1) determination of antithrombin III in a plasma sample comprising, preparing an assay mixture comprising the plasma sample, exogenous thrombin, and a heparin derivative, (effective for enhancing formation of a thrombin-antithrombin III complex and less effective than unmodified heparin for enhancing heparin cofactor II activity against thrombin), incubating the assay mixture, determining the uncomplexed thrombin in the incubated assay mixture and correlating the determined uncomplexed thrombin to the antithrombin III in the plasma sample;
- (2) determination of antithrombin III in a plasma sample containing endogenous heparin cofactor II comprising, preparing an assay mixture comprising the plasma sample, exogenous thrombin and a heparin derivative, incubating the assay mixture, determining the uncomplexed thrombin in the incubated assay mixture and correlating the determined uncomplexed thrombin to antithrombin III in the plasma sample, where the endogenous heparin cofactor II contributes about 15% or less to the determined inhibition of thrombin by antithrombin III;
- (3) a modified heparin comprising a heparin derivative effective for enhancing the **antithrombin** activity against **thrombin** and being less effective than unmodified heparin for enhancing heparin cofactor II activity against **thrombin**;
- (4) the preparation of a heparin derivative comprising enzymatically digesting heparin with chondroitinase;
- (5) a reagent useful in a thrombin-based, antithrombin III assay comprising, a lyophilized composition comprising thrombin and a heparin derivative (as above);
- (6) a kit for a thrombin based antithrombin III assay, useful for determining antithrombin III in a plasma sample comprising, a diluent composition comprising chondroitinase ACI-treated heparin and an alkali metal-halide salt, a reagent composition comprising chondroitinase ACI-treated heparin, an alkali metal-halide salt and thrombin and a chromogenic thrombin substrate;
- (7) the preparation of a high-calibrator reference plasma suitable for use as a standard for the determination of a plasma constituent comprising, obtaining a normal reference plasma, lyophilizing a volume, V1, of the normal reference plasma, and reconstituting the lyophilized normal reference plasma to form a reconstituted plasma of volume, V2, where the ratio of V1:V2 ranges from 7:8 to 1:8; and
- (8) a reference plasma suitable for use as a standard for determination of a plasma constituent, the reference plasma comprising antithrombin III at more than 120% of the normal.
- USE The method is useful for evaluating the hemostasis of a patient of developing thrombosis, and especially, to diagnostic assays for determining the level of antithrombin III present in a plasma sample from a patient. The method is particularly employed with patients known to be or suspected of being at risk of thrombosis and especially those with an antithrombin deficiency e.g. genetic deficiencies (especially, Type I and Type II), and acquired deficiencies such as those occurring in consumptive coagulopathies (especially, DVT, DIC, pulmonary emboli), other disease states (especially, severe liver disease, nephrotic syndrome), surgery, preganancy, trauma and certain courses of therapy (e.g. L-asparaginase).

ADVANTAGE - The antithrombin III assay is not influenced by the antithrombin activity of heparin cofactor II, which can be

suitably performed with automated analyzers, and which are simpler to perform, maintaining sensitivity, accuracy, reproducibility and is relatively inexpensive (especially without using expensive and unstable reagents).

Dwg.0/7

FS CPI

FA AB; DCN

MC CPI: B04-B04D4; B04-C02E1; B04-H19; B04-L05; B04-N02;

B11-C08E3; B12-K04A; D05-A02C; D05-C08; D05-H09

TECH UPTX: 19990806

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: The (modified) heparin derivative is prepared by enzymatically digesting heparin to form a (modified) heparin derivative and at least one (especially unsaturated) disaccharide, and the assay mixture is formed by combining the plasma sample with thrombin and with the modified heparin composition. The heparin derivative is prepared by enzymatically digesting heparin with a chondroitinase (especially chondroitinase ACI-digested heparin). The heparin derivative is at least two (especially 5) times less effective than unmodified heparin for enhancing heparin cofactor II activity against thrombin. The endogenous heparin cofactor II contributes about no more than 10 (especially 5) % to the determined inhibition of thrombin by antithrombin III. The method (7), has a ratio of V2:V1 of 3:4 - 1:4 (especially 2:3). The normal reference plasma comprises antithrombin III at a concentration of 90-110% of normal and at least 120 (especially 140) % of normal, reconstituted plasma comprising antithrombin III.

Preferred Reagent: The lyophilized composition comprises thrombin and a modified heparin composition prepared by enzymatically digesting heparin, especially by lyophilizing a reagent solution comprising 8-96 IU/ml thrombin and 0.5-6 U/ml heparin derivative.

ABEX UPTX: 19990806

EXAMPLE - ATIII assays were performed on each HCII-normal samples with protocols involving the heparin derivative. The HCII samples were diluted 1/40 in the heparin derivative diluent composition (25 mul plasma sample and 975 mul diluent composition), and equilibrated to 37 degreesC. 200 mul Of this mixture was mixed with 200 mul of the heparin-derivative-thrombin reagent composition and was also equilibrated to 37 degreesC to form an assay mixture. The assay mixture which included 175 mM sodium chloride was incubated for 2 minutes at 37 degreesC. 200 mul Of the chromogenic thrombin substrate was added to the incubated assay mixture and allowed to react with the uncomplexed thrombin for exactly 1 or 2 minutes, depending on the relative concentrations of thrombin and substrate, to form a developed assay mixture. The reactions were stopped using 200 mul glacial acetic acid or 2% citric acid. The absorbance of the developed assay mixture was read at OD405 using a spectrophotometer. A standard curve was prepared.

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L62 ANSWER 22 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN AN 1999-279597 [24] WPIX
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PI EP 915340 A1 19990512 (199924)* GE 18 G01N033-86 <--
R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
RO SE SI
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DE 19749197 A1 19990512 (199925) C12Q001-56 <--
AU 9891397 A 19990527 (199932) C12Q001-56 <--
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DNN N1999-209694 DNC C1999-082276

TI Determination of anti-coagulation potential of a sample.

DC B04 D16 S03

IN KRAUS, M

PA (DADE-N) DADE BEHRING MARBURG GMBH; (KRAU-I) KRAUS M

CYC 29

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CA 2252983
                     A1 19990507 (199942) EN
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     AU 751525
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     US 2003207343 A1 20031106 (200374)
                                                      C12Q001-56
                                                                     <--
ADT EP 915340 A1 EP 1998-118971 19981007; DE 19749197 A1 DE 1997-1049197
     19971107; AU 9891397 A AU 1998-91397 19981106; CA 2252983 A1 CA
     1998-2252983 19981106; JP 11225796 A JP 1998-315511 19981106; US
     2002019021 A1 US 1998-187035 19981106; AU 751525 B AU 1998-91397 19981106;
     US 2003207343 A1 Div ex US 1998-187035 19981106, US 2003-419104 20030421
FDT AU 751525 B Previous Publ. AU 9891397
PRAI DE 1997-19749197
                          19971107
IC
     ICM C12Q001-56; G01N033-86
     ICS C07K014-00; C07K016-00; C08B037-10; C12N009-74; G01N033-15
ICA G01N033-50
          915340 A UPAB: 19990624
ΔR
     NOVELTY - Determination of anti-coagulation potential of a
     sample is carried out in the presence of exogeneous thrombomodulin
          DETAILED DESCRIPTION - Determination of anti-coaqulation
     potential of a sample in the presence of an exogenous
     thrombomodulin comprises: (a) adding to the sample exogenous
     thrombomodulin, which can form a protein C-
     activating-complex with thrombin, whereby the
     protein C can be exogenous or endogenous, an
     activator for thrombin, where the prothrombin
     is either endogenous or exogenous, phospholipids, calcium ions
     and other reagents for the optimization of the coagulation
     time test; (b) starting the prothrombin-
     activator reaction; and (c) determining the formation of
     thrombin by either measuring the time of
     thrombin formation, fibrin clot formation or
     formation of labeled thrombin. INDEPENDENT CLAIMS are also
     included for: (1) a test kit used to carry out the method above;
     and (2) a method for determining anti-thrombin III
     activity of the protein C system of a sample,
     comprising the step described above.
          USE - The method is used for the selective diagnosis of
     protein defects or protein lack in a patients sample (all claimed).
FS
     CPI EPI
FΑ
     AB; DCN
MC
     CPI: B04-H19; B04-N02; B11-C08E; B12-K04A2;
          D05-H09
     EPI: S03-E14H1; S03-E14H5; S03-E14H6
TECH
                    UPTX: 19990624
     TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: The
     thrombin formation is evaluated in contrast to thrombin
     formation without the addition of protein C to the
     sample; or by adding a test substance instead of thrombomodulin.
     The thrombomodulin used also promotes the inhibition of
     thrombin by anti-thrombin III. The concentration of the
     activator is such that coagulation takes place within
     20-300 (especially 30-150) s. The activators are common
     activators used as derived from placenta, lung or brain. The
     thrombinmodulin, derived from human or animal sources is added
     separately from the activator to the sample. The
     thrombin inhibition caused by thrombomodulin can be
     reversed by binding of glycosamino glycan, especially heparin sulfate.
L62 ANSWER 23 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1999-095346 [08] WPIX
DNN N1999-069309
                       DNC C1999-028092
ΤI
    Determining enzymatic activity of blood coagulation
```

factor XIII - by detecting degree of fibrin cross-linking formed by blood coagulation factor XIII using purified fibrin monomer. DC B04 D16 S03 IN CHANG, S; CHUNG, S; HUH, J; KIM, H; LEE, J; SEONG, H; CHUNG, ; SEONG, ; CHANG, S J; CHUNG, S G; HUH, J W; KIM, H C; LEE, J S; SEONG, H M (GREC) KOREA GREEN CROSS CORP PA CYC 22 A1 19981223 (199908) * EN PΤ WO 9858078 33 C120001-56 RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE W: JP US A1 19991117 (199953) EN EP 956363 C12Q001-56 R: AT CH DE FR GB LI SE KR 99002006 A 19990115 (200011) G01N033-49 W JP 2000501295 20000208 (200018) 20 C12Q001-56 B1 19991001 (200108) KR 222292 G01N033-49 <--JP 3193728 B2 20010730 (200146) 8 C12Q001-56 <--US 6406874 B1 20020618 (200244) C12Q001-56 <--EP 956363 B1 20031001 (200365) EN C12Q001-56 <--R: AT CH DE FR GB LI SE DE 69818639 E 20031106 (200381) C12Q001-56 WO 9858078 A1 WO 1998-KR160 19980616; EP 956363 A1 EP 1998-929875 ADT 19980616, WO 1998-KR160 19980616; KR 99002006 A KR 1997-25516 19970618; JP 2000501295 W WO 1998-KR160 19980616, JP 1999-504191 19980616; KR 222292 B1 KR 1997-25516 19970618; JP 3193728 B2 WO 1998-KR160 19980616, JP 1999-504191 19980616; US 6406874 B1 WO 1998-KR160 19980616, US 1999-242436 19990217; EP 956363 B1 EP 1998-929875 19980616, WO 1998-KR160 19980616; DE 69818639 E DE 1998-618639 19980616, EP 1998-929875 19980616, WO 1998-KR160 19980616 FDT EP 956363 A1 Based on WO 9858078; JP 2000501295 W Based on WO 9858078; JP 3193728 B2 Previous Publ. JP 200001295, Based on WO 9858078; US 6406874 B1 Based on WO 9858078; EP 956363 B1 Based on WO 9858078; DE 69818639 E Based on EP 956363, Based on WO 9858078 PRAI KR 1997-25516 19970618 IC ICM C12Q001-56; G01N033-49 ICS C12Q001-37; C12Q001-48; G01N033-573; G01N033-72; G01N033-86 AB 9858078 A UPAB: 19990224 Determining the enzymatic activity of blood coagulation factor XIII by detecting the degree of fibrin cross-linking formed by the blood coagulation factor XIII using purified fibrin monomer as a substrate which is free of contaminating blood coagulation factor XIII. USE - The method is especially used for determining the activity of transglutaminase (claimed). Dwg.0/5 FS CPI EPI FA AB MC CPI: B04-B04D3; B04-B04D5; B04-H19; B04-L04; B11-C08E; B12-K04A; D05-H09 EPI: S03-E14H; S03-E14H4 L62 ANSWER 24 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1999-036163 [04] AN WPIX DNN N1999-027154 DNC C1999-011034 Diagnosis of thrombophilia - by determining ΤI anticoagulatory activity of thrombin. DC B04 D16 J04 S03 IN HUND, S; MUELLER-BERGHAUS, G; POETZSCH, B PA (PLAC) MAX PLANCK GES FOERDERUNG WISSENSCHAFTEN; (KERC-N) KERCKHOFF-KLINIK GMBH CYC 22 7 PΙ DE 19724239 A1 19981210 (199904)* G01N033-50 <--

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A1 19981217 (199905) GE
     WO 9857178
                                                      G01N033-86
                                                                     <--
        RW: AT BE CH CY DE DK ES FI FR GB GR IE IT LU MC NL PT SE
        W: CA JP US
                     A1 20000405 (200021) GE
     EP 990157
                                                      G01N033-86
                                                                     <--
        R: AT CH DE ES FR GB IT LI NL
    DE 19724239 A1 DE 1997-1024239 19970609; WO 9857178 A1 WO 1998-EP3407
ADT
     19980608; EP 990157 A1 EP 1998-934924 19980608, WO 1998-EP3407 19980608
    EP 990157 A1 Based on WO 9857178
FDT
PRAI DE 1997-19724239
                         19970609
     ICM G01N033-50; G01N033-86
         C12Q001-56; G01N033-53; G01N033-577;
     ICS
          G01N033-68
     DE 19724239 A UPAB: 19990127
AB
    Diagnosing thrombophilia [congenital predisposition to
     thrombosis] comprises determining the anticoagulatory
     activity of thrombin in a blood or plasma
     sample.
          ADVANTAGE - In some patients clinically diagnosed as
     thrombophilic [9 out of 57 in an example], interaction of
     thrombin-thrombomodulin complex with protein
     C results in less activated protein C
     (APC) formation than would be expected from the amount of thrombin
     present.
     Dwq.0/2
     CPI EPI
FS
FA
    AB
MC
     CPI: B04-B04D4; B04-H19; B04-N02; B11-C08;
          B12-K04A2; D05-H09; J04-B01
     EPI: S03-E14H1; S03-E14H4
L62
    ANSWER 25 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1998-448979 [39]
                        WPIX
DNN
    N1998-350152
                        DNC C1998-136190
    Evaluation of reaction kinetics - using modified threshold method
TI
    B04 D16 S03
DC
IN
    MEYERS, W
     (DADE-N) DADE BEHRING MARBURG GMBH; (BEHW) BEHRING DIAGNOSTICS GMBH
PΑ
CYC
    27
                     A1 19980902 (199839)* GE
                                                15
PΤ
     EP 861687
                                                      B01J019-00
         R: AL AT BE CH DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO
            SE SI
     DE 19707897
                     A1 19980910 (199842)
                                                      G01N037-00
     JP 10253636
                     A 19980925 (199849)
                                                12
                                                      G01N033-86
                                                                      <--
                     A 19980827 (199903)
     CA 2230546
                                                      C12Q001-00
                                                                      <--
     US 6245569
                     B1 20010612 (200135)
                                                      G01N033-00
                                                                      <---
    EP 861687 A1 EP 1998-101330 19980127; DE 19707897 A1 DE 1997-1007897
ADT
     19970227; JP 10253636 A JP 1998-45535 19980226; CA 2230546 A CA
     1998-2230546 19980226; US 6245569 B1 US 1998-30887 19980226
PRAI DE 1997-19707897
                          19970227
    ICM B01J019-00; C12Q001-00; G01N033-00;
          G01N033-86; G01N037-00
          C12M001-34; C12Q001-56; G01N021-27; G01N021-75; G01N031-00;
          G01N033-49; G01N033-50; G01N033-557;
          G06F019-00
AB
     EΡ
           861687 A UPAB: 19981028
      Method for evaluating reaction kinetics comprises: determining
     the variation in a reaction-dependent parameter and measuring the
     time (T) required for the parameter to exceed a limiting value
     (d); establishing reaction-specific values for an initial value d0, a
     maximum value dmax (>d0) and a step number n, thereby establishing a step
     width ds = (dmax-d0)/n; for the region d = d0 to dmax, determining the
     value Tn for each dn = (d0+(ds)*n); from the series of values T0 to Tmax,
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determining the differences Di = Ti-Ti-1 for i = 1, ..., n; from the values D1,..n, determining the highest value Dmax and the lowest value Dmin and calculating the quotient Q = Dmax/Dmin; and using only those measurements for which Q at most Q0 for further evaluation. USE - For determining blood clotting parameters, especially prothrombin time. Dwq.0/14 CPI EPI FS FA AB MC CPI: B04-B04D5; B04-H19; B11-C08; B12-K04A; D05-H09 EPI: S03-E04A1; S03-E14H; S03-E14H1 L62 ANSWER 26 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1998-011964 [02] AN WPIX DNN N1998-009444 DNC C1998-004360 TT Measuring thrombin activity for aggregation reagent for assay of blood coagulation factor - involves supplying insoluble carrier with fibrinogen to examined object. DC B04 D16 S03 (TOKU) TOKUYAMA SODA KK PA CYC A 19971014 (199802)* PΤ JP 09266798 5 C12Q001-56 ADT JP 09266798 A JP 1996-77301 19960329 PRAI JP 1996-77301 19960329 ICM C12Q001-56 IC ICS G01N033-50; G01N033-86 AB JP 09266798 A UPAB: 19980112 Measuring thrombin activity for aggregation reagent for assay of blood coagulation factor involves supplying an insoluble carrier with fibrinogen to an examined object. Then thrombin activity is determined by measuring the amount of aggregation of the carrier. ADVANTAGE - Thrombin activity measurement is reduced. Dwg.1/3 CPI EPI FS FΑ AB; GI MC CPI: B04-B04D5; D05-H09 EPI: S03-E14H ANSWER 27 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN L62 1998-009937 [02] AN WPTX DNN N1998-007727 DNC C1998-003717 TΤ Method for performing blood tests - on blood samples treated with thrombin inhibitor, useful to perform wide rage of tests. B04 D16 S03 DC MELBER, K; MENSSEN, H D; STRASSER, A W M; THIEL, E; STRASSER, A W IN (RHEI-N) RHEIN BIOTECH GES NEUE BIOTECHNOLOGISCHE; (UYFR-N) UNIV FRANKLIN PA BENJAMIN; (UYBE-N) UNIV BERLIN FREIE; (MENS-I) MENSSEN H D; (THIE-I) THIEL CYC 67 DE 19620443 PΙ A1 19971127 (199802) * 9 G01N033-50 <--WO 9744661 A1 19971127 (199802) GE 33 G01N033-50 <--RW: AT BE CH DE DK EA ES FI FR GB GH GR IE IT KE LS LU MC MW NL OA PT SD SE SZ UG W: AL AU BA BB BG BR CA CN CU CZ EE GE HU IL IS JP KP KR LC LK LR LT LV MG MK MN MX NO NZ PL RO SG SI SK TR TT UA US UZ VN AU 9728936 A 19971209 (199824) G01N033-50 DE 19620443 C2 19980917 (199841) G01N033-50 <--EP 912891 A1 19990506 (199922) GE G01N033-50 <--R: AT BE CH DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE W 19991012 (199954) JP 11511856 26 G01N033-86

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B1 20021016 (200276) GE
     EP 912891
                                                      G01N033-50
         R: AT BE CH DE DK ES FI FR GB GR IE IT LI LU MC NL PT SE
                   G 20021121 (200277)
     DE 59708506
                                                      G01N033-50
                                                                      <--
     US 6521460
                     B1 20030218 (200317)
                                                      G01N033-86
                                                                      <--
     ES 2185937
                     T3 20030501 (200341)
                                                      G01N033-50
                                                                      <--
     JP 3523883
                     B2 20040426 (200428)
                                                11
                                                      G01N033-86
ADT
    DE 19620443 A1 DE 1996-1020443 19960521; WO 9744661 A1 WO 1997-EP2343
     19970507; AU 9728936 A AU 1997-28936 19970507; DE 19620443 C2 DE
     1996-1020443 19960521; EP 912891 A1 EP 1997-923007 19970507, WO
     1997-EP2343 19970507; JP 11511856 W JP 1997-540286 19970507, WO
     1997-EP2343 19970507; EP 912891 B1 EP 1997-923007 19970507, WO 1997-EP2343
     19970507; DE 59708506 G DE 1997-508506 19970507, EP 1997-923007 19970507,
     WO 1997-EP2343 19970507; US 6521460 B1 WO 1997-EP2343 19970507, US
     1999-180806 19990208; ES 2185937 T3 EP 1997-923007 19970507; JP 3523883 B2
     JP 1997-540286 19970507, WO 1997-EP2343 19970507
FDT AU 9728936 A Based on WO 9744661; EP 912891 A1 Based on WO 9744661; JP
     11511856 W Based on WO 9744661; EP 912891 B1 Based on WO 9744661; DE
     59708506 G Based on EP 912891, Based on WO 9744661; US 6521460 B1 Based on
     WO 9744661; ES 2185937 T3 Based on EP 912891; JP 3523883 B2 Previous Publ.
     JP 11511856, Based on WO 9744661
PRAI DE 1996-19620443
                         19960521
     ICM G01N033-50; G01N033-86
          C12Q001-00; G01N001-18; G01N033-53;
          G01N033-569; G01N033-80; G01N035-00
ICA
     G01N033-49
AB
     DE 19620443 A UPAB: 19980112
       Method for performing blood tests comprises treating a
     freshly collected blood sample with at least 1 thrombin
     inhibitor and using the sample to determine blood chemistry and
     optionally haematology parameters.
          ADVANTAGE - The same sample can be used to perform a wide range of
     tests. Only one blood sample has to be taken, which is of
     benefit to patients which may lack sufficient blood already ..
     Dwg.0/0
FS
     CPI EPI
FA
     AB
MC
     CPI: B04-B04D4; B04-B04M; B11-C08E; B12-K04A; D05-H09
     EPI: S03-E13B2; S03-E14H; S03-E14H4; S03-E15
L62
     ANSWER 28 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     1997-087078 [08]
AN
                        WPIX
DNN
    N1997-071762
                        DNC C1997-028266
TI
     Determn. thrombotic risk - by detection of Protein-
     C and Protein-S activity.
DC
     B04 D16 S03
TN
     CAMPBELL, P A; PREDA, L
PA
     (INLI) INSTRUMENTATION LAB SPA
CYC
PΤ
     WO 9642018
                     A1 19961227 (199708) * EN 28
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                                                                      <--
        RW: AT BE CH DE DK ES FI FR GB GR IE IT LU MC NL PT SE
         W: CA JP MX US
     EP 830608
                     A1 19980325 (199816) EN
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         R: AT BE CH DE DK ES FR GB IT LI NL SE
     US 5780255
                  A 19980714 (199835)
                                                      C12Q001-37
                                                                      <--
     JP 11504718
                     W 19990427 (199927)
                                                26
                                                      G01N033-86
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     MX 9709961
                     A1 19981001 (200019)
                                                      G01N033-86
                                                                      <--
     EP 830608
                     B1 20001004 (200050)
                                                      G01N033-86
                                          ΕN
                                                                      <--
         R: AT BE CH DE DK ES FR GB IT LI NL SE
     DE 69610558
                    E 20001109 (200064)
                                                      G01N033-86
                                                                      <--
     ES 2152531
                     T3 20010201 (200112)
                                                      G01N033-86
                                                                      <--
     JP 3248621
                     B2 20020121 (200207)
                                                12
                                                      G01N033-86
                                                                     <--
ADT
    WO 9642018 A1 WO 1996-US9036 19960606; EP 830608 A1 EP 1996-919107
     19960606, WO 1996-US9036 19960606; US 5780255 A US 1995-488510 19950609;
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JP 11504718 W WO 1996-US9036 19960606, JP 1997-503149 19960606; MX 9709961 A1 MX 1997-9961 19971209; EP 830608 B1 EP 1996-919107 19960606, WO 1996-US9036 19960606; DE 69610558 E DE 1996-610558 19960606, EP 1996-919107 19960606, WO 1996-US9036 19960606; ES 2152531 T3 EP 1996-919107 19960606; JP 3248621 B2 WO 1996-US9036 19960606, JP 1997-503149 19960606

FDT EP 830608 A1 Based on WO 9642018; JP 11504718 W Based on WO 9642018; EP 830608 B1 Based on WO 9642018; DE 69610558 E Based on EP 830608, Based on WO 9642018; ES 2152531 T3 Based on EP 830608; JP 3248621 B2 Previous Publ. JP 11504718, Based on WO 9642018

PRAI US 1995-488510 19950609

REP EP 236985; EP 406971; EP 434377; EP 445626; EP 633473; WO 9310261

IC ICM C12Q001-37; G01N033-86

ICS C12N009-48; C12N009-74; C12Q001-56; G01N033-50

AB WO 9642018 A UPAB: 19970220

Determining thrombotic risk in a subject comprises: (1) (a) treating first plasma samples (FPS) with a first reagent to induce or activate coagulation, a second reagent (SR) which activates endogenous protein C in the plasma and a third reagent (TR) comprising calcium salts, phospholipids and/or tissue thromboplastin;

- (b) treating a second plasma sample (SPS) with FR, a buffer or other material which does not activate protein C and TR;
- (c) measuring the time and/or rate necessary for conversion of endogenous fibrinogen to fibrin in both samples;
- (d) calculating the difference in ratio between the time and/or rate of the samples;
- (e) repeating steps (a)-(c) on a sample of normal control plasma, and
- (f) determining the difference or ratio in the times and/or rates obtd. in steps (d) and (e) where the difference is indicative of the thrombotic risk; or (2)
- (a') treating FPS with a first reagent (FR') comprising a synthetic substrate (ss) and SR;
- (b') treating SPS with FR' and a buffer or other material which does not activate protein C;
- (c') measuring the rate of hydrolysis of ss in both samples;
 - (d') calculating the difference between the rates;
- (e') repeating steps (d')-(c') on normal control
 plasma, and
- (f') determining the difference or ratio in the rates obtd. in steps (d') and (e'), the difference being indicative of thrombotic risk.

Also claimed are kits comprising a first container comprising:

- (1) (a) a first container comprising FR, SR and TR and (b) a second container for adding a second plasma sample, FR, buffer or other material which does not activate protein C and TC, and
- (2) (a') a first container comprising FR', and SR; (b') a second container for adding to a SPS FR' and a buffer or other material which does not activate protein C.
- USE The global test can be a diagnostic aid for the evaluation of thrombotic risk in patients suffering from hereditary and non-hereditary thrombophilia, i.e. disorders of the haemopoietic system in which there is a tendency toward the occurrence of thrombosis, or otherwise in patients undergoing particularly pharmacologic treatments such as extroprogestinics. Additionally patients undergoing surgery can be evaluated to determine thrombotic risk. The test is used in vitro to diagnose clotting anomalies involved in the inhibitory system, due to

```
either protein C (PC) or S (PS) deficiency or the
     presence of molecular anomalies of Factor Va or to the presence of anti-a
     PC antibodies.
     Dwq.0/1
FS
     CPI EPI
FA
     AB; DCN
MC
     CPI: B04-B01B; B04-B04G; B04-H19; B04-N02; B05-A01B; B05-B01P;
          B11-C08D3; B12-K04A; D05-H09
     EPI: S03-E14H1; S03-E14H5
     ANSWER 29 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
L62
AN
     1996-200281 [20]
                        WPIX
DNN
    N1996-168115
                        DNC C1996-063239
     Determination of individual's inflammation index in anti-
TT
     coagulated whole blood - by using whole blood
     fibrinogen and haematocrit or haemoglobin
     measurements.
חכ
     B04 S03
TN
     BULL, B S; LEVINE, R A; WARDLAW, S C
     (BULL-I) BULL B S; (LEVI-I) LEVINE R A; (WARD-I) WARDLAW S C; (LEVI-I)
PA
     LEVIN R A; (WARD-I) WARDROW S C
CYC
PΙ
     US 5506145
                     A 19960409 (199620) *
                                                 6
                                                      G01N033-86
                     A2 19960605 (199627) EN
     EP 715170
                                                 6
                                                      G01N033-49
                                                                      <--
         R: AT BE CH DE DK ES FR GB GR IE IT LI NL PT SE
                   A 19960613 (199631)
     AU 9520211
                                                      G01N033-72
                     A 19960603 (199631)
     NO 9504897
                                                      G01N033-68
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                     A 19960625 (199635)
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     JP 08166389
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     CA 2149580
                     Α
                        19960603 (199639)
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                     Α
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     CN 1123914
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     TW 321721
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     RU 2122212
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                                                      G01N033-86
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    US 5506145 A US 1994-348345 19941202; EP 715170 A2 EP 1995-118964
ADT
     19951201; AU 9520211 A AU 1995-20211 19950523; NO 9504897 A NO 1995-4897
     19951201; JP 08166389 A JP 1995-170802 19950706; CA 2149580 A CA
     1995-2149580 19950517; FI 9505784 A FI 1995-5784 19951201; CN 1123914 A CN
     1995-108425 19950714; TW 321721 A TW 1995-106027 19950613; AU 690972 B AU
     1995-20211 19950523; RU 2122212 C1 RU 1995-113187 19950714
FDT
    AU 690972 B Previous Publ. AU 9520211
PRAI US 1994-348345
                          19941202
REP
    No-SR. Pub
IC
     ICM C12Q001-56; G01N033-49; G01N033-68;
          G01N033-72; G01N033-86
         G01N015-05; G01N033-53
ICA
    A61K049-00
AB
        5506145 A UPAB: 19960520
     Deg of mammalian donor systemic inflammation is determined from a sample
     of donor anticoagulated whole blood A sample of the
     blood is drawn from a donor into a transparent tube (2) containing an
     elongated float (4). The sample is centrifuged in the tube. The amount of
     fibrinogen/fibrin in the blood sample in the tube is
     measured. A haematocrit or haemoglobin value for the
     blood sample in the tube is determined. A number I, indicative of
     the degree of systemic inflammation, is computed by solving equation (i) I
     = a(f) + b(h) + c where f is the measured fibrinogen/fibrin
     level in the sample; h is the determined haematocrit or
     haemoglobin value in the sample; and a, b and c are empirically
     derived constants. The numerical value of I is correlated with the deg. of
     mammalian donor systemic inflammation.
          USE - Method determines the presence or absence of
     inflammatory condition in patient by quantifying fibrinogen content and
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haematocrit or haemoglobin content in sample of patient's blood. Patient may be human, or process may be used in veterinary work. ADVANTAGE - Method is not susceptible to systemic abnormalities that render Westergren erythrocyte sedimentation rate method unreliable. Amount of blood required is small and time required for procedure is short. Dwg.1/1 FS CPI EPI AB; GI FA CPI: B04-B04D2; B11-C08E; B12-K04A MC EPI: S03-E14H ANSWER 30 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN L62 1996-130880 [14] WPIX AN DNN N1996-109976 DNC C1996-040926 Determn. of fibrinogen concentration in undiluted plasma sample TIcomprises addition of novel reagent containing thrombin or protease, in presence of high concentration of salt. DC B04 D16 S03 IN ENOMOTO, M (NNTR) NIPPON SHOJI KAISHA LTD; (AZWE-N) AZWELL INC; (NNTR) NIPPON SHOJI PA KK CYC A2 19960306 (199614) * EN PΙ EP 699909 19 G01N033-86 <--R: DE FR GB JP 08070895 A 19960319 (199621) 10 C12Q001-56 <--A3 19960619 (199635) EP 699909 G01N033-86 <--US 5851836 A 19981222 (199907) G01N033-49 <--JP 2994557 B2 19991227 (200006) 10 C12Q001-56 <---EP 699909 B1 20011128 (200201) EN G01N033-86 <--R: DE FR GB DE 69524161 E 20020110 (200211) G01N033-86 ADT EP 699909 A2 EP 1995-113736 19950901; JP 08070895 A JP 1994-209940 19940902; EP 699909 A3 EP 1995-113736 19950901; US 5851836 A US 1995-521868 19950831; JP 2994557 B2 JP 1994-209940 19940902; EP 699909 B1 EP 1995-113736 19950901; DE 69524161 E DE 1995-624161 19950901, EP 1995-113736 19950901 JP 2994557 B2 Previous Publ. JP 08070895; DE 69524161 E Based on EP 699909 FDT PRAI JP 1994-209940 19940902 1.Jnl.Ref; EP 537490; EP 570354; EP 632270; JP 05219993; US 5292664; WO REP 9407145 IC ICM C12Q001-56; G01N033-49; G01N033-86 ICS C12Q001-37 AB 699909 A UPAB: 19960405 Method for determn. of fibrinogen (I) concentration comprises: (1) addition of thrombin, or a protease having similar activity, to an undiluted sample (if plasma) in a reaction mixture containing a salt (II) at high concentration, then (2) measurement of the coagulation time. The concentration of (II) is set at a level giving a coagulation time of 5-100 secs. at 37deg.C. using a mixture of a fibrinogen-containing sample (275 mg/dl) and a reagent (III) containing thrombin (100NIHU/ml and HEPES (RTM:buffer) (100mM; pH 7.35;) the volume ratio sample (III) being from 1-2 (pref. 1:1.0-1.8). Salt (II) is 1 of :- NaCl (0.25-3.0 concentration), NaBr (0.1-1.0), NaI (0.1-0.4), KCl (0.25-1.5), KBr (0.1-1.0), KI (0.1-0.4), MgCl2 (0.04-0.25), CaCl2 (0.04-0.25). A pref. (III) contains 1.0-2.5M NaCl and 0.1-0.8M NaBr, and an especially pref. reaction mixture comprises 0.25-1.0M NaCl, 0.05-0.2 KF or NAF, 2-50 mM Na citrate and, as a discrepancy preventive (IV), 0.001-0.5 w/v%

of a surfactant. Also claimed are reagents per se. These comprise salt a t

high concentration (set at a level giving a coagulation time

of 5-100 secs., measured under conditions as described above) and 20-500 NIHU/ml of thrombin or a protease. An especially pref. reagent comprises 40-200 (NIHU/ml. thrombin, 30-200 ml. buffer (pH 7.0-8.0), 1.0-3.5M NaCl and 0.3-1.0 MNaBr. Alternatively, 2 reagents may be used, a first comprising (IV) and a second consisting of thrombin or a protease and (II) may be in the first or second reagent. ADVANTAGE - The method uses undiluted samples of plasma, and conventional equipment for measurement of coagulation time. Samples having a low content of (I) can be assayed using a normal amount of thrombin without use of expensive peptides or prolonging the coagulation time. The results correlate well with those obtd. by the conventional dilution method and can be used for routine blood tests. Dwg.0/9 CPI EPI AB; DCN CPI: B04-B04D4; B04-H19; B04-L05C; B05-A01A; B05-A01B; B11-C08E; B12-K04; D05-A02C; D05-H09 EPI: S03-E14H ANSWER 31 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1995-233339 [31] WPIX DNN N1995-181839 DNC C1995-107722 Determn. of thrombin-induced platelet aggregation in presence of fibrin - comprises use of fibrin aggregation inhibitor to suppress fibrin clot formation. B04 D16 S03 REERS, M (DADE-N) DADE BEHRING MARBURG GMBH; (BEHW) BEHRINGWERKE AG 2.0 EP 661383 A2 19950705 (199531) * GE 5 C12Q001-56 R: AT BE CH DE DK ES FR GB GR IE IT LI LU NL PT SE DE 4344919 A1 19950706 (199532) 5 C12Q001-56 A 19950706 (199534) AU 9481788 C12N009-99 A 19950701 (199539) CA 2138931 C120001-56 <--A 19950808 (199540) JP 07203994 4 C12Q001-56 <--US 5563041 A 19961008 (199646) 6 C12Q001-56 <--EP 661383 A3 19971217 (199818) C12Q001-56 <--AU 702099 B 19990211 (199918) C12N009-99 EP 661383 B1 20010321 (200117) GE C12Q001-56 R: AT BE CH DE DK ES FR GB GR IE IT LI LU NL PT SE DE 59409698 G 20010426 (200124) C12Q001-56 < - -ES 2155842 T3 20010601 (200137) C12Q001-56 <--EP 661383 A2 EP 1994-119803 19941215; DE 4344919 A1 DE 1993-4344919 19931230; AU 9481788 A AU 1994-81788 19941229; CA 2138931 A CA 1994-2138931 19941222; JP 07203994 A JP 1994-326514 19941228; US 5563041 A US 1994-365759 19941229; EP 661383 A3 EP 1994-119803 19941215; AU 702099 B AU 1994-81788 19941229; EP 661383 B1 EP 1994-119803 19941215; DE 59409698 G DE 1994-509698 19941215, EP 1994-119803 19941215; ES 2155842 T3 EP 1994-119803 19941215 AU 702099 B Previous Publ. AU 9481788; DE 59409698 G Based on EP 661383; ES 2155842 T3 Based on EP 661383 PRAI DE 1993-4344919 19931230 EP 336353; EP 537490; US 5246832 ICM C12N009-99; C12Q001-56 A61K035-14; C07K005-08; C07K005-10; C07K007-06; C12N009-48; C12N009-74; C12Q001-00; G01N033-48; G01N033-49; G01N033-86

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In the qualitative or quantitative determn. of thrombin-induced

platelet aggregation in the presence of fibrin, interference from fibrin clot formation is suppressed with a fibrin-aggregation inhibitor (I). Also claimed is a diagnostic reagent comprising thrombin and (I). USE - The assay may be used to determine the platelet aggregation inhibitory activity of thrombin inhibitors. ADVANTAGE - (I) inhibits fibrin clot formation at high thrombin concns. without inhibiting platelet aggregation. Dwg.0/2 CPI EPI FS FA AB; DCN MC CPI: B04-C01; B04-M01; B12-K04A2; D05-A02C EPI: S03-E14H1; S03-F03 5563041 A UPAB: 19961115 ABEQ US A method for the qualitative or quantitative determination of platelet aggregation induced by thrombin in the presence of fibrin, wherein the formation of an interfering fibrin clot is prevented by the presence of an inhibitor of fibrin aggregation and the experimentally induced platelet aggregation is unaffected. Dwq.0/2 L62 ANSWER 32 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1995-061015 [08] AN WPIX DNN N1995-048479 DNC C1995-027174 Method for the quantification of active-plasminogen-TТ activator-inhibitor-type-1 - present in a sample of blood or plasma is useful in the field of haemostasis. DC B04 D16 S03 IN NIEUWENHUIZEN, W PA (ALKU) AKZO NOBEL NV CYC 23 PΙ WO 9501452 A1 19950112 (199508)* EN 41 C12Q001-56 RW: AT BE CH DE DK ES FR GB GR IE IT LU MC NL PT SE W: AU CA FI JP KR US AU 9473844 A 19950124 (199520) C12Q001-56 EP 706580 A1 19960417 (199620) EN C12Q001-56 R: AT BE CH DE DK ES FR GB GR IE IT LI LU MC NL PT SE JP 08512133 W 19961217 (199710) 43 G01N033-53 US 5753457 A 19980519 (199827) C12Q001-06 <--WO 9501452 A1 WO 1994-EP2127 19940628; AU 9473844 A AU 1994-73844 TOA 19940628; EP 706580 A1 EP 1994-923704 19940628, WO 1994-EP2127 19940628; JP 08512133 W WO 1994-EP2127 19940628, JP 1995-503274 19940628; US 5753457 A WO 1994-EP2127 19940628, US 1996-481284 19960221 AU 9473844 A Based on WO 9501452; EP 706580 A1 Based on WO 9501452; JP FDT 08512133 W Based on WO 9501452; US 5753457 A Based on WO 9501452 PRAI EP 1993-201880 19930628 06Jnl.Ref; EP 339302; EP 450086; WO 8600413; WO 890005; WO 9323561 IC ICM C12Q001-06; C12Q001-56; G01N033-53 ICS G01N033-566; G01N033-86 AB 9501452 A UPAB: 19950301 Quantification of active plasminogen-activator-inhibition-type-1 present in a sample comprises taking 2 portions from the sample and: (a) determining in 1 portion a valve corresp. to the total amount of complex between (I) and plasminogen activator (PA), the so-called PA-(I) complex with PA already present and the moment of sampling; and (b) calculating the amount of PA-(I) present at the moment of sampling; (c) determining in the other portion a value corresp. to the total amount of (II) present after addition of an excess of the active form of tissue type PA (t-PA) to the portion; and (d) calculating the amount of PA-(I) present after addition of an excess of the active form of t-PA to the portion; and (e) subtracting the total amount of complex from the total amount of PA-(I)

resulting in the amount of active (I) that was present at the moment of sampling in a volume of the sample equivalent to the volume of the portions, where the portion of step (a) is contacted with means for inhibiting complexation of PA with (I) from the moment of sampling through step (a). Step (a) comprises: (i) contacting the portion with 1 immobilised capturing agent (CAP), which (1) is directed at PA partic. of the PA-(I) complex and (2) is present in an amount and with a specificity sufficient for the mol. forms of PA, pref. of the PA-(I) complex present in the portion to be complexed to it; (ii) contacting the portion with 1 tagging agent (TAG) which: (1) is provided with a detectable marker (*); (2) is directed at plasminogen-activation-inhibitor (PAI), pref. PAI of PAI-(I) and (3) is present in an amount and with a specificity sufficient for the mol. forms of (I) present in the portions to be complexed to it; (iii) carrying out appropriate incubation and washing steps; (iv) determining a value corresp. to an amount of (*) that has been bound by an immobilised CAP. Step (c) comprises: (i) contacting the other portion with an excess of the active form of t-PA, thereby allowing non-bound active (I) present in the portion before the addition to form PA-(I); (ii) contacting this portion with TAG and CAP as in step (a), (iii) subsequently determining a value corresp. to the amount of (*) that has been bound to the immobilised CAP. Also claimed are: (A) use of a t-PA determn. assay to determine the complexed form of t-PA in a method for determining the specific known concentration of PA-(I) in the reference sample be used; (B) use of a method comprising: (1) subjecting the reference sample to conditions such that virtually all t-PA present is converted to t-PA-(I) co Dwg.4/4 CPI EPI AB; GI CPI: B04-B04D4; B04-B04D5; B04-G21; B04-L03B; B11-C07A4; B12-K04A2; B14-D07C; D05-H09; D05-H10; D05-H11A EPI: S03-E14H1 L62 ANSWER 33 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1994-007456 [01] WPIX 1994-007188 [01] DNC C1994-002979 Test for quantitative thrombin time - using plasma dilutions, excess fibrinogen and thrombin, giving more accurate results than standard tests. ALVING, B; HENDRICKS, G; REID, T; ALVING, B M; REID, T J (USSA) US DEPT OF ARMY; (ALVI-I) ALVING B M; (REID-I) REID T J CYC 19 WO 9325578 A1 19931223 (199401) * EN 30 C07K007-10 RW: AT BE CH DE DK ES FR GB GR IE IT LU MC NL PT SE W: AU CA JP AU 9347681 A 19940104 (199417) C07K007-10 W 19960227 (199643) 29 JP 08501682 C12Q001-56 WO 9325578 A1 WO 1993-US5315 19930603; AU 9347681 A AU 1993-47681 19930603; JP 08501682 W WO 1993-US5315 19930603, JP 1994-501586 19930603 AU 9347681 A Based on WO 9325578; JP 08501682 W Based on WO 9325578 19920605; US 1993-21033 PRAI US 1992-893631 19930222 2.Jnl.Ref; US 4379142; US 4496653; US 4767742; US 4952562; US 5019393; US 5118790; US 5187102; US 5196404 ICM C07K007-10; C12Q001-56 ICS A61K009-22; A61K031-445; A61K037-00; A61K037-02; A61K037-43; C07K007-08; G01N033-50; G01N033-86 9325578 A UPAB: 19950223

(A) quantitating plasma levels of specific inhibitors of

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thrombin, comprising: (a) preparing serial dilutions of a thrombin-specific inhibitor (TSI) in known concns. to give samples; (b) mixing the samples with purified human fibrinogen (PHF); (c) adding purified human alpha-thrombin (PHAT) to the samples prepared in (b); (d) measuring the clotting times of the solns. in (c); (e) plotting a standard curve from the effect of TSI based on the results obtained in (d); (f) diluting a plasma sample from patients receiving a TSI; (g) mixing the patient samples with a solution of fibrinogen of step (f) (sic); (h) adding a thrombin solution to the samples prepared in step (g); (i) measuring the clotting times of the solns. of step (h); and (j) determining the concentration of the inhibitor in the patient samples using the standard curve. USE/ADVANTAGE - The new method of measuring thrombin time is not affected by the presence of abnormal plasmas which cause prolongation of standard tests. The method can be performed on standard laboratory instruments, and are fast and inexpensive. The method can be used in analysis of thrombosis. Dwg.0/4 Dwg.0/4 CPI AB CPI: B04-B04D2; B04-H19; B11-C07; B12-K04A2 ANSWER 34 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN L62 1993-395336 [49] WPIX DNN N1993-305443 DNC C1993-176033 Determining risk of thrombosis - by measuring procoagulant activity of platelets, namely resting activity and/or excitability of platelets. B04 D16 S03 HEMKER, H C; WAGENVOORD, R J (BAXT) BAXTER DIAGNOSTICS INC; (DADE-N) DADE PROD AG; (DADE-N) DADE PRODN AG 21 US 5266462 A 19931130 (199349)* 29 C12Q001-56 WO 9324840 A1 19931209 (199350) EN 63 G01N033-86 <--RW: AT BE CH DE DK ES FR GB GR IE IT LU MC NL PT SE W: AU CA JP A 19931230 (199415) G01N033-86 AU 9344090 A1 19940713 (199427) EP 605674 EN G01N033-86 R: AT BE CH DE DK ES FR GB IE IT LI NL SE JP 0650947**9** W 19941027 (199502) C12Q001-56 В AU 658073 19950330 (199521) C12Q001-56 <--CA 2114719 С 19970826 (199748) C12Q001-56 <--B1 19980121 (199808) EP 605674 EN G01N033-86 <--R: AT BE CH DE DK ES FR GB IE IT LI NL SE DE 69316572 E 19980226 (199814) G01N033-86 T3 19980516 (199826) ES 2114053 G01N033-86 <--JP 3143694 B2 20010307 (200116) 31 C12Q001-56 US 5266462 A US 1992-892865 19920603; WO 9324840 A1 WO 1993-US5436 ADT 19930603; AU 9344090 A AU 1993-44090 19930603; EP 605674 A1 EP 1993-912364 19930603, WO 1993-US5436 19930603; JP 06509479 W WO 1993-US5436 19930603, JP 1994-500887 19930603; AU 658073 B AU 1993-44090 19930603; CA 2114719 C CA 1993-2114719 19930603; EP 605674 B1 EP 1993-912364 19930603, WO 1993-US5436 19930603; DE 69316572 E DE 1993-616572 19930603, EP 1993-912364 19930603, WO 1993-US5436 19930603; ES 2114053 T3 EP 1993-912364 19930603; JP 3143694 B2 WO 1993-US5436 19930603, JP 1994-500887 19930603 AU 9344090 A Based on WO 9324840; EP 605674 A1 Based on WO 9324840; JP FDT 06509479 W Based on WO 9324840; AU 658073 B Previous Publ. AU 9344090,

Based on WO 9324840; EP 605674 B1 Based on WO 9324840; DE 69316572 E Based

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on EP 605674, Based on WO 9324840; ES 2114053 T3 Based on EP 605674; JP 3143694 B2 Previous Publ. JP 06509479, Based on WO 9324840 PRAI US 1992-892865 19920603 REP 5.Jnl.Ref ICM C12Q001-56; G01N033-86 IC ICS A01N037-10; C12Q001-32; C12Q001-37; G01N033-50; G01N033-92 AB 5266462 A UPAB: 19960322 Determining the risk of thrombosis in a patient by determining the pro-coagulant activity of resting platelets comprises; (a) mixing a sample containing platelets from a patient with a substrate (I) which can be converted by a pro-coagulant phospholipid dependent enzyme or enzyme complex; (b) contacting and reacting the mixture with the enzyme or enzyme complex to form an activated substrate; (c) determining the amount of the formed activated substrate in the sample; (d) comparing the amount of activated substrate with the amount of formed activated substrate from one or more control individuals. Also claimed is a method for determining the risk of thrombosis in a patient by determining the excitability of platlets by (i) incubating a sample containing platelets from a patient with thrombin or thormbin plus collagen; (ii) mixing the prod. with (I); (iii) as for step (b) above; (iv) as for step (c) above; and (v) comparing the excitability of platelets from the patient with the excitability of platelets from one or more control individuals. The method is also claimed for determining if an agent (II) will effectively inhibit platelet activation. The sample containing platelets is incubated iwth thrombin (or thrombin plus collagen) and (II), then mixed with (I) and reacted with the enzyme or enzyme complex. The amount of formed activated substrate from the sample containing (II) is compared with the amount of formed activated substrate from a (II)-free sample. USE/ADVANTAGE - The resting activty and/or excitability of platelets can be used to determine patients at risk of thrombosis (as claimed). The test is based on the amount of procoagulant phospholipids which are exposed at the outer membrane of platelets. The flip-flop reaction of procoagulant phospholipids together with the ability to determine the presence of pro-coagulant phospolipids in the outer membrane also provides the basis for the evaluation fo (II) (e.g. drugs) which, e.g. inhibit the flip-flop reaction. Dwg.0/18 Dwg.0/18 FS CPI EPI FA AB; DCN MC CPI: B04-B01B; B04-B02C3; B04-B04D3; B04-B04D5; B05-B01P; B11-C08E3; B12-K04A2; D05-A02C; D05-H09 EPI: S03-E14H1 ABEO EP 605674 B UPAB: 19980223 Determining the risk of thrombosis in a patient by determining the pro-coagulant activity of resting platelets comprises; (a) mixing a sample contg. platelets from a patient with a substrate (I) which can be converted by a pro-coagulant phospholipid dependent enzyme or enzyme complex; (b) contacting and reacting the mixt. with the enzyme or enzyme complex to form an activated substrate; (c) determining the amt. of the formed activated substrate in the sample; (d) comparing the amt. of activated substrate with the amt. of formed activated substrate from one or more control individuals. Also claimed is a method for determining the risk of thrombosis in a patient by determining the excitability of platlets by (i) incubating a sample contg. platelets from a

patient with thrombin or thormbin plus collagen; (ii) mixing the prod. with (I); (iii) as for step (b) above; (iv) as for step (c) above; and (v) comparing the excitability of platelets from the patient with the excitability of platelets from one or more control individuals. The method is also claimed for determining if an agent (II) will effectively inhibit platelet activation. The sample contg. platelets is incubated iwth thrombin (or thrombin plus collagen) and (II), then mixed with (I) and reacted with the enzyme or enzyme complex. The amt. of formed activated substrate from the sample contg. (II) is compared with the amt. of formed activated substrate from a (II)-free sample. USE/ADVANTAGE - The resting activty and/or excitability of platelets can be used to determine patients at risk of thrombosis (as claimed). The test is based on the amt. of procoagulant phospholipids which are exposed at the outer membrane of platelets. The flip-flop reaction of procoagulant phospholipids together with the ability to determine the presence of pro-coagulant phospolipids in the outer membrane also provides the basis for the evaluation fo (II) (e.g. drugs) which, e.g. inhibit the flip-flop reaction. Dwq.0/18 L62 ANSWER 35 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 1993-379548 [48] AN WPIX DNN N1993-293156 DNC C1993-168407 Measuring protein C or S activity in plasma TI - by activation with thrombomodulin and determn. of its effect on endogenous thrombin formation, especially for assessing risk of thrombosis. DC B04 D16 J04 S03 AIACH, M G H; PITTET, J R IN (INMR) BIO MERIEUX PA CYC 1 PΙ FR 2689640 A1 19931008 (199348) * 21 G01N033-52 ADT FR 2689640 A1 FR 1992-4184 19920406 PRAI FR 1992-4184 19920406 IC ICM G01N033-52 AB 2689640 A UPAB: 19940120 Determn. of protein C and/or S in a plasma sample comprises: a) preparing a mixture of: (i) plasma sample; (ii) a predetermined quantity of thrombomodulin; (iii) at least one activated coagulation factor and/or at least one coagulation factor activator to allow the formation of endogenous thrombin; b) incubating the obtd. mixture in conditions allowing activation of protein C and expression of activated protein C; c) adding a thrombin substrate to the mixture; and d) quantifying the protein C and/or S activity in the sample by the determn. of enzyme activity of the thrombin on the substrate. Pref. the mixture is incubated for 3-6 (5) minutes at 37 deg. C. The thrombomodulin may be human or animal, natural or recombinant, and/or modified, especially by an enzyme such as chondroitinase, trypsin, elastase or analogues, and is pref. soluble. The activated coagulation factors are pref. activated factors XII and/or X. The substrate is pref. fibrinogen. ADVANTAGE - A simple, easily automated, sensitive process which does not require the use of an exogenous activator and can be used even for low concns. of proteins C and/or S.

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Dwg.
     CPI EPI
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     AB: GI
MC
     CPI: B04-B04A6; B04-B04D3; B04-B04D4; B11-C08E3;
          B12-K04A2; D05-A02C; D05-H09; J04-C02
     EPI: S03-E14H1
L62 ANSWER 36 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1993-321661 [41]
                        WPIX
DNC C1993-143097
TI
     Turbidimetric agglutination assay for use in centrifugal
     analyser - involves comparing turbidity decrease of
     analyte with known analyte concentration after reaction under centrifugal
     conditions, useful for e.g. clotting proteins.
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     A89 B04 D16 J04
     KRAUS, M
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     (BEHW) BEHRINGWERKE AG; (DADE-N) DADE BEHRING MARBURG GMBH
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                     A1 19931007 (199341) *
PΤ
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                     A 19931005 (199401)
     CA 2093307
                                                      G01N033-546
                     A 19940125 (199408)
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                                                      G01N033-543
     AU 661764
                     В
                        19950803 (199539)
                                                      G01N021-59
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                     Α
                        19961105 (199650)
                                                14
                                                      G01N033-546
                     B1 19991215 (200003)
     EP 568797
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                                                      G01N033-546
         R: AT BE CH DE DK ES FR GB IT LI LU NL PT SE
                     G 20000120 (200011)
     DE 59309900
                                                      G01N033-546
     ES 2141735
                     T3 20000401 (200023)
                                                      G01N033-546
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     JP 3327484
                     B2 20020924 (200264)
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ADT DE 4211351 A1 DE 1992-4211351 19920404; EP 568797 A1 EP 1993-104120
     19930313; AU 9335653 A AU 1993-35653 19930402; CA 2093307 A CA
     1993-2093307 19930402; JP 06018528 A JP 1993-76036 19930402; AU 661764 B
     AU 1993-35653 19930402; US 5571728 A US 1993-41210 19930401; EP 568797 B1
     EP 1993-104120 19930313; DE 59309900 G DE 1993-509900 19930313, EP
     1993-104120 19930313; ES 2141735 T3 EP 1993-104120 19930313; JP 3327484 B2
     JP 1993-76036 19930402
FDT
    AU 661764 B Previous Publ. AU 9335653; DE 59309900 G Based on EP 568797;
     ES 2141735 T3 Based on EP 568797; JP 3327484 B2 Previous Publ. JP 06018528
PRAI DE 1992-4211351
                          19920404
     ICM
         G01N021-59; G01N033-50; G01N033-543;
          G01N033-546
         G01N021-82; G01N033-53
ICA
    G01N033-531; G01N033-86
          4211351 A UPAB: 19931130
AB
     Determn. of an analyte in a sample is effected by incubating the sample
     with an analyte-specific binding reagent (immobilised) on a particulate
     support, where: (a) the reaction takes place under the influence of a
     centrifugal force which remains constant throughout the measurement
     process; (b) the particle concentration in the reaction mixture is at least 0.1
     weight%; (c) the decrease in turbidity is measured (sic measurement
     is commenced) immediately after the mixing and distribution phase
     instigated by sample addition; and (d) the analyte concentration is determined
by
     comparing the turbidity decrease for the sample with the
     values obtained under identical conditions for samples with known analyte
     contents.
          USE/ADVANTAGE - The method may be used to determine
     proteins or peptides of the clotting, fibrinolysis or complement
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system, especially the **fibrin** cleavage prod. D-dimer. The method gives quantitative results, with good precision and reproducibility within the **time** (3-8 min.) required for use in

conventional automatic centrifigual analysers. Dwg.0/0

FS CPI

FA AB

MC CPI: A12-L04; A12-V03C2; B04-B04A; B04-B04C6; B04-C01; B11-C07A6; B12-K04; D05-H09; D05-H13; J04-B01

ABEQ EP 568797 A UPAB: 19931220

Determn. of an analyte in a sample is effected by incubating the sample with an analyte-specific binding reagent (immobilised) on a particulate support, where: (a) the reaction takes place under the influence of a centrifugal force which remains constant throughout the measurement process; (b) the particle concn. in the reaction mixt. is at least 0.1 wt.%; (c) the decrease in turbidity is measured (sic measurement is commenced) immediately after the mixing and distribution phase instigated by sample addn.; and (d) the analyte concn. is determined by comparing the turbidity decrease for the sample with the values obtained under identical conditions for samples with known analyte contents.

USE/ADVANTAGE - The method may be used to determine proteins or peptides of the clotting, fibrinolysis or complement system, esp. the fibrin cleavage prod. D-dimer. The method gives quantitative results, with good precision and reproducibility within the time (3-8 min.) required for use in conventional automatic centrifugal analysers.

Dwg.0/6

ABEQ US 5571728 A UPAB: 19961211

Determining the concn. of an analyte in <8 mins. comprises:

- a) mixing and distributing in an automatic centrifugal analyzer a sample of a biological material contg. the analyte with 1 binding partner specific for the analyte, said binding partner being immobilized on a particulate carrier material wherein the concn. of the particulate carrier material in the mixt. is >0.09 wt.%;
- b) maintaining constant centrifugal acceleration of 10-10000 g during the mixing and distributing step;
- c) determining a decrease in absorption as a measurement of turbidity immediately upon completion of the mixing and distributing step, and
- d) determining the concn. of the analyte by comparing the measurement of turbidity obtained in step c) with values measured under identical conditions for samples of known analyte content. Dwg.0/6

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L62 ANSWER 37 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
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AN 1993-182573 [22] WPIX

DNN N1993-140319 DNC C1993-080921

TI Method for diagnosing blood coagulation disorders e.g. thromboembolism - using in-vitro coagulation assay including activated protein C.

DC B04 D16 S03

IN DAHLBACK, B; DAHLBAECK, B

PA (DAHL-I) DAHLBACK B; (DAHL-I) DAHLBAECK B; (TACT-N) TAC THROMBOSIS & COAGULATION AB

CYC 19

PΙ WO 9310261 A1 19930527 (199322) * EN C12Q001-56 RW: AT BE CH DE DK ES FR GB GR IT LU MC NL SE W: AU CA JP US SE 9103332 A 19930514 (199329) G01N033-86 A 19930615 (199340) AU 9221980 C12Q001-56 <--B 19931220 (199402) SE 470274 G01N033-86 <--A1 19940803 (199430) EP 608235 EN C12Q001-56 <--

R: AT BE CH DE DK ES FR GB GR IT LI LU MC NL SE

JP 07501217 W 19950209 (199515) C12Q001-56 <--

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US 5443960
                    A 19950822 (199539)
                                                     C12Q001-56
                                                                     <--
                    B1 19960110 (199607) EN
                                                     C12Q001-56
     EP 608235
                                               19
                                                                     <--
        R: AT BE CH DE DK ES FR GB GR IT LI LU MC NL SE
     DE 69207607 E 19960222 (199613)
                                                     C120001-56
                   B 19960215 (199614)
     AU 666484
                                                     C120001-56
                                                                    <--
                  T3 19960316 (199618)
    ES 2081618
                                                     C12Q001-56
                                                                    <--
     JP 2562000
                  B2 19961211 (199703)
                                                     C12Q001-56
                                                                    <--
    CA 2119761
                    C 19971216 (199810)
                                                     C12Q001-56
                                                                    <--
     US 5443960
                    B1 19990112 (199910)
                                                     C12Q001-56
                                                                     <--
ADT WO 9310261 A1 WO 1992-SE310 19920513; SE 9103332 A SE 1991-3332 19911113;
     AU 9221980 A AU 1992-21980 19920513; SE 470274 B SE 1991-3332 19911113; EP
     608235 A1 EP 1992-913443 19920513, WO 1992-SE310 19920513; JP 07501217 W
     WO 1992-SE310 19920513, JP 1993-509186 19920513; US 5443960 A WO
     1992-SE310 19920513, US 1994-199328 19940527; EP 608235 B1 EP 1992-913443
     19920513, WO 1992-SE310 19920513; DE 69207607 E DE 1992-607607 19920513,
     EP 1992-913443 19920513, WO 1992-SE310 19920513; AU 666484 B AU 1992-21980
     19920513; ES 2081618 T3 EP 1992-913443 19920513; JP 2562000 B2 WO
     1992-SE310 19920513, JP 1993-509186 19920513; CA 2119761 C CA 1992-2119761
     19920513; US 5443960 B1 WO 1992-SE310 19920513, US 1994-199328 19940527
FDT AU 9221980 A Based on WO 9310261; EP 608235 A1 Based on WO 9310261; JP
     07501217 W Based on WO 9310261; US 5443960 A Based on WO 9310261; EP
     608235 B1 Based on WO 9310261; DE 69207607 E Based on EP 608235, Based on
     WO 9310261; AU 666484 B Previous Publ. AU 9221980, Based on WO 9310261; ES
     2081618 T3 Based on EP 608235; JP 2562000 B2 Previous Publ. JP 07501217,
     Based on WO 9310261; US 5443960 B1 Based on WO 9310261
PRAI SE 1991-3332
                         19911113; US 1991-811303
     1.Jnl.Ref; EP 434377; SE 464135; WO 9101282; WO 9102812; 6.Jnl.Ref
REP
IC
     ICM C12Q001-56; G01N033-86
     ICS
         G01N033-50
AB
          9310261 A UPAB: 19931115
     An in vitro method for the diagnosis of blood
     coagulation disorders, in partic. thromboembolic
     diseases in a human or for the detection of the risk for a human to
     acquire blood coagulation disorders, where the
     disorders are not expressed by Protein S deficiency or defective Factor
     VIIIa, or the disorders are not related to Protein S deficiency,
     comprising (i) incubating a sample containing human coagulation
     factors with (a) an exogenous reagent (I) which at least partially
     activates the blood coagulation system of the
     sample, (b) an activated exogenous Protein C
     (APC) or exogenous PC together with exogenous reagents (II) that transform
     exogenous PC to APC, components, such as phospholipid and Ca(2+)
     salt, are necessary for efficient reaction of the activated
     coagulation factors introduced in step (1, a) and (d) if desired,
     an exogenous sustrate for an eznyme, which activity is influenced by
     activated Protein C, the final assay media
     pref. having at least a patient plasma sample content that is
     greater than 10%, in partic. greater than 20% or 35% (v/v), (ii) directly
     monitoring a substrate conversion rate for a blood
     coagulation eznyme which activity is influenced by
     activated Protein C and (iii)
     comparing the conversion rate determined in step (ii)
     with a standard value obtd. from samples of normal
     individuals subjected to steps (i) and (ii) under identical conditions;
     where the finding of a sample conversion rate that is not
     normal compared to the standard value is taken
     as an indication of the human suffering from the disorder or as being at
     risk for acquiring the disorder, in partic. an enhanced conversion
     rate is taken as an indication of a thromboembolic
     disease or a risk for acquiring such disease.
          Reagent (I) may be e.g. Factor Ax, Factor IXa or an APTT reagent.
          USE/ADVANTAGE - The method can be used for screening and
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diagnosis of e.q. hereditary thrombophilia, in pregnant

individuals, individuals undergoing surgery, individuals taking anti-conception drugs, etc. $\mbox{\rm Dwg.}\ 0/0$

FS CPI EPI

FA AB

MC CPI: B04-B01B; B04-B02C3; B04-B04A6; B04-B04D3;

B04-B04D4; B05-A01B; B05-B01P; B11-C08E; B12-K04A2;

D05-H09

EPI: S03-E14H1

ABEQ US 5443960 A UPAB: 19951004

In vitro screening and diagnosis of activated protein C(APC) resistance comprises (i) incubating human plasma with (a) exogenous APC, protein C and an exogenous reagent (b) an exogenous reagent partially activating a coagulation factor and opt. (c) an exogenone substrate for an enzyme affected by APC; (ii) measuring substrate conversion rate for a coagulation factor affected by APC; and (iii) comparing this with a std. value. Higher than std. values indicate APC resistance. Resistance is recognised by a low anti-coagulant response to exogenous APC not related to protein S deficiency or defective factor VIII/VIIIa, and has a low anti-coagulant response to exogenous APG in the absence of APC Ig inhibitors.

USE - Used for detecting APC resistance e.g. in screening and diagnosing thromboembolic diseases such as hereditary thrombopilia. Also for determining the risk of thrombosis in pregnancy, surgery and during contraceptive use.

Dwq.0/0

ABEQ EP 608235 B UPAB: 19960222

An in vitro method for diagnosing in a human, or for determining the risk for a human to acquire manifestation of, blood coagulation disorder designated APC resistance and recognized by an abnormally low anticoagulant response to exogenous activated Protein C (abbreviated APC) even in presence of normal levels of functional Protein S, a Factor VIIIa, which is normally degraded by APC, and absence of lupus anticoagulants, said method comprising determining for a plasma sample comprising coagulation factors and derived from a human, the anticoagulant activity of exogenous APC by measuring the substrate conversion rate obtained for a blood coagulation enzyme, the activity of which is influenced by APC, by the following steps: (i) incubating said plasma sample with (1) exogenous APC, or exogenous Protein C together with current exogenous reagents to transform the exogenous Protein C to APC; (2) an exogenous Reagent (I), which at least partially activates the blood coagulation system of said sample and is selected in a manner known per se to cause activation of a coagulation factor used for the measurement in step (ii); (3) components such as phospholipid(s) and Ca** salt, that are necessary to efficient reaction of the activated coaquiation factors introduced by step (i)(2); and if desired, (4) an exogenous substrate for an enzyme, the activity of said enzyme being influenced by APC; (ii) directly measuring said substrate conversion rate obtained in (i), and (iii) comparing the conversion rate measured in step (ii) with a standard value obtained from samples from normal individuals, which samples have been subjected to steps (i) and (ii) under essentially the same conditions as the plasma sample from said human; in which method a substrate conversion rate obtained for a plasma sample in step (ii), that is higher than the standard value indicates that said human suffers from or runs the risk of acquiring manifestation of said disorder. Dwg.0/0

7

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L62 ANSWER 38 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     1992-066428 [09]
AN
                        WPIX
DNN N1992-049884
                        DNC C1992-030418
     Immunoassays using soluble fibrin monomer - comprising
TT
     fibrin-like material, avoiding solubility maintaining reagent or
     polymerisation inhibitor, for plasmin activator
     determn..
DC
     B04 S03
     KUDRYK, B J; PROCYK, R
IN
     (NYBL-N) NEW YORK BLOOD CENTER INC; (NYBL-N) NEW YORK BLOOD CEN; (PROC-I)
PΑ
     PROCYK R; (NYBL-N) NY BLOOD CENT INC; (NYBL-N) NEW YORK BLOOD CENT INC
CYC
PΙ
     EP 472205
                     A 19920226 (199209) *
         R: AT BE CH DE ES FR GB GR IT LI LU NL SE
     AU 9182628
                    A 19920227 (199218)
                     A 19920224 (199220)
     CA 2049710
                                                      G01N033-573
     JP 04233458
                     A 19920821 (199242)
                                                13
                                                      G01N033-53
                                                                      <--
     AU 644205
                     B 19931202 (199404)
                                                      G01N033-68
                                                                      <--
     EP 472205
                     B1 19970108 (199707)
                                          EN
                                                18
                                                      G01N033-86
                                                                      <--
         R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE
                   E 19970220 (199713)
     DE 69124030
                                                      G01N033-86
                                                                      <--
                     A 20000613 (200035)
     US 6074837
                                                      C12Q001-56
                                                                     <--
     KR 171609
                     B1 19990501 (200051)
                                                      G01N033-50
                                                                      < - -
ADT
    EP 472205 A EP 1991-114134 19910823; CA 2049710 A CA 1991-2049710
     19910822; JP 04233458 A JP 1991-208131 19910820; AU 644205 B AU 1991-82628
     19910820; EP 472205 B1 EP 1991-114134 19910823; DE 69124030 E DE
     1991-624030 19910823, EP 1991-114134 19910823; US 6074837 A Cont of US
     1990-572189 19900823, Cont of US 1992-946826 19920917, Cont of US
     1994-308482 19940919, US 1995-468460 19950606; KR 171609 B1 KR 1991-14643
     19910823
    AU 644205 B Previous Publ. AU 9182628; DE 69124030 E Based on EP 472205
FDT
PRAI US 1990-572189
                          19900823; US 1992-946826
                                                         19920917;
     US 1994-308482
                          19940919; US 1995-468460
                                                         19950606
REP
     2.Jnl.Ref; EP 94720; WO 8605814; WO 8900005
IC
     ICM C12Q001-56; G01N033-50; G01N033-53;
          G01N033-573; G01N033-68; G01N033-86
         C12P021-02; C12P021-06; C12Q001-37; G01N003-68
AB
           472205 A UPAB: 19931006
     Reagent is a fibrinolytic material having a solubility and stability
     similar to fibrinogen in that it remains soluble and stable at
     physiological conditions at a concentration used in the assay in the absence of
     fibrin polymerisation inhibitors or reagents for
     maintaining solubility.
          USE/ADVANTAGE - Quantitative determn. of soluble fibrin
     monomers, plasmin activator inhibitor activity,
     tissue-plasminogen activator in plasma and
     immunoassays. Difficulty of solubilising fibrin under
     physiological conditions, useful for form of fibrin, or a
     substance with the properties of fibrin, that does not require
     special conditions for maintaining solubility especially in biochemical and
     immunological assays that require fibrin monomer or soluble
     fibrin.
     1/6
FS
     CPI EPI
FA
     AB; GI
MC
     CPI: B04-B02C3; B04-B04D; B11-C07A; B11-C08; B12-K04A
     EPI: S03-E14H1
ABEO EP
           472205 B UPAB: 19970212
     An assay method that requires a soluble fibrin or a
     soluble fibrin monomer reagent as one of the components of the
     assay, wherein the reagent is a fibrin-like material having a
     solubility and stability similar to fibrinogen in that it remains soluble
```

and stable at physiological conditions at a concentration employed in the assay and substantially identical to that produced by a process comprising the following steps: (1) partially reducing fibrinogen with a low amount of reducing agent, at slightly elevated temperature, under non-denaturing conditions, in the absence of divalent cations, and for a time sufficient to allow almost complete cleavage of susceptible disulphide bonds of the fibrinogen to free thiol groups, then (2) blocking said thiol groups with a blocking reagent, then optionally (3) reacting the product of step (2) with a clotting enzyme in physiological buffers in the absence of divalent cations to release fibrinopeptides A and B, and thereafter (4) terminating the activity of said clotting enzyme. Dwg.0/6

AN

TТ

DC

IN PA

PΙ

REP

IC

AB

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ANSWER 39 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
L62
     1991-193318 [26]
                       WPIX
DNN
    N1991-147980
                       DNC C1991-083712
     Improved fibrinogen assay avoids repeated calibration curves - avoids
     repeated calibration curves since single calibration only is required to
     convert sensor signal to concentration.
     B04 J04 S03 S05
     CALLAHAN, J B; HOFFMAN, J F; SWOPE, C H; CALLAHAN, J
     (ALKU) AKZO NV; (ALKU) AKZO NOBEL NV
CYC
    21
     WO 9108460
                    A 19910613 (199126) *
        RW: AT BE CH DE DK ES FR GB GR IT LU NL SE
        W: AU CA FI JP KR US
                    A 19910626 (199139)
     AU 9168983
     FI 9202312
                    A 19920521 (199235)
                                                      G01N
     EP 502103
                    A1 19920909 (199237) EN
                                                      G01N021-00
        R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE
                  A 19921125 (199301)
     ZA 9009564
                                               13
                                                     G01N000-00
     JP 05503008
                    W 19930527 (199326)
                                                5
                                                     C12Q001-56
     US 5223437
                    A 19930629 (199327)
                                                3
                                                     G01N021-00
     AU 641459
                    B 19930923 (199345)
                                                      C12Q001-56
     EP 502103
                    A4 19930505 (199526)
ADT FI 9202312 A WO 1990-US6988 19901130, FI 1992-2312 19920521; EP 502103 A1
     WO 1990-US6988 19901130, EP 1991-900512 19901130; ZA 9009564 A ZA
     1990-9564 19901128; JP 05503008 W WO 1990-US6988 19901130, JP 1991-501113
     19901130; US 5223437 A Cont of US 1989-443948 19891201, US 1991-696569
     19910508; AU 641459 B AU 1991-68983 19901130; EP 502103 A4 EP 1991-900512
    EP 502103 A1 Based on WO 9108460; JP 05503008 W Based on WO 9108460; AU
FDT
     641459 B Previous Publ. AU 9168983, Based on WO 9108460
PRAI US 1989-443948
                         19891201
     1.Jnl.Ref; US 3658480; US 3833864; US 3861877; US 3989382; US 4659550; US
     4720787; EP 184242; EP 59277; US 3432268; US 3905769
         C12Q001-56; G01N000-00
         G01N021-59; G01N021-75; G01N021-77; G01N033-86
          9108460 A UPAB: 19950301
     A method for optically measuring the concentration of fibrinogen in a
     blood plasma sample comprises: (a) providing a sample of
     plasma containing fibrinogen in a container; (b) adding
     thrombin to the sample, and mixing; (c) measuring the initial
     optical transmittance Ti for the reaction mixture; (d) allowing the
     thrombin and fibrinogen in the mixture to react; (e) measuring a
     final optical transmittance Tf for the reaction mixture; (f)
     comparing Tf with Ti to compute a delta value; and (g) determining the
     concentration of fibrinogen based on the delta value.
          USE/ADVANTAGE - Prior art methods of determining fibrinogen
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concentration by clotting methods depend on repeated construction of calibration curves, involve considerable calculation and are time consuming. Also the quantity being measured is often instrument and time dependent. The present method

eliminates the need to establish a standard curve repeatedly, as it need be done only once. This is constructed so that it remains unchanged by variations, instrument, or sample. Once the correlation equation for R is established, it can be permanently stored in the computer software. @(12pp Dwg.No.0/0) 0/0 FS CPI EPI FA AB MC CPI: B04-B04D2; B04-B04D4; B11-C07B2; B11-C09; B12-K04A; J04-B01B EPI: S03-E04B1; S03-E14H1; S05-C01 ABEQ JP 05503008 W UPAB: 19931116 A method for optically measuring the concn. of fibrinogen in a blood plasma sample comprises: (a) providing a sample of plasma contg. fibrinogen in a container; (b) adding thrombin to the sample, and mixing; (c) measuring the initial optical transmittance Ti for the reaction mixt.; (d) allowing the thrombin and fibrinogen in the mixt. to react; (e) measuring a final optical transmittance Tf for the reaction mixt.; (f) comparing Tf with Ti to compute a delta value; and (g) detg. the concn. of fibrinogen based on the delta value. USE/ADVANTAGE - The method eliminates the need to establish a standard curve repeatedly, as it need be done only once. This is constructed so that it remains unchanged by variations, instrument, or sample, Once the correlation equation for R is established, it can be permanently stored in the computer software. ABEQ US 5223437 A UPAB: 19931116 Optically measuring a concn. of fibrinogen in a blood plasma sample comprises adding thrombin to a sample of plasma contg. fibrinogen in a container, mixing the thrombin with the sample, measuring an initial optical transmittance for the obtd. reaction mixt., allowing the thrombin and fibrinogen to react, measuring a final optical transmittance for the reaction mixt., comparing the final transmittance measurement to the initial transmittance measurement to compute a delta value and determining the concn. of fibrinogen based on the delta value. Pref., the thrombin is at a concn. of 100 N1H units and the plasma sample is diluted in a 1:10 ratio with Na barbital. ADVANTAGE - Method has improved efficiency. The effects of instrument and channel variation in measuring the changes in optical transmission, are eliminated. Dwg.0/3 L62 ANSWER 40 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN AN 1986-108161 [17] WPIX DNN N1986-079665 DNC C1986-046181 TI Monitoring coagulation of fibrinogen - avoiding early false positives due to noise. DC B04 J04 S03 IN LIPSCOMB, M S PA (INNO-N) INNOVATIVE MEDICAL SYSTEMS; (ORTH) ORTHO DIAGNOSTIC SYSTEMS INC CYC 16 PΙ EP 178910 A 19860423 (198617) * EN R: AT BE CH DE FR GB IT LI LU NL SE AU 8548728 A 19860424 (198624) A 19860616 (198630) A 19870616 (198729) A 19880119 (198805) JP 61128172 ES 8704637 US 4720787 A B CA 1236921 19880517 (198824)

19920108 (199203)

R: AT BE CH DE FR GB IT LI LU NL SE

DE 3585129 G 19920220 (199209)

EP 178910

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JP 06230011
                     A 19940819 (199438)
                                                11
                                                      G01N033-86
                                                                      <--
                     B2 19950906 (199540)
     JP 07082020
                                                9
                                                      G01N033-86
                                                                      <--
     JP 2500848
                    B2 19960529 (199626)
                                                12
                                                      G01N033-86
                                                                      <--
ADT JP 61128172 A JP 1985-229787 19851015; ES 8704637 A ES 1985-547898
     19851015; US 4720787 A US 1985-786925 19851011; JP 06230011 A Div ex JP
     1985-229787 19851015, JP 1993-342245 19851015; JP 07082020 B2 JP
     1985-229787 19851015; JP 2500848 B2 Div ex JP 1985-229787 19851015, JP
     1993-342245 19851015
FDT
    JP 07082020 B2 Based on JP 61128172; JP 2500848 B2 Previous Publ. JP
     06230011
PRAI GB 1984-26004
                          19841015
    5.Jnl.Ref; A3...8627; A3...8846; DE 2635081; DE 3230672; DE 3439344; EP
     107498; EP 109234; EP 117470; EP 128009; EP 131789; EP 77063; EP 80032; EP
     87686; EP 88540; FR 2408839; GB 2040292; No-SR.Pub; US 4279616; WO 8301198
IC
     C12Q001-56; G01N033-49
     ICM G01N033-86
     ICS C12Q001-56; G01N033-49
AB
     EP
           178910 A UPAB: 19970502
     The presence of a coagulation component (I) in a mixture of
     coagulation reagent and patient sample uses a sensor for
     determining increases in optical density and provides a coordinating
     signal. (a) The value of the signal proportional to the optical property
     of the mixture at various times during a predetermined interval
     after formation of the mixture is measured and stored; (b) the value of the
     signal at the end of the predetermined interval is determined; (c) the
     stored signal values are scanned starting with the last acquired signal to
     determine the time T1 when the measured signal is less than or
     equal to X times the determined end value; (d) the stored signal
     values are scanned starting with the last acquired signal to determine the
     time T2 when the measured signal is less than or equal to Y
     times the determined end value when X is greater than and less
     than 1; (e) a function relating the measured signal to time is
     produced by performing a curve fitting analysis of the measured signal
     values over a time period bounded by T1 and T2; and (f) the
     time at which the value of the signal is equal to Z times
     the determined end value is determined from the function where Z = 0-1
     exclusive whereby the (I) clotting detection time is
     determined.S ADVANTAGE - Method shows high sensitivity for
     detection of clotting times due to reduction of noise
     effects in the clotting time. It provides
     statistically valid data which may be used to set confidence levels on the
     final clotting time results, especially valuable in that it
     permits singlet sample testing as opposed to prior art required double
     sample testing. This reduces by half the amount of reagents and personnel
     required and increases throughput capacity.
     Dwq.0/0
FS
     CPI EPI
FA
     AB
MC
     CPI: B04-B04D2; B11-C08; B12-H04; B12-K04A; J04-B01;
          J04-C04
     EPI: S03-E14H1
ABEQ EP
           178910 B UPAB: 19930922
       Method for measuring blood clotting
     time by monitoring the prsence of a coagulation
     component in a mixture of coagulation reagent and patient sample
     and employing a sensor for determining increases in optical density and
     providing a signal proportional thereto, comprising: a) measuring and
     storing the value of the signal proportional to the optical property of
     the mixture at a plurality of times during a predetermined
     interval after formation of the mixture; b) determining the value (Vmax)
     of the signal at the end of the predetermined interval; c) scanning the
     stored signal values starting with the last acquired signal to determine
```

the time T1 when the measured signal is less than or equal to X

times the determined end value (Vmax); d) scanning the stored signal values starting with the last acquired signal to determine the time T2 when the measured signal is less than or equal to Y times the determined end value wherein l is above 1 which is above Y which is above X which is above O, and X and Y are determined experimentally; e) statistically calculating a function relating the measured signal to time by performing a curved fitting analysis of the measured signal values over a time period bounded by times T1 and T2; and f) determining the blood clotting time from said function, said blood clotting time being the time at which the value of the signal is equal to Z times the statistically determined end value wherein Z is between 1 and 0 and Z is determined empirically.

ABEQ US 4720787 A UPAB: 19930922

System for monitoring presence of a coagulation component in a mixt. of coagulation reagent and patient sample involves measuring component clotting detection time and employing a sensor to determine increase in optical density and providing a signal in coordination.

Values of the signal proportional to the optical property of the mixt. are measured and stored during a predetermined interval after mixt. formation. The last stored value of the signal is determined at the end of the predetermined interval, all stored signal values are scanned starting with the last stored signal value to determine a time T1 when the measured signal exceeds or equals a fraction X times the last stored signal value. The stored signal values starting with the last value are further scanned starting with the last stored signal value to determine a time T2 when the measured signal is less than or equal to a fraction Y times the last stored signal, where Y is less than 1 and X is less than Y. A function relating stored signal values to times is produced by curve fitting analysis of the stored signal for times between T1 and T2. From this function, the time at which the value of the signal equals Z times the last stored signal value is determined, where Z is 0-1 (exclusive). Hence component clotting detection time is determined.

USE - For fibrinogen determn. and in thrombin, partial thromboplastin and prothrombin coagulation tests.

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ANSWER 41 OF 41 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
L62
AN
     1985-237368 [39]
                        WPIX
DNN
    N1985-177516
                        DNC C1985-102812
TI
     Test composition for partial thromboplastin time
     determination - contains a platelet factor reagent and a
     hydrocarbyl sulphonic or sulphuric acid or salt.
DC
    B04 B05 S03
PA
     (ALKU) AKZO NV; (WARN) GOEDECKE AG; (WITT-I) WITT P
CYC
    13
                    A 19850919 (198539) *
PΙ
    DE 3407280
                                                16
                        19850925 (198539)
                     Α
        R: AT BE CH DE FR GB IT LI LU NL SE
     JP 60203200
                    A 19851014 (198547)
     US 4672030
                     Α
                        19870609 (198725)
                        19900425 (199017)
     EP 155565
                     В
        R: AT BE CH DE FR GB IT LI LU NL SE
    DE 3577349
                     G 19900531 (199023)
     JP 06004040
                     B2 19940119 (199406)
                                                      C12Q001-56
    DE 3407280 A DE 1984-3407280 19840228; EP 155565 A EP 1985-102281
ADT
     19850228; JP 60203200 A JP 1985-38636 19850227; US 4672030 A US
     1985-702284 19850215; JP 06004040 B2 JP 1985-38636 19850227
    JP 06004040 B2 Based on JP 60203200
PRAI DE 1984-3407280
                          19840228
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A3...8647; EP 107383; EP 49877; No-SR.Pub
     C12Q001-56; G01B033-86; G01N033-86
IC
     ICM C12Q001-56
     ICS G01B033-86; G01N033-86
AB
         3407280 A UPAB: 19970909
     A test composition for partial thromboplastin time
     (PTT) determination with a platelet factor reagent also contains
     at least one cpd. of formula (Ia) (R-O-SO3-X) or (Ib) (R-SO3-X) (where R
     is opt. substd. singly or multiply unsatd. branched or straight chain
     aliphatic or cycloaliphatic hydrocarbyl or an opt. substd. aromatic gp; X
     is H or a cation).
          USE/ADVANTAGE - The test composition can be used to carry out
     screening tests for endogenous coagulation systems, and in the
     monitoring of heparin therapy. The reconstituted reagent is stable at 37
     deg.C, room temperature and on storage in a fridge The reagent is more
sensitive
     towards heporin than previous reagents.
     Dwg.0/0
FS
     CPI EPI
FΑ
     AB
     CPI: B10-A09A; B10-A09B; B12-K04
MC
     EPI: S03-E14H9
           155565 B UPAB: 19930925
ABEQ EP
     Test kit for the PTT determination with a platelet factor
     reagent, characterised in that it additionally contains at least one
     compound of the formulae R-O-SO3X or R-SO3X in which R stands for an
     optionally substituted mono- or polyunsaturated, straight-chain or
     branched aliphatic or cycloaliphatic hydrocarbon radical or an optionally
     substituted aromatic radical, and X represents a hydrogen atom or a
     cation, is additionally added to the reagent in an amount such
     that 11 of the reconstituted solution contains 0.05 to 100 mmol thereof.
          4672030 A UPAB: 19930925
ABEQ US
     Test kit for determn. of partial thromboplastin time
     (PTT) contg. test reagents comprises a PTT reagent and a cpd. (I) of
     formula R-O-SO3X or R-SO3X (where R is mono- or poly-unsatd. opt. branched
     (cyclo) aliphatic hydrocarbon gp. or aromatic gp. and is opt. substd.; and
     X is H or a cation).
          Pref. 1 l. reconstituted test reagent soln. contains 0.05-100
     (0.2-20) mmol. (I). (I) may be dodecylbenzene sulphonic acid-Na salt or Na
     lauryl sulphate.
          USE/ADVANTAGE - In diagnostic tests for the endogenous
     coagulation system. The test kit offers improved sensitivity esp.
     to low mol. heparin fractions, and enables control of heparin therapy.
=> d his
     (FILE 'HOME' ENTERED AT 09:30:52 ON 07 JUL 2004)
                SET COST OFF
     FILE 'WPIX' ENTERED AT 09:31:08 ON 07 JUL 2004
              1 S (US20040053351 OR US6743596)/PN OR (WO2001-US32564 OR US2003-
L1
                E BIOMER/PA
            178 S E6-E17
L2
                E INMR/PACO
            641 S E3, E4
L3
                E AKZO NOBEL/PA
           2016 S E3-E29
1.4
                E ALKU/PACO
           4288 S E3-E5
L5
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E FISCHER T/AU

E BAGLIN T/AU

176 S E3,E10

L6

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L7
              5 S E3, E4
                E TEJIDOR L/AU
L8
              8 S E3, E4
                E G01N033-86/IC, ICM, ICS
L9
            857 S E3-E7
                E G01N033-86/ICA, ICI
L10
             23 S E3, E4
L11
              1 S E38
L12
            877 S L9-L11
             23 S L2-L8 AND L12
L13
            151 S L2-L8 AND (S03-E14H1 OR B14-F08 OR C14-F08 OR B12-H04 OR C12-
L14
            151 S L2-L8 AND (B04-B04D? OR C04-B04D?)/MC
L15
                E G01N033-49/IC, ICM, ICS
           2694 S E3-E5
L16
                E G01N033-49/ICA, ICI
L17
             94 S E3, E4
L18
              1 S E101
L19
           2775 S L16-L18
                E G01N033-50/IC, ICM, ICS
          14057 S E3-E5
L20
                E G01N033-50/ICA, ICI
            328 S E3, E4
L21
L22
              8 S E98
L23
          14354 S L20-L22
                E C12Q001-56/IC, ICM, ICS
L24
            608 S E3-E7
                E C12Q001-56/ICA,ICI
L25
             30 S E3
L26
            632 S L24, L25
L27
             69 S L2-L8 AND L19, L23, L26
            198 S L15, L27
L28
L29
              6 S L28 AND FIBRIN/BIX
L30
             23 S L28 AND ACTIVAT?/BIX
              8 S L28 AND THROMBIN/BIX
L31
L32
              4 S L28 AND PROTEIN C/BIX
              3 S L28 AND THROMBOMODULIN/BIX
L33
              4 S L28 AND TISSUE FACTOR/BIX
L34
L35
             40 S L1,L13,L29-L34
            163 S L28 NOT L35
L36
L37
            176 S L2-L8 AND (P813 OR P815 OR R611)/M0,M1,M2,M3,M4,M5,M6
             96 S L37 NOT L28, L36
L38
                SEL DN AN L35 5 6 8 11 13-15 17 25 27 36 40
L39
             12 S E1-E35
L40
             12 S L1,L39 AND L1-L39
L41
            185 S L12 AND L19, L23
L42
            325 S L12 AND L26
L43
             77 S L41 AND L42
             28 S L41 AND FIBRIN/BIX
L44
             37 S L41 AND THROMBIN/BIX
L45
             12 S L41 AND PROTEIN C/BIX
L46
              4 S L41 AND TISSUE FACTOR/BIX
L47
L48
            107 S L41 AND TIME/BIX
L49
             39 S L48 AND L44-L47
                SEL DN AN L49 2 8-10 13 17 25-27 29 31-33 35-38
L50
             22 S L49 NOT E36-E84
             31 S L40, L50
L51
L52
             53 S L43-L47 NOT L49-L51, L2-L8
                SEL DN AN L52 4 7 21 23 24 27 28 31 41 44
L53
             10 S E85-E114
             41 S L51, L53 AND L1-L53
L54
L55
             41 S L54 AND (?COAGUL? OR ?CLOT? OR ACTIVAT? OR INITIAT? OR THROMB
             41 S L54 AND (METHOD? OR PLASMA OR BLOOD OR SERUM OR CATION? OR H
L56
L57
             41 S L54-L56 AND (A61B OR C12Q OR G01N033)/IC, ICM, ICS, ICA, ICI
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L58	41	s	L57 AND	(B04-H19	OR	C04-H19	OR	B04-B04D?	OR	C04-B04D?	OR	B04-
L59	41	s	L54-L58	, L1								
L60	35	S	L59 AND	?THROMB?	/BI	X						
L61	8	S	L59 AND	(HEME? O	R H	AEM?)/BI	K					
L62	41	S	L59-L61									•

FILE 'WPIX' ENTERED AT 10:40:01 ON 07 JUL 2004

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